

KINETICS OF BIOLOGICAL NITROGEN REMOVAL
IN A SINGLE SLUDGE ACTIVATED SLUDGE PROCESS

by

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the requirements for the Degree of Master of
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DECLARATION BY CANDIDATE .

I, EPHRAIM SEHAYEK, hereby declare that this
thesis is my own work and that it has not been
submitted for a degree at another university.

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SYNOPSIS

The objective of this investigation was to develop experimental set-ups and operational procedures for accurate and reliable evaluation of the kinetic constants and sewage characteristics that influence nitrification and denitrification in the single sludge nitrification-denitrification activated sludge process. The kinetic constant that crucially affects nitrification has been identified as the maximum nitrification rate constant, μ_{nmT} ; the kinetic constants and sewage characteristic that crucially affect denitrification are the substrate utilization rate constant for slowly biodegradable COD in an aerobic environment, K_{mpT} , the ratio of substrate utilization rate constant for slowly biodegradable COD in an anoxic environment to that in an aerobic environment, $\eta = K'_{mpT}/K_{mpT}$ and the ratio of easily to total biodegradable COD in the influent, f_{ca} , respectively.

Series reactor systems under cyclic flow and load had previously been used extensively by Van Haandel, Ekama and Marais to determine the constants above by calibration. With this operational set up the main problem encountered was that the combination of series configuration *together* with cyclic flow and load led to an unstable response in a particular reactor so that it was not suitable for accurate evaluations by calibration. A refinement, using a series reactor system under constant flow and COD load and imposing *cyclic TKN load* indicated a practical problem: extremely accurate metering of the TKN pulse feeding rate and the recycle rates is required to obtain reliable estimates of the kinetic constants (by calibration against the general dynamic steady state model). This accuracy was not obtainable with the pumps available giving rise to imprecise and inaccurate evaluations.

To overcome the problems set out above a constant flow completely mixed *single* reactor system was proposed to be operated under a sequence of anoxic and aerobic periods. Application of the general dynamic steady state program to this system required extension of the program to accommodate the sequential anoxic aerobic operating condition. The approach proved singularly successful: the experimental data showed stable reproducibility and the general

dynamic steady state model simulated the response under constant flow and load very successfully to allow precise evaluation of the constants by calibration. This was found to be the case also, but to a lower degree of precision, for square wave flow and load conditions with anoxic aerobic sequences superimposed. One problem encountered was to simulate closely the response when (as was often the case) the experiment was performed on a day when the influent characteristics (TKN/COD ratio and COD) deviated appreciably from the mean values for the antecedent days. In this situation a satisfactory solution was to use the simulated data from the dynamic steady state program as initial input to commence the simulation for the particular day over which the experiment was performed with its particular influent flow and load. By this means the constants could still be evaluated with precision.

A useful development was that it was found possible to *manually* calculate μ_{nmT} from the experimental nitrate response of the system with good accuracy. The denitrification kinetic constants, K_{mpT} and η , could also be manually determined *provided* the operational conditions were such that the concentration of the stored COD mass, X_s , remained high.

Evaluation of the denitrification kinetic constants requires knowledge of the easily biodegradable COD fraction in the influent waste, f_{ca} . The magnitude of f_{ca} was determined by an independent experimental set up as described by Ekama and Marais.

The various experiments undertaken in this investigation indicate that for different waste influents μ_{nm20} could vary from 0,33 to 0,60 [d^{-1}], f_{ca} from 0,10 to 0,20 [$mg\ COD.mg\ COD^{-1}$] and K_{mp20} from 2,3 to 3,0 [$mg\ COD.mg\ VSS^{-1}.d^{-1}$], possibly due to presence of varying industrial fractions in the respective influents. Variation in K_{mp20} has not such a marked effect on the simulations except at high anoxic sludge mass fractions but μ_{nmT} and f_{ca} are the parameters that crucially affect nitrification and denitrification respectively - accurate experimental estimation of μ_{nmT} and f_{ca} , using the proposed influent waste as feed source is essential therefore for meaningful predictions of nitrification and denitrification behaviour in any anoxic-aerobic activated sludge process.

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LIST OF SYMBOLS

<u>SYMBOL</u>	<u>DESCRIPTION</u>
a	Mixed liquor recycle ratio from an aerobic to a predenitrification reactor.
b_h	Endogenous respiration rate constant for active sludge [d^{-1}].
b'_h	Death rate constant for active sludge [d^{-1}].
b_n	Death rate constant for nitrifiers [d^{-1}].
f	Inert residue in the endogenous respiration approach.
f'	Inert residue in the death regeneration approach.
f_{ca}	Ratio of easily to total biodegradable material in the influent.
f_{ma}	Maximum fraction of substrate that can be stored on the active sites. (Maximum ratio of stored material/active sludge).
f_n	Mass fraction of nitrogen in the organic sludge [$mg\ N.mg\ VSS^{-1}$].
f_{na}	Ratio of ammonia to total TKN in the influent.
f_{ns}	Ratio of TKN to organic material in the stored material [$mg\ N.mg\ COD^{-1}$].
f_{nu}	Unbiodegradable soluble TKN fraction in the influent.
f_{up}	Unbiodegradable particulate influent COD fraction [$mg\ VSS.mg\ COD^{-1}$].
f_{us}	Unbiodegradable soluble influent COD fraction.
f_{xl}	Primary (predenitrification) anoxic sludge mass fraction.
f_{xm}	Maximum anoxic sludge mass fraction to allow stable nitrification.
f_{xt}	Total unaerated sludge mass fraction.

q	Volumetric wastage flow rate [$\ell.d^{-1}$].
r	General symbol indicating the rate of a particular mechanism or the reaction rate of one of the variables in the activated sludge process [$mg.\ell^{-1}.d^{-1}$] or [$mg.\ell^{-1}.h^{-1}$]. index h refers to hydraulic rate index obs refers to observed rate index r refers to reaction rate.
s	Underflow recycle ratio.
t_a	Actual anoxic retention time [h].
t_f	Feeding period [h].
t_2-t_1	Time interval between any two moments [h].
Alk	Alkalinity expressed in ppm as $CaCO_3$.
C_c	Concentration of a variable C [$mg.\ell^{-1}$].
COD	Carbonaceous material concentration [$mg\ COD.\ell^{-1}$].
D	Dissolved oxygen (D.O) concentration [$mg\ O.\ell^{-1}$].
K	Empirical denitrification rate constant [$mg\ N.mg\ VSS^{-1}.h^{-1}$] or [$mg\ N.mg\ VSS^{-1}.d^{-1}$].
K_1	High rate (primary phase) predenitrification rate constant [$mg\ N.mg\ VSS^{-1}.h^{-1}$] or [$mg\ N.mg\ VSS^{-1}.d^{-1}$].
K_2	Low rate (secondary phase) predenitrification rate constant [$mg\ N.mg\ VSS^{-1}.h^{-1}$] or [$mg\ N.mg\ VSS^{-1}.d^{-1}$].
K_3	Low rate post-denitrification rate constant [$mg\ N.mg\ VSS^{-1}.h^{-1}$] or [$mg\ N.mg\ VSS^{-1}.d^{-1}$].
K_{mp}	Substrate utilization rate constant for slowly biodegradable COD in an aerobic environment [$mg\ COD.mg\ VSS^{-1}.d^{-1}$].
K'_{mp}	Substrate utilization rate constant for slowly biodegradable COD in an anoxic environment [$mg\ COD.mg\ VSS^{-1}.d^{-1}$].
K_{ms}	Substrate utilization rate constant for easily biodegradable COD [$mg\ COD.mg\ VSS^{-1}.d^{-1}$].
K_n	Half saturation value for nitrification [$mg\ NH_3-N.\ell^{-1}$].
K_r	Conversion rate constant for organic nitrogen [$\ell.mg\ VSS^{-1}.d^{-1}$].

K_{sp}	Half saturation value for slowly biodegradable COD (active site approach) $[\text{mg COD.mg VSS}^{-1}]$.
K_{ss}	Half saturation value for easily biodegradable COD $[\text{mg COD.l}^{-1}]$.
K_v	Adsorption rate constant for adsorption of particulate biodegradable material $[\text{l.mg VSS}^{-1}.\text{d}^{-1}]$.
$M(X)$	Mass of sludge present in an activated sludge system $[\text{mg VSS}]$. index a refers to active sludge index n refers to nitrifiers index v refers to volatile solids index x refers to any parameter.
N_a or NH_3	Ammonia concentration $[\text{mg NH}_3\text{-N.l}^{-1}]$.
N_{ae}	Effluent ammonia concentration $[\text{mg NH}_3\text{-N.l}^{-1}]$.
NON	Organic nitrogen concentration $[\text{mg NON-N.l}^{-1}]$.
N_n or NO_3	Nitrate concentration $[\text{mg NO}_3\text{-N.l}^{-1}]$
N_s	Heterotrophic nutrient nitrogen requirement $[\text{mg N.l}^{-1}]$.
O	General symbol denoting oxygen uptake rate in an aerobic reactor $[\text{mg O.l}^{-1}.\text{h}^{-1}]$. index c refers to oxygen uptake rate associated with oxidation of carbonaceous material. index n refers to oxygen uptake rate associated with oxidation of ammonia. index sbp refers to oxygen uptake rate associated with oxidation of slowly biodegradable COD. index sbs refers to oxygen uptake rate associated with oxidation of easily biodegradable COD. index t refers to total oxygen uptake rate.
P	COD equivalence of organic sludge = COD/VSS ratio $[\text{mg COD.mg VSS}^{-1}]$.
Q	General symbol indicating flow rate $[\text{l.d}^{-1}]$ index a refers to actual flow rate index b refers to base flow rate.

K_{sp}	Half saturation value for slowly biodegradable COD (active site approach) $[\text{mg COD.mg VSS}^{-1}]$.
K_{ss}	Half saturation value for easily biodegradable COD $[\text{mg COD.l}^{-1}]$.
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P	COD equivalence of organic sludge = COD/VSS ratio $[\text{mg COD.mg VSS}^{-1}]$.
Q	General symbol indicating flow rate $[\text{l.d}^{-1}]$ index a refers to actual flow rate index b refers to base flow rate.

R_h	Hydraulic (nominal, unless otherwise stated) retention time [d].
R_s	Sludge age [d].
R_{sm}	Minimum sludge age required for nitrification [d].
R_o	Nominal retention time during which the high rate (primary phase) predenitrification rate constant, K_{1T} is operative [d].
R_1	Total nominal retention time in the predenitrification reactor [d].
R_3	Nominal retention time in the post-denitrification reactor [d].
S	General symbol indicating COD substrate [$\text{mg COD} \cdot \ell^{-1}$]. index b refers to biodegradable substrate. index bp refers to slowly biodegradable substrate. index bs refers to easily biodegradable substrate index t refers to total carbonaceous material index u refers to unbiodegradable soluble COD.
S_f	Factor of safety introduced to ensure efficient nitrification.
TKN	Total kjeldahl nitrogen concentration [$\text{mg N} \cdot \ell^{-1}$].
V	Total reactor volume [ℓ].
VSS	Volatile suspended solids concentration [$\text{mg VSS} \cdot \ell^{-1}$].
X	General symbol indicating sludge concentration [$\text{mg VSS} \cdot \ell^{-1}$]. index a refers to active sludge index e refers to endogenous residue index i refers to inert material index n refers to nitrifiers index s refers to stored material index v refers to volatile solids.
Y_h	Yield coefficient of active heterotrophic sludge [$\text{mg VSS} \cdot \text{mg COD}^{-1}$].
Y_n	Yield coefficient of nitrifiers [$\text{mg VSS} \cdot \text{mg N}^{-1}$].
α	Proportionality constant between extent of high rate nitrate removal and influent biodegradable COD concentration = $f_{ca}(1 - PY_h)/2,86$ [$\text{mg NO}_3\text{-N} \cdot \text{mg COD}^{-1}$].

Δ	General symbol denoting a net change.
ΔN	Total nitrogen removal [$\text{mg NO}_3\text{-N.l}^{-1}$].
ΔN_1 or ΔN_{sbs}	Nitrogen removed due to utilization of easily biodegradable COD in a predenitrification reactor [$\text{mg NO}_3\text{-N.l}^{-1}$].
ΔN_2 or ΔN_{sbp}	Nitrogen removed due to utilization of slowly biodegradable COD in a predenitrification reactor [$\text{mg NO}_3\text{-N.l}^{-1}$].
ΔN_3	Nitrogen removed due to utilization of particulate COD in a post-denitrification reactor [$\text{mg NO}_3\text{-N.l}^{-1}$].
η	Ratio of substrate utilization constant for slowly biodegradable COD in an anoxic environment to that in an aerobic environment = $K'_{\text{mp}}/K_{\text{mp}}$.
μ_n	Specific growth rate of nitrifiers [d^{-1}].
μ_{nm}	Maximum specific growth rate of nitrifiers [d^{-1}].
μ'_{nm}	Effective maximum specific growth rate of nitrifiers [d^{-1}].

Suffixes to Subscripts

T	refers to temperature [$^{\circ}\text{C}$].
i	refers to influent.
p	refers to peak value.

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CHAPTER 1

INTRODUCTION

Three nitrification-denitrification activated sludge systems have been developed; the three sludge, two sludge and single sludge system, respectively. In the three sludge system the respective carbonaceous energy removal, nitrification and denitrification phases are achieved by having three separate reactor systems each with a settler (Fig.1a). Denitrification is achieved by adding an external energy source typically methanol, to the third reactor, this reactor being maintained in an anoxic state. In the two sludge system the energy removal and nitrification phases are consolidated to take place in *one* reactor and that of denitrification in a second, each with a separate settler (Fig.1b). In the single sludge process aerobic energy removal, nitrification and denitrification are mediated by the same sludge mass in a series reactor system having aerated and unaerated reactors (Fig.1c), or in a single reactor which is sequentially made aerobic and anoxic. The energy source for denitrification in the single sludge process is the organic material in the influent.

The three and two sludge systems have not been applied extensively in practice even though theoretically they are competent systems, the main reason against their use being the high cost of the energy source (usually methanol) for denitrification. As a consequence the single sludge system has received extensive attention as an alternative to the three and two sludge systems.

Modelling of the *three* sludge system has been accomplished relatively easily because the carbonaceous energy removal, nitrification and denitrification respectively, take place in separate reactors. Modelling of the *two* sludge system, where energy removal and nitrification occur in the first reactor, also has not presented any serious difficulties, the reason for this being that the energy removal and nitrification reactions are mediated by two entirely different groups of organisms - the autotrophic nitrifiers and heterotrophic energy removers - their respective kinetic behaviour in the same reactor can be modelled virtually independently of each

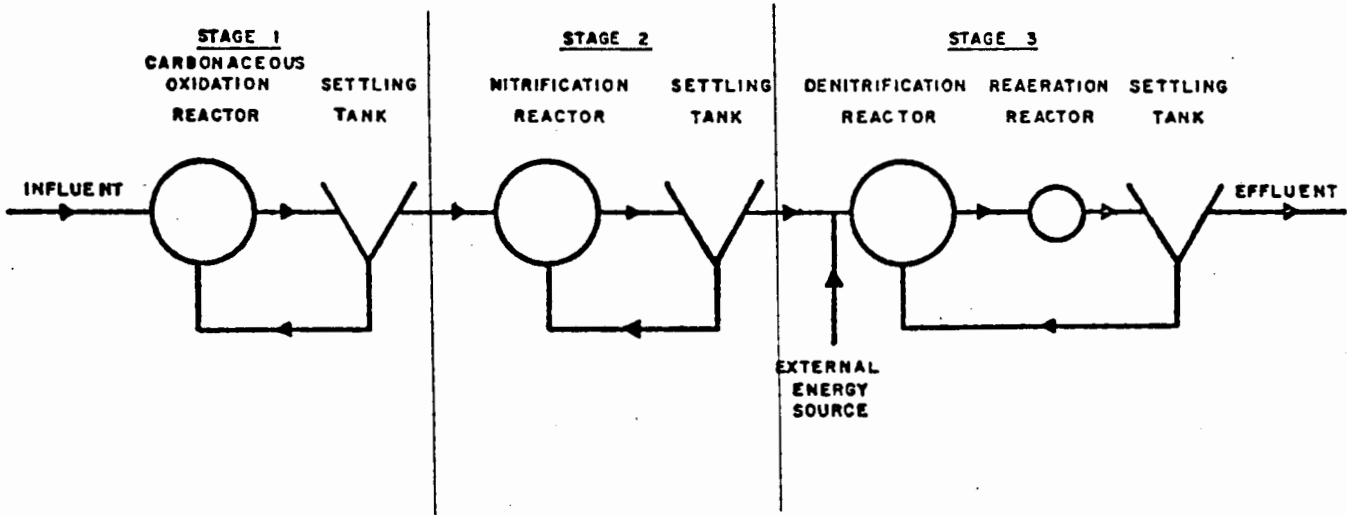


Fig.1a: Three sludge systems for denitrification with an external energy source.

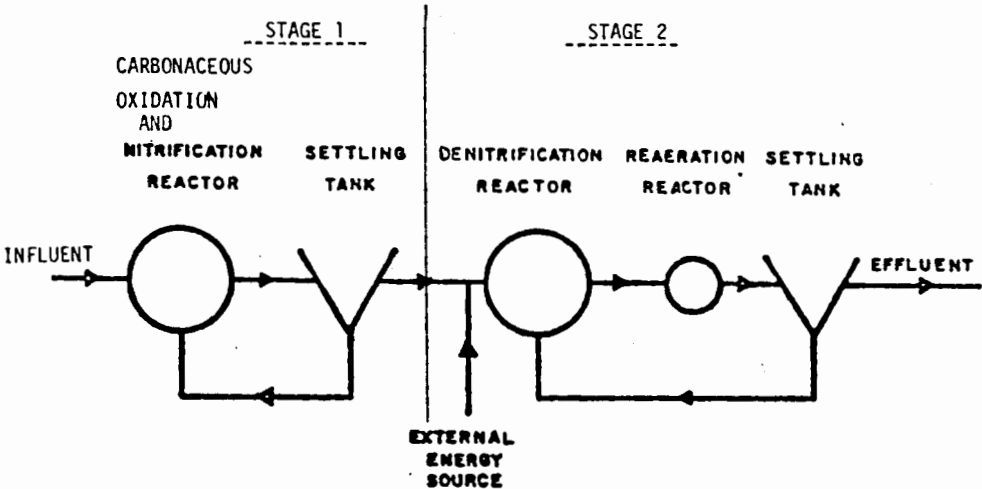


Fig.1b: Two sludge systems for denitrification with an external energy source.

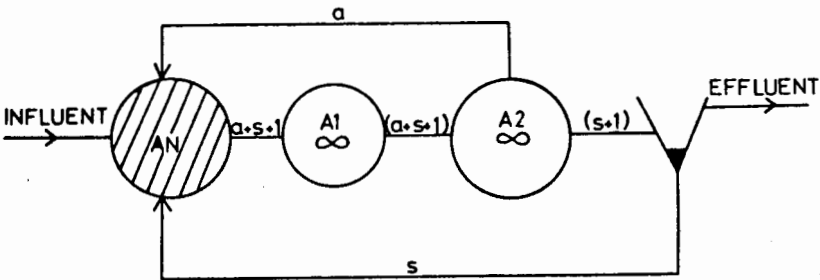


Fig.1c: Single sludge predenitrification series reactor system.

other; the two reactions are linked only insofar as the reactor and the process parameters (sludge age, temperature) are common to both, and that ammonia is abstracted from the wastewater for growth by both groups of organisms.

The denitrification phase in both the three and two sludge systems is entirely divorced from the preceding energy removal and nitrification phases. In addition, as the energy source for denitrification is very specific (say methanol), the organism type that develops in the denitrification reactor is also very specific to the energy source (as an electron donor) and nitrate (as an electron acceptor).

In contrast to the two and three sludge systems, modelling of the single sludge process requires description of a complex system of interacting reactions: energy removal for heterotrophic growth takes place from the same substrate source under sequential aerobic and anoxic (denitrification) phases, by the *same* organism mass; nitrification is affected by anoxic conditions in that growth of nitrifiers can take place only in an aerobic environment, but endogenous respiration takes place under both aerobic and anoxic conditions, and so on. Despite the complexity of the behaviour of the single sludge nitrification-denitrification process, because denitrification, like aerobic energy removal, is mediated by heterotrophic facultative organisms, Van Haandel, Ekama and Marais (1981) found that response of the autotrophs in an anoxic state may be described by the general model for aerobic systems developed by Dold, Ekama and Marais (1980), if it is assumed that only about 40% of the heterotrophs can utilize nitrate instead of oxygen as an electron acceptor. With this assumption they obtained very good correlation between the theoretically predicted and observed response at 20°C.

Accepting the basic model of Van Haandel *et al*, accurate description of the processes depends, *inter alia*, on the values assigned to the kinetic constants and sewage characteristics. These are determined by calibration of the model against response data. Because of the large number of parameters involved in the general model, it is possible to simulate satisfactorily the experimentally

observed data using different sets of constants, the reason for this being that some of the constants tend to act in a compensatory fashion with others. Assignment of precise values to the constants therefore requires *inter alia* experimental set ups that have good discriminative power for the particular constants to be evaluated. To obtain this discrimination, generally it is necessary to induce transient states in the system. Up to the present, this has been obtained by setting up series systems having anoxic and aerobic reactors operated under cyclic (usually square wave) flow and load conditions. Intrinsic to this system are the recycles to supply the mixed liquor and nitrate to the appropriate reactors. With this operational set up it has been found that the combination of series configuration, cyclic flow and load and recycles causes unstable response in a particular reactor and in consequence this approach has not been as successful as originally hoped in obtaining precise and accurate evaluations of constants by calibration.

The lack of precision and accuracy has provided the incentive to seek alternative set-ups and operational procedures to achieve estimates of greater reliability. For example, if the series configuration is selected such that a very short retention time is present in a particular reactor, so short that the reaction of interest is not complete, a transient type situation will be induced and the need for a cyclic flow and load might be eliminated and this cause for instability removed. Alternatively a *single* reactor might be operated under a time sequence of aerobic and anoxic conditions under constant or cyclic flow and load conditions. In such a system transients will be induced every time the operation is changed from aerobic to anoxic. A single reactor system also will not be affected by recycles as the recycle, in effect, is intra reactor.

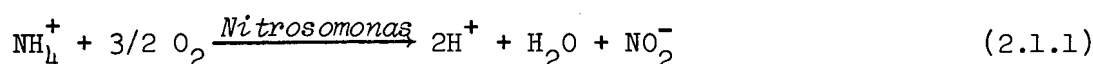
This report describes the research conducted into the development of different experimental set ups and operational procedures in the search for accurate and reliable evaluations of the kinetic constants and the sewage characteristics that influence nitrification and denitrification in single sludge systems.

CHAPTER 2

LITERATURE REVIEW

2.1 Autotrophic Nitrification2.1.1 Stoichiometry of nitrification

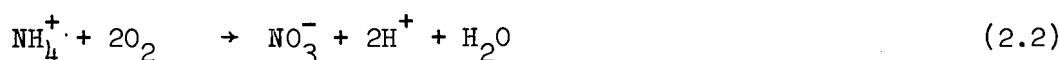
In biological nitrification oxidation of ammonia to nitrate is mediated by autotrophic bacteria and takes place in two steps. The first oxidation step, which yields nitrite, is mediated by autotroph *Nitrosomonas* (sp). Stoichiometrically:



and the second oxidation step, which yields nitrate, is mediated by autotroph *Nitrobacter* (sp). Again stoichiometrically:



The first step (Eq. 2.1.1) is the rate limiting one; nitrite is converted to nitrate almost as soon as it is formed. It is for this reason that most biologically nitrified ammonia is in the nitrate form. From a kinetic point of view the rate limiting step mediated by *Nitrosomonas*, therefore, becomes the step of importance. Hence the equivalent stoichiometric equation for conversion of ammonia to nitrate is written:



Considering the *Nitrosomonas*, Haug and McCarty (1971) accepted an empirical formula for the protoplasm of *Nitrosomonas* as $\text{C}_5\text{H}_7\text{NO}_2$. From reported data they selected a yield coefficient $Y_n = 0,15$ (mg VSS.mg N^{-1}) for autotrophic sludge production and apparently, using this yield coefficient it was possible to develop* the following

* Haug *et al* (1971) do not set out exactly how Eq.(2.3) was developed. However, the production of 1 mole $\text{C}_5\text{H}_7\text{NO}_2$ (113 g) from 55 moles NH_4^+ (770 g-N) corresponds to a yield coefficient $Y_n = 113/770 = 0,15$ (mg *Nitrosomonas*.mg N^{-1})

Nitrification of one mole of ammonia to nitrate produces 2 moles of H^+ ($= 2g H^+$) or equivalently consumes $2.50 = 100 g$ of alkalinity expressed as $CaCO_3$ (Loewenthal and Marais, 1976) i.e.

$$14 g N \equiv 2 g H^+ \equiv 100 g CaCO_3$$

or

$$1 mg N \equiv 7.14 mg CaCO_3$$

i.e. for each 1 mg of NH_4^+-N nitrified there is a consumption of 7.14 mg alkalinity expressed as $CaCO_3$.

2.1.3 Oxygen requirements of nitrification

Equation (2.2) describes the stoichiometric oxygen requirements for nitrification and states that nitrification of 1 mole of ammonia ($= 14 g$ expressed as NH_4^+-N) to nitrate requires 2 moles of oxygen ($= 64 g O$), i.e. the nitrification of 1 mg NH_4^+-N requires $64/14 = 4.57 mg O$.

2.2 Nitrification Kinetics

2.2.1 Nitrification kinetics in aerobic systems

Investigations of (Downing, Painter and Knowles, 1964) indicate that the relationship by Monod (1950) satisfactorily describes the rate of nitrification, i.e.

$$\mu_{nT} = \mu_{nmT} N_a / (N_a + K_{nT}) \quad (2.4)$$

μ_{nT} = Specific growth rate of nitrifiers [d^{-1}]

μ_{nmT} = Maximum specific growth rate of nitrifiers [d^{-1}]

K_{nT} = Monod half saturation coefficient [$mg NH_3-N.l^{-1}$]

N_a = Ammonia (substrate) concentration [$mg NH_3-N.l^{-1}$]

The index 'T' refers to temperature [$^{\circ}C$]

The basic equation for the rate of synthesis is given by:

$$dX_n/dt = \mu_{nT} X_n = (\mu_{nmT} N_a / (N_a + K_{nT})) X_n \quad (2.5)$$

X_n = Concentration of nitrifiers [$\text{mg } X_n \cdot \text{l}^{-1}$]

Now the mass of ammonia removed in relation to the mass of nitrifier synthesised is given by

$$\Delta X_n = Y_n \Delta N_a \quad (2.6a)$$

where Y_n = Yield coefficient for nitrifiers [$\text{mg VSS} \cdot \text{mg NH}_3\text{-N}^{-1}$]

Hence the rates can be expressed by

$$dX_n/dt = Y_n (dN_a/dt) \quad (2.6b)$$

Substituting for dX_n/dt from Eq. (2.5) yields

$$\frac{dN_a}{dt} = -\left(\frac{\mu_{nmT}}{Y_n}\right) \cdot \frac{N_a}{N_a + K_{nT}} \cdot X_n \quad (2.7)$$

The death rate of nitrifiers is given by

$$dX_n/dt = -b_{nT} X_n$$

b_{nT} = Death rate constant for nitrifiers [d^{-1}]

2.2.2 Nitrification process kinetics

The process response depends on the hydraulic regime (plug flow or completely mixed) together with the basic kinetic reactions. Assuming nitrification in a completely mixed reactor under constant flow and load conditions (depicted by Fig. 2.1), a mass balance on the accumulation of nitrifier mass, $M(\Delta X_n)$ is given by

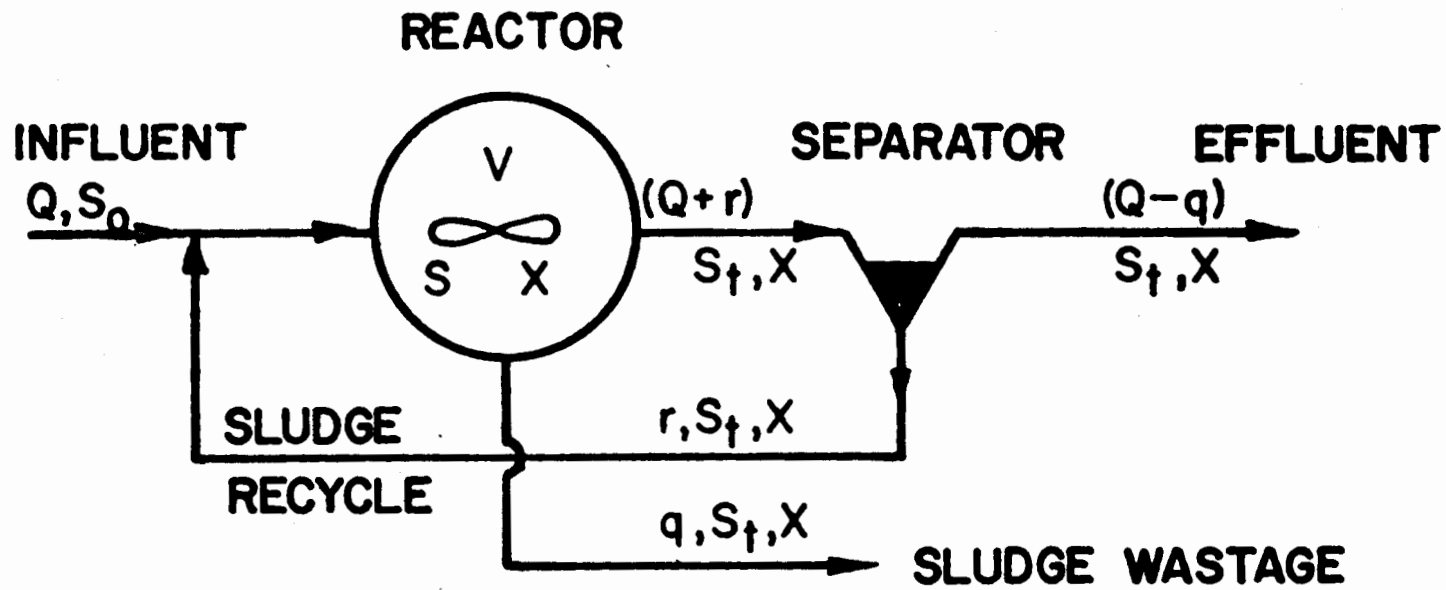
$$M(\Delta X_n) = V \cdot \Delta X_n = [\mu_{nmT} \cdot N_a / (N_a + K_{nT})] \cdot X_n \cdot V \cdot \Delta t - b_{nT} \cdot X_n \cdot V \cdot \Delta t - X_n \cdot q \Delta t$$

where

V = Process volume [l]

q = Volumetric withdrawal rate of sludge from the reactor [$\text{l} \cdot \text{d}^{-1}$]

Dividing through by $V \cdot \Delta t$



COMPLETELY MIXED ACTIVATED SLUDGE PROCESS WITH RECYCLING

Fig.2.1: A diagrammatic representation of the completely mixed activated sludge process with hydraulic control of sludge age and sludge recycle.

$$\therefore \Delta X_n / \Delta t = [\mu_{nmT} N_a / (N_a + K_{nT})] \cdot X_n - b_{nT} X_n - X_n q / V \quad (2.8a)$$

$$\text{Now, } \frac{V X_n}{q X_v} = \frac{\text{mass of sludge in the process per day}}{\text{mass of sludge wasted per day}} = \frac{V}{q}$$

$V/q = R_s$ defines the sludge age, R_s [d].

Under constant flow and load conditions eventually a steady state will be achieved when

$$dX_n / dt = 0$$

Equating Eq. (2.8a) to zero for steady state and substituting $1/R_s$ for q/V gives

$$[\mu_{nmT} N_a / (N_a + K_{nT})] X_n - b_{nT} X_n - X_n / R_s = dX_n / dt = 0 \quad (2.8b)$$

From Eq. (2.8b) by solving for N_a , the ammonia concentration in a single completely mixed aerobic reactor under constant flow and load conditions is determined i.e.

$$N_{ae} = \frac{K_{nT}(b_{nT} + 1/R_s)}{[\mu_{nmT} - (b_{nT} + 1/R_s)]} \quad (2.8)$$

where

$$N_{ae} = \text{Effluent ammonia concentration [mg NH}_3\text{-N.l}^{-1}\text{]}$$

Equation (2.8) provides a basis for estimation of the minimum aerobic sludge age required for nitrification, R_{sm} , (i.e. the sludge age below which nitrification ceases). This will occur when the effluent ammonia concentration (N_{ae}) equals the influent ammonia concentration (N_{ai}), i.e. when $N_a = N_{ai}$. Substituting $N_a = N_{ai}$ and noting that $K_{nT} \approx 1.0$ [mg N.l⁻¹] and $N_{ai} > 20$ [mg N.l⁻¹] i.e. $N_{ai} \gg K_{nT}$, in which event $K_{nT}/N_{ai} \approx 0$, Eq. (2.8) reduces to $[\mu_{nmT} - (b_{nT} + 1/R_s)] = 0$ or

$$R_{sm} = 1/(\mu_{nmT} - b_{nT}) \quad (2.8c)$$

To illustrate the effect of sludge age on N_{ae} a plot of N_{ae} vs. R_s is shown in Fig 2.2 a with $b_{n20} = 0,04 [d^{-1}]$, $K_{n20} = 1,0 [mg N.l^{-1}]$ for different values of μ_{nmT} . It may be noted that as R_s increases slightly higher than R_{sm} so the effluent ammonia concentration rapidly reduces to a small value. This behaviour has been repeatedly observed in practice - a plant usually either nitrifies almost completely or not at all.

2.2.2a Presence of an anoxic zone

If a fraction f_{xt} of the total sludge mass is unaerated (anoxic), a fraction $(1-f_{xt})$ is aerobic. The effect of the anoxic zone on nitrification can be explained by hypothesizing that endogenous respiration takes place throughout the aerated and unaerated zones whereas growth can take place only in the fraction of sludge mass that is aerated. A nitrifier mass balance gives Eq. (2.9) for the nitrification behaviour in a nitrification-denitrification process (Ekama, Van Haandel and Marais 1979) i.e.

$$N_{ae} = \frac{[K_{nT}(b_{nT} + 1/R_s)]}{[(1-f_{xt})\mu_{nmT} - (b_{nT} + 1/R_s)]} \quad (2.9)$$

Comparing Eq. (2.9) with the corresponding aerobic one [Eq. (2.8)], the presence of the anoxic sludge mass fraction, (f_{xt}) equivalently reduces the value of μ_{nmT} i.e.

$$\mu'_{nmT} = (1-f_{xt})\mu_{nmT}$$

$$\mu'_{nmT} = \text{Effective maximum specific growth rate } [d^{-1}]$$

If f_{xt} is fixed then the minimum sludge age for unaerated-aerobic systems can be calculated by substituting R_{sm} for R_s , $N_{ae} = N_{ai}$ and hence $K_{nT}/N_{ai} \approx 0$ in Eq. (2.9). Solving for R_{sm} , for any given f_{xt} , yields

$$R_{sm} = 1/(\mu'_{nmT} - b_{nT}) = 1/[\mu_{nmT}(1-f_{xt}) - b_{nT}] \quad (2.10a)$$

Conversely, if the sludge age is fixed, the maximum anoxic sludge mass fraction to allow nitrification (f_{xm}) can be calculated by substituting f_{xm} for f_{xt} , $N_{ae} = N_{ai}$ and hence $K_{nT}/N_{ai} \approx 0$ in Eq. (2.9). Solving for f_{xm} , for any given R_s , yields:

$$f_{xm} = 1 - (b_{nT} + 1/R_s)/\mu_{nmT} \quad (2.10b)$$

As stated previously, theoretically complete nitrification is achieved almost immediately R_s exceeds R_{sm} (see Fig. 2.2a) but clearly it is undesirable to operate at R_s close to R_{sm} as this may lead to unstable nitrification. It is necessary therefore to calculate a sludge age so much longer than the minimum in order that effluent ammonia concentration (N_{ae}) will always be low. In practice two factors influence the selection of the sludge age:

- (1) μ_{nmT} tends to vary around the mean value. Consequently the sludge age should at least be larger than that indicated by μ'_{nmT} in Eq. (2.10a).
- (2) Under cyclic load and flow conditions particularly the latter, the nitrification efficiency declines because during the high flow period the contact time with the nitrifiers is too short to allow complete nitrification.

In order to ensure that the mass of nitrifiers is sufficiently large to deal with the forementioned influences, the sludge age should be appreciably greater than the minimum. From simulations of nitrification systems, Ekama *et al* (1979) found that if R_s is at least 1.25 times R_{sm} , nitrification is virtually complete even under cyclic flow and load conditions. In the analysis above the same provision can be made by applying a factor of safety in the calculation of f_{xm} . This is accomplished by incorporating a factor of safety, S_f , in Eq. (2.10b) as follows:

$$f_{xm} \approx 1 - S_f(b_{nT} + 1/R_s)/\mu_{nmT} \quad (2.11a)$$

In Eq. (2.11a) the value of f_{xm} is subject to a restriction:

Arkley and Marais (1981) found that when f_{xm} exceeds 0,45 to 0,50, the settling characteristics of the sludge deteriorate to such an extent that the required size of settlers makes the plant uneconomical. Equation (2.11a), therefore, is subjected to the constraint that

$$f_{xm} < 0,50 \quad (2.11b)$$

This restriction has important implications in the design of nitrification-denitrification systems; these will be dealt with in Section 2.3.4.

2.2.3 Choice of nitrification constants

In order to calculate the kinetic behaviour of nitrification it is necessary to determine the magnitudes of all the kinetic constants affecting nitrification i.e. Y_n , b_{nT} , K_{nT} and μ_{nmT} . Some of these do not have a significant effect on nitrification rate so that great accuracy in determining their magnitude is not important. A comparative study of the effect of each of these parameters on the nitrification behavioural pattern will indicate which of the constants needs to be evaluated with great accuracy and which not.

Downing *et al* (1964), Knowles, Downing and Barret (1965), Lijklema (1973), Gujer (1977) and Ekama and Marais (1978) reported various values for Y_n , b_{nT} , K_{nT} and μ_{nmT} . The range of values for each of the constants, respectively, is given in Table 2.1.

Table 2.1 Ranges of values of nitrification constants

Constant	Range	Units
Y_n	0,05-0,15	mg VSS.mg N ⁻¹
b_{n20}	0,04-0,07	d ⁻¹
K_{n20}	0,2 -1,0	mg N.l ⁻¹
μ_{nmT}	0,33-0,65	d ⁻¹

In order to investigate the influence of each of the constants individually, a set of average values is accepted as a standard for comparison and the effects of changing each of the constants, keeping the others at their standard value, are investigated.

2.2.3(a) The significance of Y_n in nitrification

Two aspects determine the significance of Y_n in nitrification, i.e. practical considerations and theoretical considerations.

2.2.3(a.1) Practical significance of Y_n in nitrification

In a typical domestic waste the ratio of the influent TKN/COD $\approx 0,1$ [mg N.mg COD⁻¹]. By applying TKN/COD = 0,1 and other typical domestic sewage and operational characteristics (given in Table 2.2) to the various formulations used by Marais and Ekama (1976) for a completely mixed reactor under steady state conditions of constant flow and load, it is possible to compare the mass of nitrifier that is produced relative to the total mass of sludge that is generated for a given sludge age; this is depicted in Fig. 2.2. From Fig. 2.2 it is apparent that X_n never constitutes more than 2 per cent of the total sludge and therefore from a practical point of view the evaluation of X_n is of no consequence. Hence from Eq. (2.6a) it follows that the evaluation of Y_n is also of no consequence.

Table 2.2 Operational and sewage characteristics used in Fig. 2.2

<u>Operational Characteristics:</u>	
T	= 12°C
R _h	= 1,0[d]
<u>Sewage Characteristics:</u>	
S _{ti}	= 600 [mg COD.l ⁻¹]
f _{up}	= 0,09 [mg VSS.mg COD ⁻¹]
f _{us}	= 0,05 [mg COD.mg COD ⁻¹]
f _{nu}	= 0,0 [mg N.mg N ⁻¹]
f _{na}	= 0,75 [mg NH ₃ -N.mg N ⁻¹]

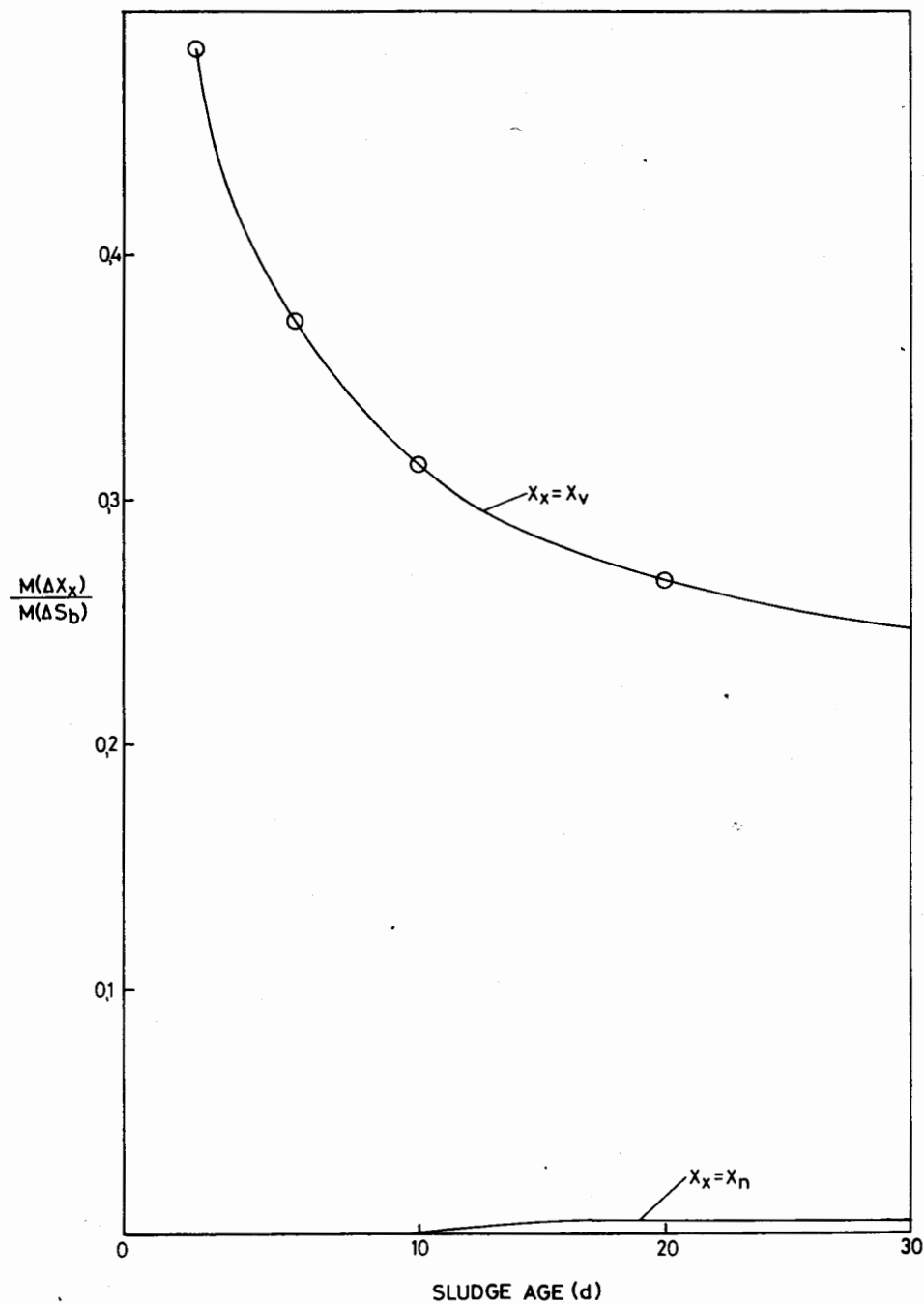


Fig.2.2: Comparison of the mass of nitrifier produced in a CMASP under constant flow and load relative to the mass of heterotrophs produced.

2.2.3(a.2) Theoretical significance of Y_n in nitrification

A theoretical evaluation of X_n is needed in order to determine the nitrification rate under changing conditions of flow and ammonia concentration. Associated with X_n is the value accorded to the maximum specific nitrification rate constant μ_{nmT} . The concentration of nitrifiers, X_n and μ_{nmT} are interrelated in the sense that $\mu_{nmT} \cdot X_n$ defines the rate of nitrification; if for the same rate X_n is estimated at a high figure, μ_{nmT} must necessarily be low and *vice versa* to keep $\mu_{nmT} \cdot X_n = \text{constant}$.

The evaluation of X_n requires knowledge of Y_n and b_{nT} . In the absence of data to fix these two parameters accurately it is necessary, for consistency in reporting μ_{nmT} values, to standardise the values of Y_n and b_{nT} . In all the models proposed by Marais and his group the following standard values were accepted: $Y_n = 0,10$ (mg VSS. mg N^{-1}), $b_{n20} = 0,04$ [d^{-1}]. All the μ_{nmT} values reported by them are relative to these two values.

Under constant flow and load conditions it is possible to describe the behavioural pattern of nitrification without the knowledge of X_n . This is achieved by eliminating X_n as done in the derivation of Eq. (2.8), i.e. *under conditions of constant flow and load* the value of Y_n becomes immaterial.

2.2.3(b) Choice of value for K_{nT}

By accepting a set of standard average values for the nitrification constants i.e. $\mu_{nm20} = 0,5[d^{-1}]$ and $b_{n20} = 0,04[d^{-1}]$, Fig. 2.2b compares the effect of K_{nT} on R_{sm} and on N_{ae} for $R_s > R_{sm}$. From Fig. 2.2b it is obvious that K_{nT} has hardly any effect on R_{sm} . Furthermore, for operational sludge ages of at least 1,25 times the minimum sludge age, the effluent ammonia concentration is low irrespective of the K_{nT} value. Hence knowledge of the exact value of K_{nT} is of no practical significance and an average value of $K_{n20} = 1,0$ [mg $N \cdot \ell^{-1}$] is accepted.

2.2.3(c) Choice of value for b_{nT}

By accepting a set of standard average values for the nitrification constants i.e. $\mu_{nm20} = 0,5[d^{-1}]$ and $K_{n20} = 1,0$ [mg $N \cdot \ell^{-1}$],

Fig 2.2c compares the effect of b_{nT} on R_{sm} and on N_{ae} for $R_s > R_{sm}$. From Fig 2.2c it can be seen that the effect of b_{nT} on R_{sm} is of no great significance and the effect of b_{nT} on the effluent ammonia concentration for $R_s > 1,25 R_{sm}$ is negligible. Hence knowledge of the exact value of b_{nT} is of no great importance and an average value of $b_{n20} = 0,04[d^{-1}]$ is accepted.

2.2.3(d) The significance of μ_{nmT} in nitrification

By accepting a set of standard average values for the nitrification constants i.e. $K_{n20} = 1,0[mg N.l^{-1}]$ and $b_{n20} = 0,04[d^{-1}]$, Fig. 2.2a compares the effect of μ_{nmT} on R_{sm} and on N_{ae} .

Fig. 2.2a clearly indicates that μ_{nmT} has a marked effect on the value of R_{sm} but at $R_s > 1,3 R_{sm}$ the effect on N_{ae} is insignificant. The knowledge of R_{sm} is crucial in designing a nitrification system hence an accurate estimation of μ_{nmT} is the key to a successful prediction of nitrification behaviour in an activated sludge plant.

The value of μ_{nmT} has been observed to vary with virtually every source of sewage and to vary between different batches of the same source.

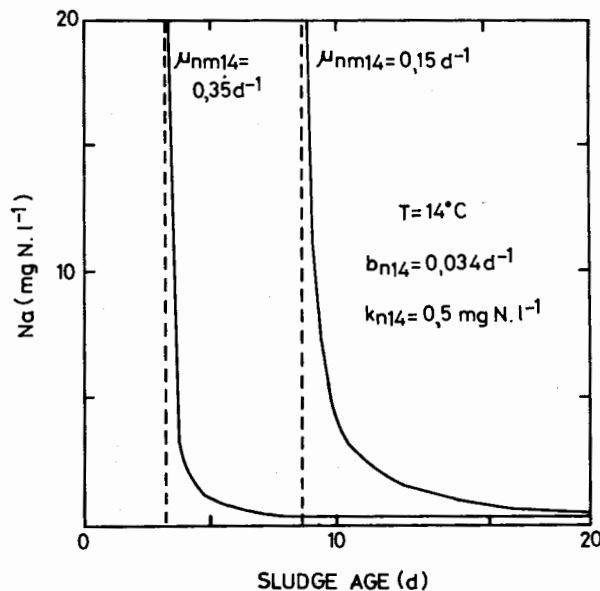


Fig.2.2a: Effect of μ_{nmT} on nitrification.

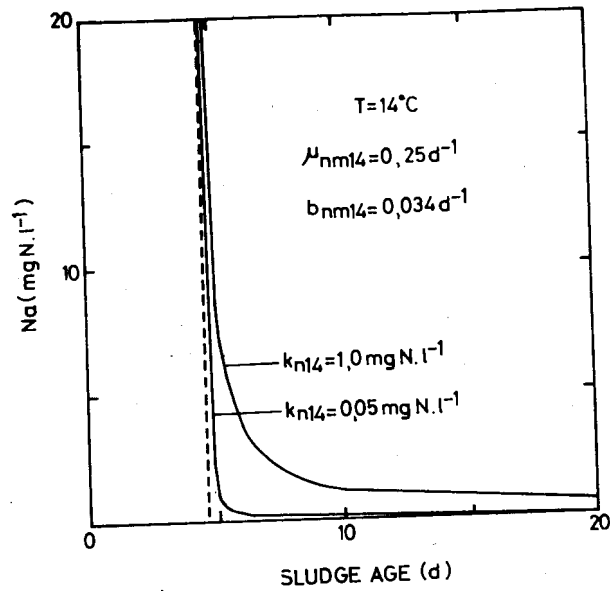


Fig.2.2b: Effect of K_{nT} on nitrification.

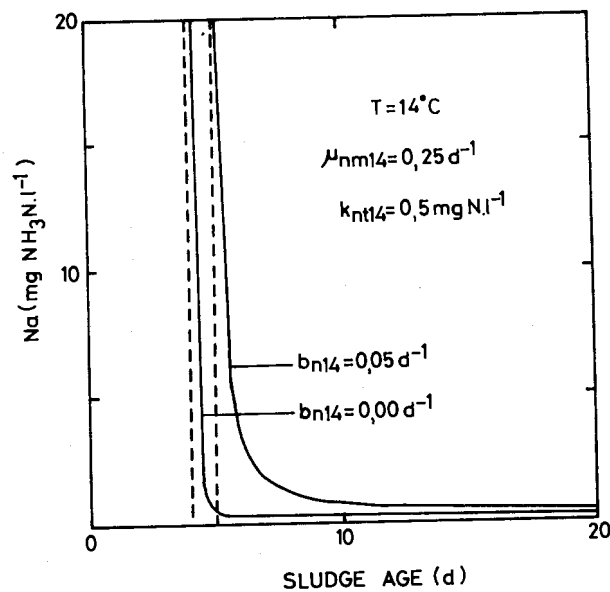


Fig.2.2c: Effect of b_{nT} on nitrification.

It therefore should be classified as a sewage characteristic rather than a kinetic constant. The reason for the marked variations in μ_{nmT} of a certain source is not completely clear. It does not appear to be a matter of toxicity but rather of inhibition and the degree of inhibition seems to be related to the fraction of industrial waste flow in the process. Due to the link between waste flow and μ_{nmT} , the value of μ_{nmT} should always be measured for the specific sewage to be dealt with.

2.2.4 Determination of μ_{nmT}

Theoretically the values of μ_{nmT} , K_{nT} and b_{nT} should be amenable to determination by calibration of Eq. (2.8) to a series of data on N_{ae} obtained over a range of sludge ages. The difficulty with this method is that in the region $R_s > R_{sm}$ the effluent ammonia is very insensitive to μ_{nmT} , b_{nT} and K_{nT} (see Figs. 2.2a, 2.2b and 2.2c). The only measurement that would be of use is to determine μ_{nmT} by operating at certain $R_s = R_{sm}$ but this is a region of instability and it requires patient and tedious experimentation to obtain reliable data.

To overcome the problems of the approach above, Ekama *et al* (1979) employed a series aerobic reactor system under cyclic loading conditions in order to create the transient conditions from which constants can be obtained by calibration. Again this approach has not been completely satisfactory. The uncertainty in this approach stems from the marked hydraulic effects of cyclic loading: slight error in the measurement of the flows results in marked variations in the respective recycles and consequently it is not possible to determine precise estimates for μ_{nmT} .

It is evident from the review above that there is a need for either improvement of the series reactor system or development of an alternative method that allows reliable determination of μ_{nmT} . These two alternatives form a substantial part of the research reported later in this thesis.

2.2.5 Incorporation of nitrification behaviour in aerobic-anoxic systems in a general model

When the general model of Dold, Ekama and Marais (1980) was adapted to aerobic-anoxic environment by Ekama *et al* (1979), the modifications concerning *nitrification* that needed to be made were:

- (1) There is no nitrification in an anoxic environment, i.e.

$$(\mu_{nmT})_{\text{anoxic}} = 0.$$
- (2) Nitrifiers can grow only in an aerobic environment (obligate aerobes) but undergo death (with its associated substrate release by lysis) in both aerobic and unaerated zones.
- (3) The model presupposes that an anoxic environment does not have a direct effect on the nitrification reaction kinetics of a subsequent aerobic environment - experimental evidence reported later in this work supports this hypothesis.

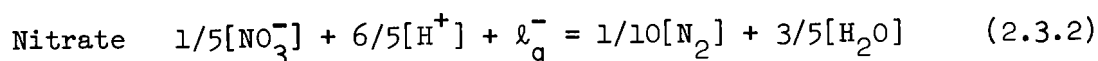
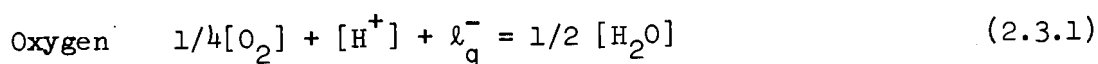
2.3 Denitrification

2.3.1 Stoichiometry of denitrification

Denitrification is a biologically mediated reaction known as dissimilative reduction of nitrate. It involves the reduction of nitrate or nitrite present in the wastewater to gaseous nitrogen, which escapes to the atmosphere.

The denitrification reaction is mediated by heterotrophic organisms; in the denitrification reaction these organisms obtain energy by the oxidation of organic material with nitrate and nitrite acting as electron acceptors. Usually the mass of nitrite present is negligible compared to that of nitrate (see Section 2.1.1) so that the reduction of nitrate only is of significance.

In terms of electron equivalents, the half reactions for the reduction of oxygen and nitrate can be compared:



Considering Eq. (2.3.1), the transfer of one electron equivalent (ℓ_q^-) is accomplished by the reduction of 1/4 mole of oxygen, i.e. 8g.

This transfer corresponds to the oxidation of 8g COD of the organic compounds (from the definition of the COD test below*).

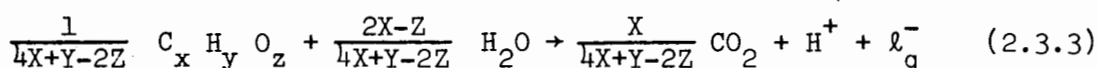
Considering Eq. (2.3.2), the transfer of one ℓ_q^- is accomplished by reduction of 1/5 mole of nitrate i.e. 62/5 g. If nitrate is measured in terms of the nitrogen atom i.e. in terms of ($\text{NO}_3\text{-N}$), then one ℓ_q^- is equivalent to the reduction of $(\frac{62}{5}) \cdot (\frac{14}{62}) = 2,80$ g ($\text{NO}_3\text{-N}$). Equating the electron equivalence of nitrate and oxygen from Eqs. (2.3.2 and 2.3.1) respectively, it follows that

$$2,80 \text{ g } (\text{NO}_3\text{-N}) \equiv 8 \text{ g O}$$

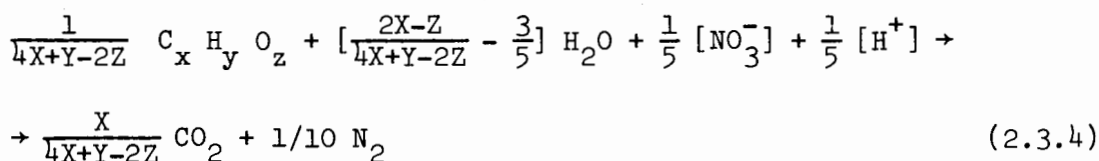
i.e.

$$1 \text{ mg } (\text{NO}_3\text{-N}) \equiv 2,86 \text{ mg O (or mg COD)}.$$

Equation (2.3.2) is a half reaction describing the reduction of nitrate. The corresponding half reaction describing the oxidation of COD (of a general formula $\text{C}_x \text{H}_y \text{O}_z$) is



Combining Eqs. (2.3.2 and 2.3.3), the following redox reaction can be written



Hence, during denitrification of 1 mole of NO_3^- , 1 mole of H^+ is consumed.

2.3.2(a) Oxygen recovery associated with denitrification

In Section 2.1.3 it was shown that for every 1 mg $\text{NH}_4^+\text{-N}$ of ammonia converted to nitrate, 4,57 mg O are consumed, and in Section 2.3.1 that for every 1 mg ($\text{NO}_3\text{-N}$) reduced to nitrogen gas, 2,86 mg O

*The COD test is a measure for the electron transfer potential of the organic matter [after Van Haandel and Marais (1981)].

are recovered equivalently. Thus, in the denitrification process of a single sludge system a fraction $(2,86/4,57) = 0,625$ of the oxygen required for nitrification can be recovered as equivalent oxygen.

2.3.2(b) Effect of denitrification on alkalinity

The conversion of one mole of ammonia to nitrate (nitrification), produces 2 moles of H^+ (Eq.2.2), i.e. the nitrification of 1 mg (NH_4^+-N) to nitrate consumes 7,14 mg alkalinity as $CaCO_3$ [Section 2.1.2]. Denitrification of one mole of nitrate to nitrogen gas consumes 1 mole of H^+ (Eq. 2.3.4). Hence for each mole of ammonia expressed as N i.e. (NH_4^+-N) ($\equiv 14$ g) converted to nitrogen gas, there is a nett production of $(2-1) = 1$ mole of H^+ (i.e. 1 g H^+) or, equivalently, a nett consumption of 50 g of alkalinity expressed as $CaCO_3$ [Loewenthal and Marais (1976)], i.e. per one mg of nitrified and subsequently denitrified nitrogen there is a nett decrease of $50/14 = 3,57$ mg of alkalinity expressed as $CaCO_3$.

2.3.3 Empirical approach to denitrification

Stern and Marais (1974) found that under *constant flow and load conditions* denitrification is approximately a zero order reaction with respect to nitrate concentration, and directly proportional to the active mass concentration. Hence under constant flow and load conditions the rate of denitrification could be described by an equation of the type

$$\Delta N/\Delta t = -K.X_a$$

Experimentally three rates were identified: in a predenitrification reactor, receiving the influent, two rates were apparent (specified by K_{1T} - and K_{2T} - rate constants) and in a post-denitrification reactor a single rate, K_{3T} - rate constant.

The rate constant K_{1T} was high, giving rise to a rapid rate of denitrification, but it terminated after a few minutes; the total removal of nitrogen associated with K_{1T} was observed to be proportional to the influent COD. The rate associated with K_{2T} continued for the balance of the time in the reactor, at about 1/7 the first rate.

The rate associated with K_{3T} was lower than that associated with K_{2T} (i.e. the value of K_{3T} was lower than that for K_{2T}) and continued for the total period of time in the post-denitrification reactor. The rates were temperature dependent but apparently independent of the sludge age for sludge ages between 10 and 22 days.

The retention time in the pre-denitrification reactor normally is long enough to allow completion of the rate associated with K_{1T} . Consequently it was possible to write the denitrification of a process having both pre- and post-anoxic reactors *under constant flow and load conditions* as follows:

$$\Delta N = \Delta N_1 + \Delta N_2 + \Delta N_3 = \alpha \cdot S_{ti} + K_{2T} \cdot (R_1 - R_0) \cdot X_a + K_{3T} R_3 X_a \quad (2.3.5)$$

where

ΔN = Total nitrogen removal [$\text{mg NO}_3\text{-N} \cdot \ell^{-1}$]

α = Predenitrification mass concentration constant during which K_{1T} is operative [$\text{mg NO}_3\text{-N} \cdot \text{mg COD}^{-1}$]

R_0 = Nominal retention time during which K_{1T} is operative [d]

R_1 = Total nominal retention time in the predenitrification reactor [d]

R_3 = Nominal retention time in the post-denitrification reactor [d]

S_{ti} = Total influent COD concentration [$\text{mg COD} \cdot \ell^{-1}$]

Note that the denitrification formulation expressed in Eq. (2.3.5) is applicable only under constant flow and load conditions, and is empirical, based on observed behaviour.

2.3.4 Application of the bi-substrate theory to denitrification

In 1978 (published in 1980) Dold, Ekama and Marais proposed that the biodegradable material in municipal waste flow is composed of an easily biodegradable fraction, f_{ca} , which is of a soluble nature (and is utilized at a high rate by the micro-organisms) and a slowly biodegradable fraction, $(1-f_{ca})$, which is of a particulate nature (and is utilized at a relatively low rate). Ekama, Van Haandel and Marais (1979) suggested that K_{1T} is associated with the utilization

2.2.3(a.2) Theoretical significance of Y_n in nitrification

A theoretical evaluation of X_n is needed in order to determine the nitrification rate under changing conditions of flow and ammonia concentration. Associated with X_n is the value accorded to the maximum specific nitrification rate constant μ_{nmT} . The concentration of nitrifiers, X_n and μ_{nmT} are interrelated in the sense that $\mu_{nmT} \cdot X_n$ defines the rate of nitrification; if for the same rate X_n is estimated at a high figure, μ_{nmT} must necessarily be low and *vice versa* to keep $\mu_{nmT} \cdot X_n = \text{constant}$.

The evaluation of X_n requires knowledge of Y_n and b_{nT} . In the absence of data to fix these two parameters accurately it is necessary, for consistency in reporting μ_{nmT} values, to standardise the values of Y_n and b_{nT} . In all the models proposed by Marais and his group the following standard values were accepted: $Y_n = 0,10$ (mg VSS. mg N^{-1}), $b_{n20} = 0,04$ [d^{-1}]. All the μ_{nmT} values reported by them are relative to these two values.

Under constant flow and load conditions it is possible to describe the behavioural pattern of nitrification without the knowledge of X_n . This is achieved by eliminating X_n as done in the derivation of Eq. (2.8), i.e. *under conditions of constant flow and load* the value of Y_n becomes immaterial.

2.2.3(b) Choice of value for K_{nT}

By accepting a set of standard average values for the nitrification constants i.e. $\mu_{nm20} = 0,5$ [d^{-1}] and $b_{n20} = 0,04$ [d^{-1}], Fig. 2.2b compares the effect of K_{nT} on R_{sm} and on N_{ae} for $R_s > R_{sm}$. From Fig. 2.2b it is obvious that K_{nT} has hardly any effect on R_{sm} . Furthermore, for operational sludge ages of at least 1,25 times the minimum sludge age, the effluent ammonia concentration is low irrespective of the K_{nT} value. Hence knowledge of the exact value of K_{nT} is of no practical significance and an average value of $K_{n20} = 1,0$ [mg $N \cdot l^{-1}$] is accepted.

2.2.3(c) Choice of value for b_{nT}

By accepting a set of standard average values for the nitrification constants i.e. $\mu_{nm20} = 0,5$ [d^{-1}] and $K_{n20} = 1,0$ [mg $N \cdot l^{-1}$],

Fig 2.2c compares the effect of b_{nT} on R_{sm} and on N_{ae} for $R_s > R_{sm}$. From Fig 2.2c it can be seen that the effect of b_{nT} on R_{sm} is of no great significance and the effect of b_{nT} on the effluent ammonia concentration for $R_s \geq 1,25 R_{sm}$ is negligible. Hence knowledge of the exact value of b_{nT} is of no great importance and an average value of $b_{n20} = 0,04[d^{-1}]$ is accepted.

2.2.3(d) The significance of μ_{nmT} in nitrification

By accepting a set of standard average values for the nitrification constants i.e. $K_{n20} = 1,0[mg\ N.l^{-1}]$ and $b_{n20} = 0,04[d^{-1}]$, Fig. 2.2a compares the effect of μ_{nmT} on R_{sm} and on N_{ae} .

Fig. 2.2a clearly indicates that μ_{nmT} has a marked effect on the value of R_{sm} but at $R_s > 1,3 R_{sm}$ the effect on N_{ae} is insignificant. The knowledge of R_{sm} is crucial in designing a nitrification system hence an accurate estimation of μ_{nmT} is the key to a successful prediction of nitrification behaviour in an activated sludge plant.

The value of μ_{nmT} has been observed to vary with virtually every source of sewage and to vary between different batches of the same source.

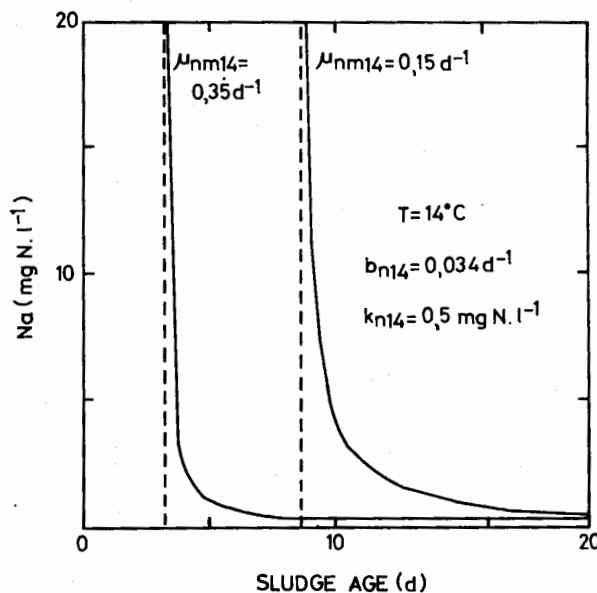


Fig.2.2a: Effect of μ_{nmT} on nitrification.

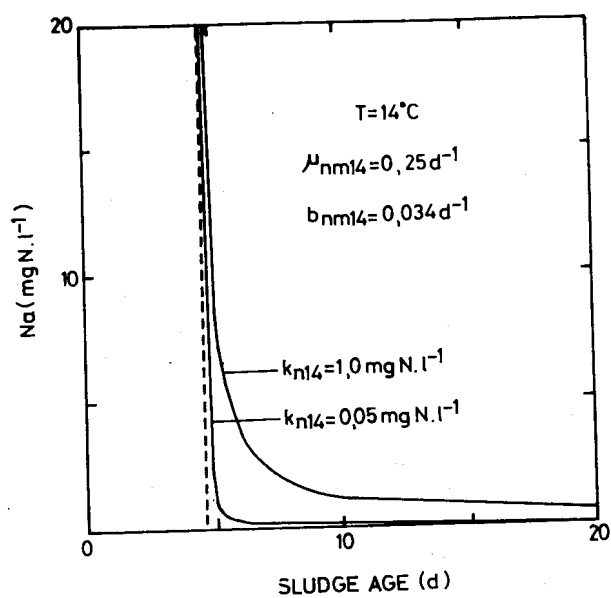


Fig.2.2b: Effect of K_{nT} on nitrification.

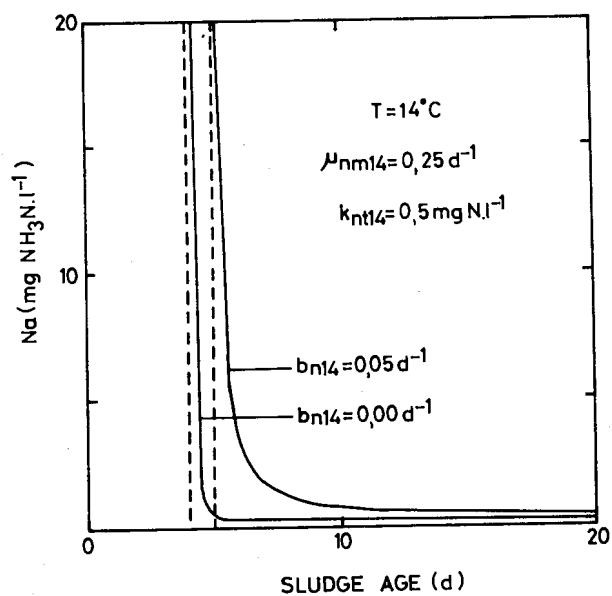


Fig.2.2c: Effect of b_{nT} on nitrification.

It therefore should be classified as a sewage characteristic rather than a kinetic constant. The reason for the marked variations in μ_{nmT} of a certain source is not completely clear. It does not appear to be a matter of toxicity but rather of inhibition and the degree of inhibition seems to be related to the fraction of industrial waste flow in the process. Due to the link between waste flow and μ_{nmT} , the value of μ_{nmT} should always be measured for the specific sewage to be dealt with.

2.2.4 Determination of μ_{nmT}

Theoretically the values of μ_{nmT} , K_{nT} and b_{nT} should be amenable to determination by calibration of Eq. (2.8) to a series of data on N_{ae} obtained over a range of sludge ages. The difficulty with this method is that in the region $R_s > R_{sm}$ the effluent ammonia is very insensitive to μ_{nmT} , b_{nT} and K_{nT} (see Figs. 2.2a, 2.2b and 2.2c). The only measurement that would be of use is to determine μ_{nmT} by operating at certain $R_s = R_{sm}$ but this is a region of instability and it requires patient and tedious experimentation to obtain reliable data.

To overcome the problems of the approach above, Ekama *et al* (1979) employed a series aerobic reactor system under cyclic loading conditions in order to create the transient conditions from which constants can be obtained by calibration. Again this approach has not been completely satisfactory. The uncertainty in this approach stems from the marked hydraulic effects of cyclic loading: slight error in the measurement of the flows results in marked variations in the respective recycles and consequently it is not possible to determine precise estimates for μ_{nmT} .

It is evident from the review above that there is a need for either improvement of the series reactor system or development of an alternative method that allows reliable determination of μ_{nmT} . These two alternatives form a substantial part of the research reported later in this thesis.

2.2.5 Incorporation of nitrification behaviour in aerobic-anoxic systems in a general model

When the general model of Dold, Ekama and Marais (1980) was adapted to aerobic-anoxic environment by Ekama *et al* (1979), the modifications concerning *nitrification* that needed to be made were:

- (1) There is no nitrification in an anoxic environment, i.e.

$$(\mu_{nmT})_{\text{anoxic}} = 0.$$
- (2) Nitrifiers can grow only in an aerobic environment (obligate aerobes) but undergo death (with its associated substrate release by lysis) in both aerobic and unaerated zones.
- (3) The model presupposes that an anoxic environment does not have a direct effect on the nitrification reaction kinetics of a subsequent aerobic environment - experimental evidence reported later in this work supports this hypothesis.

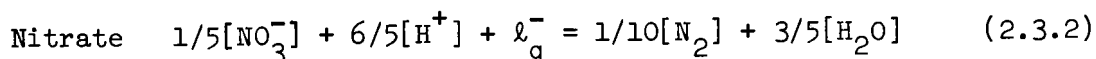
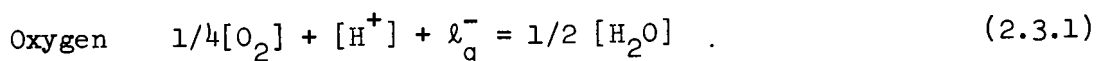
2.3 Denitrification

2.3.1 Stoichiometry of denitrification

Denitrification is a biologically mediated reaction known as dissimilative reduction of nitrate. It involves the reduction of nitrate or nitrite present in the wastewater to gaseous nitrogen, which escapes to the atmosphere.

The denitrification reaction is mediated by heterotrophic organisms; in the denitrification reaction these organisms obtain energy by the oxidation of organic material with nitrate and nitrite acting as electron acceptors. Usually the mass of nitrite present is negligible compared to that of nitrate (see Section 2.1.1) so that the reduction of nitrate only is of significance.

In terms of electron equivalents, the half reactions for the reduction of oxygen and nitrate can be compared:



Considering Eq. (2.3.1), the transfer of one electron equivalent (ℓ_q^-) is accomplished by the reduction of 1/4 mole of oxygen, i.e. 8g.

This transfer corresponds to the oxidation of 8g COD of the organic compounds (from the definition of the COD test below*).

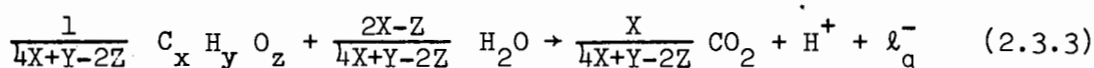
Considering Eq. (2.3.2), the transfer of one ℓ_q^- is accomplished by reduction of 1/5 mole of nitrate i.e. 62/5 g. If nitrate is measured in terms of the nitrogen atom i.e. in terms of $(\text{NO}_3\text{-N})$, then one ℓ_q^- is equivalent to the reduction of $(\frac{62}{5}) \cdot (\frac{14}{62}) = 2,80$ g $(\text{NO}_3\text{-N})$. Equating the electron equivalence of nitrate and oxygen from Eqs. (2.3.2 and 2.3.1) respectively, it follows that

$$2,80 \text{ g } (\text{NO}_3\text{-N}) \equiv 8 \text{ g O}$$

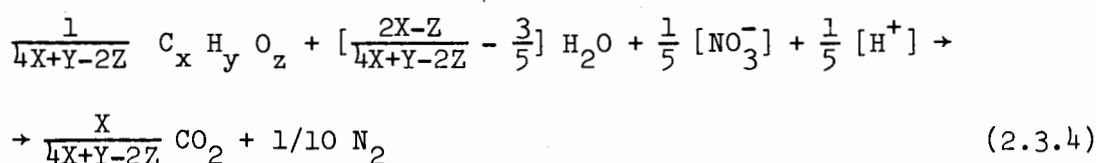
i.e.

$$1 \text{ mg } (\text{NO}_3\text{-N}) \equiv 2,86 \text{ mg O (or mg COD)}.$$

Equation (2.3.2) is a half reaction describing the reduction of nitrate. The corresponding half reaction describing the oxidation of COD (of a general formula $\text{C}_x \text{H}_y \text{O}_z$) is



Combining Eqs. (2.3.2 and 2.3.3), the following redox reaction can be written



Hence, during denitrification of 1 mole of NO_3^- , 1 mole of H^+ is consumed.

2.3.2(a) Oxygen recovery associated with denitrification

In Section 2.1.3 it was shown that for every 1 mg $\text{NH}_4^+\text{-N}$ of ammonia converted to nitrate, 4,57 mg O are consumed, and in Section 2.3.1 that for every 1 mg $(\text{NO}_3\text{-N})$ reduced to nitrogen gas, 2,86 mg O

* The COD test is a measure for the electron transfer potential of the organic matter [after Van Haandel and Marais (1981)].

are recovered equivalently. Thus, in the denitrification process of a single sludge system a fraction $(2,86/4,57) = 0,625$ of the oxygen required for nitrification can be recovered as equivalent oxygen.

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2.3.3 Empirical approach to denitrification

Stern and Marais (1974) found that under *constant flow and load conditions* denitrification is approximately a zero order reaction with respect to nitrate concentration, and directly proportional to the active mass concentration. Hence under constant flow and load conditions the rate of denitrification could be described by an equation of the type

$$\Delta N / \Delta t = -K.X_a$$

Experimentally three rates were identified: in a predenitrification reactor, receiving the influent, two rates were apparent (specified by K_{1T} and K_{2T} rate constants) and in a post-denitrification reactor a single rate, K_{3T} rate constant.

The rate constant K_{1T} was high, giving rise to a rapid rate of denitrification, but it terminated after a few minutes; the total removal of nitrogen associated with K_{1T} was observed to be proportional to the influent COD. The rate associated with K_{2T} continued for the balance of the time in the reactor, at about 1/7 the first rate.

The rate associated with K_{3T} was lower than that associated with K_{2T} (i.e. the value of K_{3T} was lower than that for K_{2T}) and continued for the total period of time in the post-denitrification reactor. The rates were temperature dependent but apparently independent of the sludge age for sludge ages between 10 and 22 days.

The retention time in the pre-denitrification reactor normally is long enough to allow completion of the rate associated with K_{1T} . Consequently it was possible to write the denitrification of a process having both pre- and post-anoxic reactors *under constant flow and load conditions* as follows:

$$\Delta N = \Delta N_1 + \Delta N_2 + \Delta N_3 = \alpha \cdot S_{ti} + K_{2T} \cdot (R_1 - R_0) \cdot X_a + K_{3T} R_3 X_a \quad (2.3.5)$$

where

ΔN = Total nitrogen removal [$\text{mg NO}_3\text{-N} \cdot \ell^{-1}$]

α = Predenitrification mass concentration constant during which K_{1T} is operative [$\text{mg NO}_3\text{-N} \cdot \text{mg COD}^{-1}$]

R_0 = Nominal retention time during which K_{1T} is operative [d]

R_1 = Total nominal retention time in the predenitrification reactor [d]

R_3 = Nominal retention time in the post-denitrification reactor [d]

S_{ti} = Total influent COD concentration [$\text{mg COD} \cdot \ell^{-1}$]

Note that the denitrification formulation expressed in Eq. (2.3.5) is applicable only under constant flow and load conditions, and is empirical, based on observed behavior.

2.3.4 Application of the bi-substrate theory to denitrification

In 1978 (published in 1980) Dold, Ekama and Marais proposed that the biodegradable material in municipal waste flow is composed of an easily biodegradable fraction, f_{ca} , which is of a soluble nature (and is utilized at a high rate by the micro-organisms) and a slowly biodegradable fraction, $(1-f_{ca})$, which is of a particulate nature (and is utilized at a relatively low rate). Ekama, Van Haandel and Marais (1979) suggested that K_{1T} is associated with the utilization

of easily biodegradable substrate while K_{2T} is associated with the utilization of slowly biodegradable substrate. Hypothesizing that the reactions associated with K_{1T} and K_{2T} take place independently and simultaneously instead of sequentially as assumed by Stern and Marais (1974), Ekama *et al* (1979) recalculated K_{1T} , K_{2T} and K_{3T} as

$$K_{1T} = 0,72 (1,2)^{T-20} [\text{mg N. mg } X_a^{-1} \cdot \text{d}^{-1}]$$

$$K_{2T} = 0,101 (1,08)^{T-20} [\quad - \quad - \quad]$$

$$K_{3T} = 0,079 (1,03)^{T-20} [\quad - \quad - \quad]$$

Utilizing the redefinition of K_{1T} and K_{2T} as proposed by Ekama *et al* (1979), Eq. (2.3.5) may be rewritten as

$$\Delta N = \Delta N_1 + \Delta N_2 + \Delta N_3 = \alpha S_{ti} + K_{2T} R_1 X_a + K_{3T} R_3 X_a \quad (2.3.6)$$

It was noted in Section 2.2.2 that, for practical reasons, the value of the maximum anoxic sludge mass fraction, f_{xm} , must not exceed $f_{xm} = 0,50$. This restriction sets an upper limit to denitrification associated with the utilization of the slowly biodegradable substrate:

For a single sludge system, having a pre-denitrification reactor only, the empirical description of denitrification (Eq. 2.3.6) reduces to

$$\Delta N = \Delta N_1 + \Delta N_2 = \alpha S_{ti} + K_{2T} R_1 X_a \quad (2.3.6a)$$

Equation (2.3.6a) indicates that the nitrogen removal associated with the utilization of slowly biodegradable substrate in the pre-denitrification reactor, ΔN_2 , is directly proportional to the nominal hydraulic retention time of the predenitrification reactor, R_1 . i.e.

$$\Delta N_2 = K_{2T} R_1 X_a = K_{2T} (R_1/R_h) X_a R_h$$

where

R_h = Total hydraulic retention time of the process [d].

Now,

$$\frac{R_1}{R_h} = \frac{V_1}{V_T} = \frac{V_1 X_a}{V_T X_a} = \frac{M(X_a)_1}{M(X_a)} = f_{x1}$$

i.e.

$$\Delta N_2 = K_{2T} \cdot f_{x1} \cdot R_h \cdot X_a$$

where

V_1 = Volume of predenitrification reactor [ℓ]

V_T = Total volume of the process [ℓ]

X_a = Active sludge *mass concentration* throughout the process [mg VSS.ℓ⁻¹]

$M(X_a)_1$ = Active sludge *mass* in the predenitrification reactor [mg VSS]

$M(X_a)$ = Total active sludge mass in the process [mg VSS]

f_{x1} = Sludge mass fraction in the predenitrification reactor [mg VSS.mg VSS⁻¹]

For a predenitrification system

$$f_{x1} \leq f_{xm}$$

The magnitude of f_{x1} governs the nitrogen removal associated with the utilization of slowly biodegradable substrate in a predenitrification reactor and is subject to an upper limit, $f_{x1} \leq 0.50$.

Returning to Eq. (2.3.6), this equation constitutes an empirical description of the denitrification behaviour under constant flow and load conditions; essentially the formulation is unrelated to any biological kinetic behaviour.

In order to explain *kinetically* the observed denitrification behaviour under constant flow and load conditions, it was necessary to obtain a general model of activated sludge *kinetics* that describes the denitrification behaviour under constant, cyclic and any other general pattern of flow and load.

2.3.5 Incorporation of denitrification behaviour in the general model

The general activated sludge theory including denitrification is an extension of the general aerobic theory. In order to appreciate the significance of this extension it is useful to examine the development of the general aerobic theory.

Marais and Ekama (1976) developed an aerobic model based on that of Lawrence and McCarty (1970). The latter's model accepted that

- (1) the Monod equation relates the specific organism growth rate to the concentration of soluble substrate surrounding the organism, and
- (2) concomitant with growth, there is a continuous loss of active mass (endogenous respiration). Furthermore, Lawrence and McCarty established the link between sludge age and food/microorganism ratio.

Marais and Ekama (1976) extended the Lawrence and McCarty model by including (1) the generation of endogenous residue due to endogenous respiration as first proposed by McKinney and Symons (1964), and (2) the accumulation of inert volatile solids due to the presence of this material in the influent (McKinney 1966). Furthermore, (3) they established the relationship between oxygen utilization and the synthesis and endogenous respiration reactions, respectively.

Important aspects arising from the model proposed by Marais and Ekama (1976) are:

- (1) A distinction is made between the active and inert sludge fractions.
- (2) The biological reactions are expressed in terms of *active* mass.
- (3) The oxygen uptake rate is determined directly from the biological reactions.

Application of this model to a single reactor with domestic wastewater as influent under *constant* flow and load conditions was in

good agreement with observed results. Application of the model to completely mixed activated sludge (CMAS) process under cyclic flow conditions, using a glucose feed to the system, also showed close correlation between predicted and observed values (Dold, Ekama and Marais 1980) [see Fig. 2.3]. However, when the model was applied to cyclically loaded CMAS processes receiving domestic wastewater as influent and operating at low sludge ages ($R_s = 2,5$ days), it was found impossible to correlate the predicted and observed oxygen consumption rates. At the instant of feed termination there was a precipitous drop in oxygen consumption rate and thereafter a continuous high rate of oxygen consumption for a period of about 2 hours followed by a slow decrease to the oxygen uptake rate associated with endogenous respiration, (see Fig. 2.4). This behaviour could not be accounted for by the Marais-Ekama (1976) model, Fig. 2.4. Despite this inadequacy, the important consequence of the Marais and Ekama model was the recognition of the oxygen utilization rate as a most sensitive parameter against which the activated sludge theory can be tested.

In 1977 Ekama and Marais, based on the work of Andrews and Busby (1973), concluded that the influent COD of domestic waters is *not* soluble but totally of a particulate nature and requires adsorption and storage on the organism, and extracellular enzymatic breakdown before it can be absorbed and metabolized by the organism. They extended Andrews' model for the degradation of particulate material and presented an improved empirical kinetic relationship to determine the rate of utilization of the particulate material. This model described the behaviour of the process under cyclic flow conditions much more realistically than the previous model based on Monod's equation for the soluble COD substrate utilization rate. However, it did not explain the precipitous drop in oxygen utilization rate under a square wave feed pattern at the termination of the feed cycle (see Fig. 2.5a). Ekama and Marais (1978) initially subscribed the drop in oxygen consumption rate to a behavioural characteristic of *autotrophic* nitrification. The acceptance of this hypothesis led to the development of a *modified* model for nitrification that assumed *inter alia* that the conversion of ammonia to nitrate is instantaneous i.e. the model excluded the Monod relationship for nitrification. In

terms of their model it was necessary to accept that only 20 per cent of the TKN was in the ammonia and 80 per cent in the organic nitrogen form. Although the reverse is indicated by chemical measurement, Ekama and Marais justified their choice of the influent nitrogen fractions by hypothesizing that in the ammonia test, in fact some of the proteinaceous material is broken down to ammonia.

The nitrification model also accepted that the nitrogen required for heterotrophic growth is obtained from the organic nitrogen form rather than the ammonia form; the excess organic nitrogen is slowly converted to free and saline ammonia by the heterotrophs' action. Incorporation of these modifications into the general model gave the response shown in Fig 2.5b for 2,5 days sludge age. The precipitous decrease in the total oxygen consumption rate was now adequately predicted. Although the model was apparently successful, the exclusion of the Monod relationship for nitrification in favour of a different nitrification theory, was difficult to justify unless very powerful supporting evidence is presented.

Work by Wilson and Marais (1976) led to a reassessment of the model. Ekama and Marais (1978) hypothesized that the precipitous drop in oxygen consumption rate at feed termination may be attributed to cessation of a *heterotrophic* energy requirement for adsorption of the carbonaceous particulate substrate, caused by feed termination. (The feed was still regarded as being *all* of a particulate nature). They included this energy (oxygen) requirement for adsorption in the mathematical model and reinstated Monod relationship for nitrification, and accepted the experimentally measured nitrogen influent fractions (i.e. approximately 80% *free and saline ammonia* and 20% *organic nitrogen*). However they retained the concept of organic nitrogen storage associated with stored COD and the slow conversion of organic nitrogen to free and saline ammonia by the heterotrophs. A comparison of the response of this modified model with the experimental response at 2,5 days sludge age is shown by Fig. 2.6. The modified model showed a significantly improved correlation between the experimental and simulated oxygen consumption rates and nitrate concentrations. Indeed, generally, the Ekama and Marais (1978) model describes the behaviour of an aerobic CMAS process very adequately.

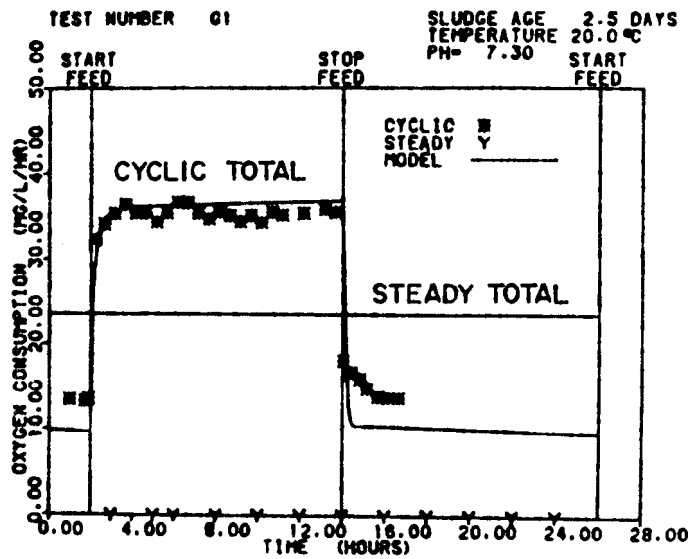


Fig.2.3: Comparison of simulated and observed response of a CMASP under cyclic flow and load with a glucose feed to the system using the Ekama & Marais(1976) model.

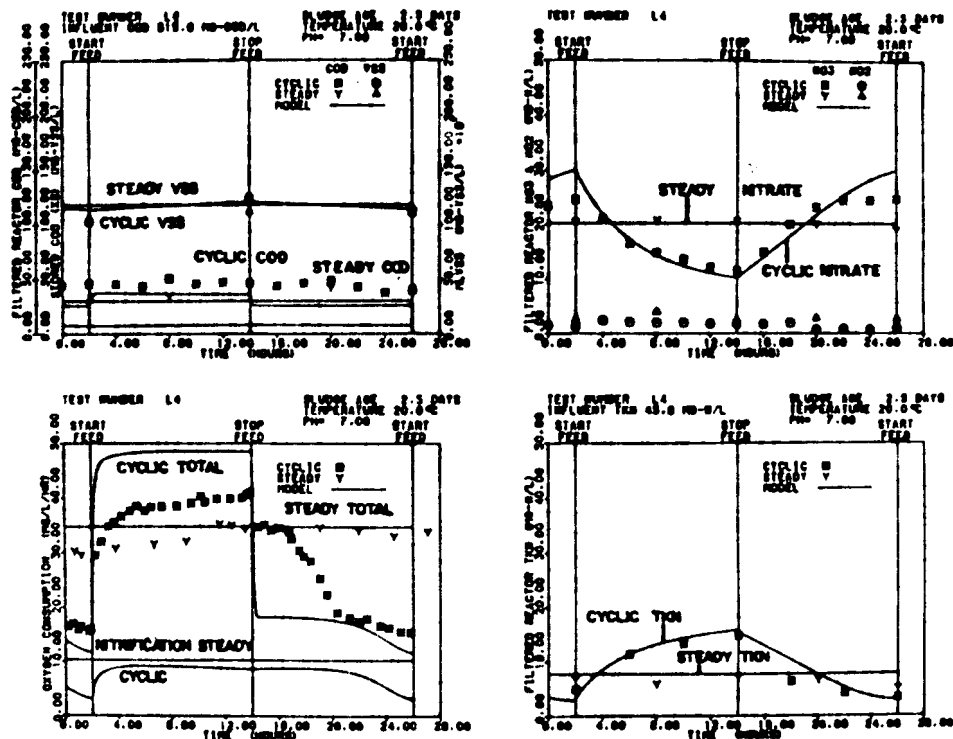


Fig.2.4: Comparison of simulated and observed response of a CMASP under cyclic flow and load with domestic sewage feed using the Ekama & Marais(1976) model.

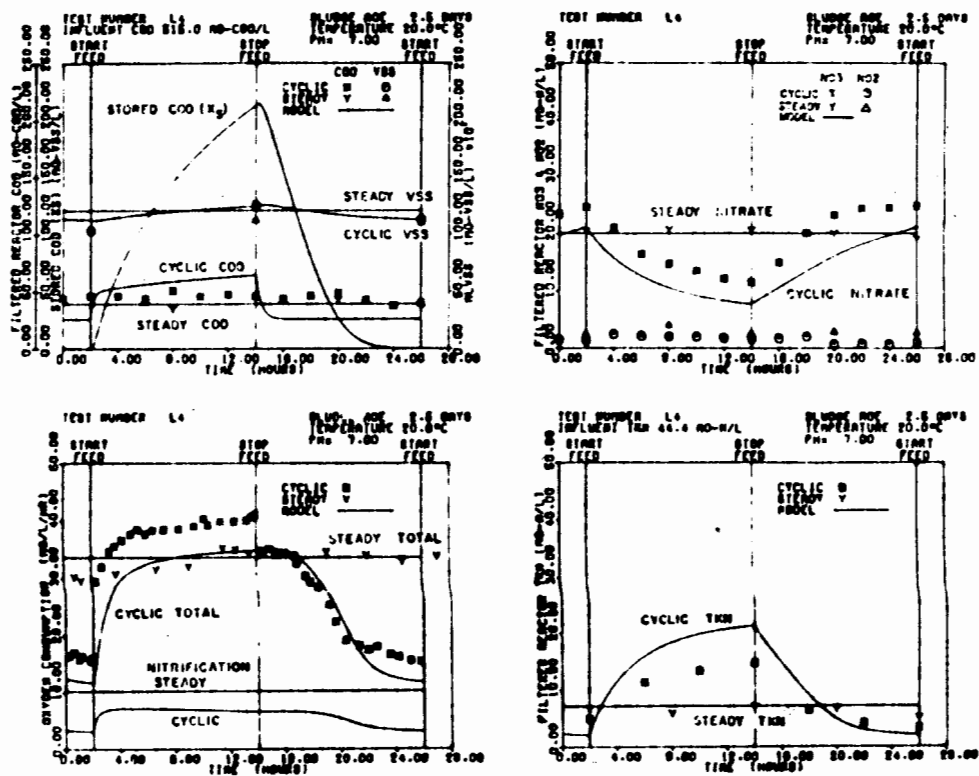


Fig.2.5a: Comparison of simulated and observed response of a CMASP under cyclic flow and load with sewage feed assuming the influent to be totally of a particulate nature [After Ekama & Marais(1977)].

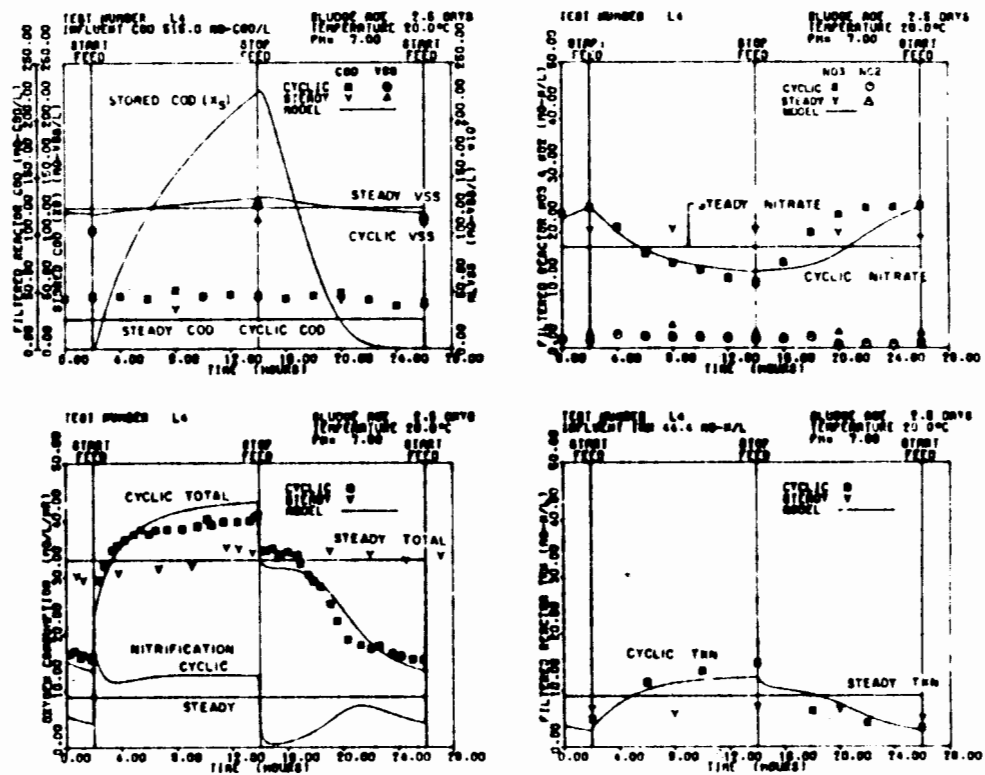


Fig.2.5b: Comparison of simulated and observed response of a CMASP under cyclic flow and load with sewage feed utilizing the Ekama & Marais(1977) model and incorporating a modified nitrification model.

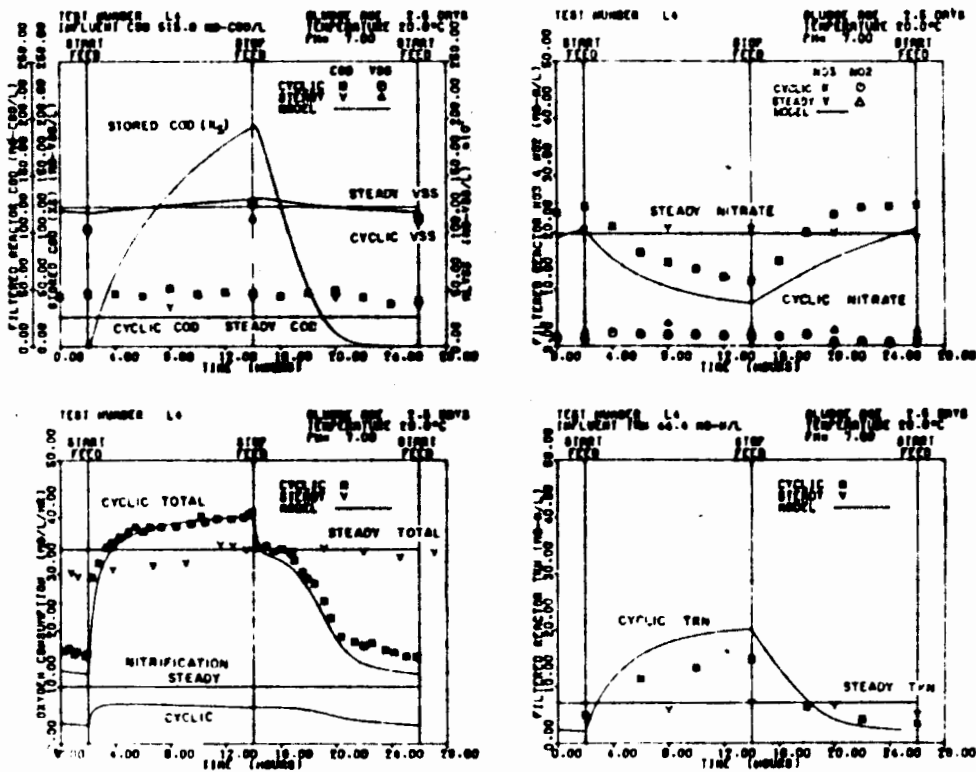


Fig.2.6: Comparison of simulated and observed response of a CMASP under cyclic flow and load with sewage feed assuming the influent to be totally of a particulate nature and incorporating the adsorption phenomena [After Ekama & Marais(1978)].

In 1979 (published in 1980) Dold, Ekama and Marais critically reviewed the model of Ekama and Marais and concluded that certain hypotheses in the Ekama and Marais model needed replacement and proposed the following:

- (1) The biodegradable substrate consists of two fractions easily biodegradable soluble and slowly biodegradable particulate. The rate of utilization of the easily biodegradable material is governed by the Monod equation (Eq. 2.3.11) while that of the slowly biodegradable material is governed by a Levenspiel (1972) type equation for surface active reactions (Eq. 2.3.13).
- (2) The Monod and Levenspiel reactions take place independently and simultaneously. Dold *et al* justified this assumption by hypothesizing that as the organisms essentially remain in a continuous state of stress due to the low food/microorganism ratio found in most types of activated sludge processes, it is unlikely that, in the range of operational sludge ages, the particulate material (stored on the active sites) will occupy any but a small fraction of the active sites. The only process where the particulate material may occupy a substantial fraction of the maximum fraction of substrate that can be stored on the active sites, (f_{ma}), is in the contact chamber of a contact stabilization process and, in aerated lagoons with short hydraulic retention time 1 - 2 days.*

*Justification for the independence of the two reactions is to be found in analysing the ratio of stored material to active mass by means of the general theory of Dold, Ekama and Marais. A measure of the fraction of *unoccupied* active sites is given by $[f_{ma} - (X_s/X_a)]$. In Fig. 2.9 a plot is shown of $[f_{ma} - (X_s/X_a)]$ vs. sludge age. It is apparent that the activated sludge process is characterized by a minimum sludge age for particulate organic substrate utilization (similar to that for nitrification) below which all the active sites are occupied and the reaction achieves a stationary upper limit of utilization per unit mass of X_a .

Above about 2 days sludge age the percentage of sites available remains relatively constant in the range 90-100 per cent so that the response to soluble waste will be virtually as if no particulate material is present, i.e. the reactions of easily biodegradable and particulate biodegradable substrate, respectively, are virtually independent of each other. The interdependence of the easily biodegradable and particulate substrate, respectively, would develop in the range below about $R_s = 2$ days; within this range the rate of substrate utilization is uncertain.

- (3) The energy requirement for adsorption can be relinquished, the precipitous drop in oxygen utilization rate at cyclic feed termination being due to termination of the easily biodegradable COD.

The modified (1979) aerobic model gave predictions very close to the observed data. However at a later stage during denitrification studies with Van Haandel, the model above showed certain inconsistencies in that the observed oxygen utilization of an aerobic reactor receiving mixed liquor from an unaerated reactor was higher than that obtained by the model prediction. All the models up to this stage assumed the synthesis-endogenous respiration approach. Basically this approach assumes that the organism is generated and subsequently undergoes mass loss due to a cell maintenance energy requirement. If this is so indeed, then a mixed liquor sample placed under batch conditions without feed addition should show an "ageing" effect. However Marais and Ekama found that the ratios of oxygen utilization rates between any two consecutive days remain constant, i.e. no ageing effect is observed.

In order to explain the behaviour characteristics described above, they hypothesized a different model for the behaviour of the organisms, i.e. the death-regeneration model. In the death-regeneration model it is hypothesized that the percentage of live mass that disappears per unit time in fact dies, lyse biodegradable substrate back into the liquid, and leaves the balance as unbiodegradable endogenous residue. The lysed biodegradable substrate is added to the biodegradable COD in the liquid and passes through the same phases of adsorption, storage and utilization. This approach, in effect, implies that endogenous respiration (cell maintenance) *per se* does not exist and that oxygen is consumed only for synthesis of new cell material from the lysed energy. In terms of this model, the average life expectation is about 1.1 days, i.e. 62 per cent of the organisms die each day and 38 per cent is regenerated giving rise to a nett decrease of 24 per cent per day. Once the sludge age exceeds about 1-2 days, the regeneration effect virtually keeps the mass at an actual organism life expectancy of 1.1 days, i.e. no ageing effect

would become apparent. Also the observations made by Ekama *et al* (1979) that high oxygen uptake rate is observed in an aerobic reactor that follows an anaerobic reactor, can be now explained: under anaerobic conditions synthesis ceases, but organism death, with associated lysis of biodegradable material continues, resulting in a build up of biodegradable material in the reactor. When effluent from the anaerobic reactor passes into an aerobic reactor, in which oxygen is available, the aerobic biodegradation of the accumulation of biodegradable substrate causes a high oxygen utilization.

The simulated response of a cyclically loaded *aerobic* CMAS process obtained by using the bi-substrate active-site death-regeneration model, when compared with observed response, gives excellent correlation not only at 2,5 days sludge age but also for a variety of process configurations under a number of influent flow and load patterns. Figure 2.7 is one simulation of a square wave, cyclically loaded 2,5 days sludge age aerobic process.

Ekama, Van Haandel and Marais (1979) extended the death-regeneration aerobic general model to include denitrification. This required making the following adjustments:

- (1) Growth of nitrifiers can take place only under aerobic conditions; death is assumed to take place irrespective of aerobic or anoxic conditions.
- (2) It is assumed that an anoxic environment does not have a direct effect on the nitrification reaction kinetics of a subsequent aerobic environment.
- (3) Growth of heterotrophs is assumed to take place in both aerobic and anoxic conditions (i.e. provided nitrate or dissolved oxygen is available); death and lyses take place irrespective of whether the conditions are aerobic or anoxic.

Calibration of the modified general model by Van Haandel and Marais (1981), using data on pre- and post-denitrification reactors generated in a plug flow regime under constant flow and load conditions indicated that:

- (1) For close fits between simulated and observed data in the post-denitrification reactor, the lysed material should be of a particulate nature.
- (2) In an anoxic reactor, the utilization rate of the particulate matter takes place at a slower rate than that in an aerobic reactor [see Eq. 2.3.18)].

To indicate the adaptations that needed to be made in order to extend the aerobic model to include denitrification, the differential equations for the general aerobic model describing heterotrophic reactions are set out below:

Let

$$A = K_{msT} \cdot S_{bs} / (S_{bs} + K_{ss}) \quad (2.3.7)^*$$

$$B = K_{vT} \cdot S_{bp} \cdot (f_{ma} - X_s/X_a) \quad (2.3.8)^*$$

$$C = K_{mpT} \cdot P \cdot X_s / (X_s P + K_{spT} X_a) \quad (2.3.9)^*$$

$$D = P \cdot b_{hT}^1 \quad (2.3.10)^*$$

Then the differential equations for energy removal are:

$$dS_{bs}/dt = -A \cdot X_a \quad (2.3.11)^*$$

$$dS_{bp}/dt = [-B + (1-f')D] \cdot X_a \quad (2.3.12)^*$$

$$dX_s/dt = [(B-C)/P] \cdot X_a \quad (2.3.13)^*$$

$$dX_a/dt = [Y_h(A + C) - D/P] \cdot X_a \quad (2.3.14)^*$$

$$dX_e/dt = [f'D/P] \cdot X_a \quad (2.3.15)^*$$

$$O_c = [(1-PY_h)(A + C)] \cdot X_a \quad (2.3.16)^*$$

These equations apply also to an anoxic environment provided the following adjustments are made:

- (1) Equation (2.3.16) describing the oxygen utilization rate in an anoxic environment needs adjustment in order to express the utilization rate in terms of nitrate instead of oxygen, i.e.

$$(dN_n/dt) = -(1-PY_h)(A+C) \cdot X_a / 2,86 \quad (2.3.17)^*$$

* The meaning of symbols in the equation is given by a list of symbols given before Chapter 1.

- (2) The utilization rate of particulate substrate under anoxic conditions has been shown earlier to be lower than that under aerobic conditions. This is incorporated into the general model by including an adjustment constant, η , linking the slowly biodegradable substrate utilization rate constant in the *aerobic* reactor to that in the *anoxic* reactor, i.e. Eq. (2.3.17) is modified to

$$(dN_n/dt) = -(1-PY_h) (A+C') \cdot X_a / 2,86 \quad (2.3.18)^*$$

where

$$C' = \eta \cdot C = \frac{K'_{mpT}}{K_{mpT}} \cdot C \quad (2.3.19)^*$$

and K'_{mpT} is defined as the slowly biodegradable substrate utilization rate constant under anoxic conditions [$\text{mg COD} \cdot \text{mg VSS}^{-1} \cdot \text{d}^{-1}$].

In effect, apart from the convenient changes from oxygen to nitrate, only one significant change was made -- the reduction of K_{mpT} to K'_{mpT} . No structural kinetic changes were made to the model.

In the light of the above modifications made in order to incorporate the denitrification behaviour in the general model, it is now possible to interpret the significance of the empirical constants discussed in Section 2.3.4 in terms of parameters involved in the general model.

2.3.6 Interpretation of the empirical approach to denitrification

From Section 2.3.5 it follows that in terms of the bi-substrate death-regeneration model, the rate of nitrate utilization dN_n/dt , can be viewed to be comprised of a rate associated with the utilization of easily biodegradable material, dN_1/dt , and a rate associated with the utilization of slowly biodegradable material, dN_2/dt , i.e.

$$dN_n/dt = dN_1/dt + dN_2/dt \quad (2.3.20)$$

where

* The meaning of symbols in the equation is given by a list of symbols given before Chapter 1.

$$dN_1/dt = -[(1-PY_h).A/2,86].X_a \quad (2.3.21)$$

$$\text{and } dN_2/dt = -[(1-PY_h).C'/2,86].X_a \quad (2.3.22)$$

In Eq. (2.3.6), ΔN_1 and ΔN_2 are the corresponding empirical equations of Eqs. (2.3.21 and 2.3.22) respectively. If "A" is substituted from Eq. (2.3.7) into Eq. (2.3.21) and "C" is substituted from Eq. (2.3.19) into Eq. (2.3.22) respectively, the significance of the empirical K_{1T} and K_{2T} respectively, in denitrification as interpreted by the general model may be given by

$$K_{1T} = [(1-PY_h)K_{msT}.S_{bs}/(K_{ss}+S_{bs})]/2,86 \quad (2.3.23)$$

$$\text{and } K_{2T} = [(1-PY_h).K'_{mpT}.P.X_s/(P.X_s+K_{spT}.X_a)]/2,86 \quad (2.3.24)$$

With regard to the utilization of influent easily biodegradable substrate (S_{bsi}) that is implied by Eq. (2.3.23), in a pre-denitrification reactor normally all the influent easily biodegradable substrate is completely utilized and hence knowledge of the exact value of K_{1T} is consequential only in so far as the rate of reaction is concerned but is inconsequential in so far as the *mass* of denitrified nitrate associated with the utilization of easily biodegradable material is concerned. This mass of nitrate removed can be expressed as

$$\Delta N_1 = (1-PY_h).S_{bsi}/2,86 \quad (2.3.25)$$

Equation (2.3.25) implies that for all practical purposes the nitrate removal due to utilization of easily biodegradable material depends solely on the value of S_{bsi} which is linked to the value of f_{ca} as follows:

$$f_{ca} = S_{bsi}/S_{bi} = S_{bsi}/[S_{ti}(1-f_{us}-P.f_{up})]$$

and the value of K_{msT} in Eq. (2.3.23) is of minor significance.

With regard to the utilization of slowly biodegradable substrate (S_{bp}) that is implied by Eq. (2.3.24), the nitrate removal due to utilization of slowly biodegradable material depends on the value of K'_{mpT} which in turn is linked to K_{mpT} by η .

2.3.7. Identification of the kinetic constants that most significantly affect denitrification

Van Haandel, Ekama and Marais, in a paper to be published

in Water Research (1981), compared the experimental and simulated response of an anoxic-aerobic series reactor system under cyclic loading conditions. They found that they could always obtain very good correlation [see Fig. 2.8(a and b)] provided it was accepted that some of the constants were affected by the influent sewage characteristics. Now, either the theory is not adequate or necessarily the constants are specific to a sewage. If the latter, then this implies that for every sewage it will be good practice to measure the constants. With regard to the former, the wide range of experiments presented by Van Haandel and Marais and the generally good qualitative consistencies obtained between observed and simulated data, provide powerful evidence that the basic kinetic description is adequate. Hence the problem develops in determining the constants for each specific waste flow. However not all the constants need to be determined with equal accuracy. By simulation, using various values for the denitrification constants, Van Haandel and Marais found that the response of the plant is very sensitive to the magnitudes of some of the constants and not so for others. Consequently those that have a sensitive effect should be determined more reliably than those having a relatively insensitive influence. The significant constants appear to be the following:

- (1) The slowly biodegradable (particulate) substrate utilization rate constant, K_{mpT} .
- (2) The ratio of slowly biodegradable substrate utilization rate constant under anoxic conditions to that under aerobic conditions, $K'_{mpT}/K_{mpT} = \eta$.
- (3) The easily biodegradable COD fraction of the influent, $f_{ca} = S_{bsi}/S_{bi}$.
- (4) The easily biodegradable substrate utilization rate constant, K_{msT} .

Accurate estimation of the denitrification constants is quite complex because a particular response may be significantly affected by some or all the other constants and it is often difficult to isolate the effects of one constant. This is apparent, in

particular, in plug flow reactor response under constant flow and load wherein the effects of the different reactions have led to the establishment of empirical constants K_{1T} , K_{2T} and K_{3T} . [see Section 2.3.4].

2.3.8 Determination of f_{ca} , K_{mpT} and η

The factors affecting denitrification have been identified in the previous section as

- (1) Nitrate removal associated with utilization of easily biodegradable material, i.e. f_{ca} , and
- (2) Nitrate removal associated with utilization of slowly biodegradable material, i.e. K_{mpT} and η .

Dold, Ekama and Marais (1980) evaluated f_{ca} by imposing square wave cyclic load conditions (12 hours on 12 hours off) on a single completely mixed aerobic reactor operating at a sludge age of about 2.5 days. At this sludge age the rate of nitrification and the rate of utilization of slowly biodegradable material is virtually negligible compared to the rate of utilization of easily biodegradable material. The precipitous drop in oxygen uptake rate at feed cessation, ΔO , is solely due to the termination of the synthesis associated with the easily biodegradable COD fraction. Hence f_{ca} is given by

$$f_{ca} = \frac{\Delta O}{(1-PY_h)} \cdot \frac{1}{S_{bi}} \cdot \frac{V}{Q_a}$$

where

V = Volume of process (single completely mixed reactor) [l]

Q_a = Actual volumetric flow rate during feeding period [$l \cdot h^{-1}$]

$S_{bi} = S_{ti}(1-f_{us}-P \cdot f_{up})$ = Influent biodegradable COD [$mg \text{ COD} \cdot l^{-1}$]

ΔO = The precipitous drop in oxygen uptake rate at feed cessation [$mg \text{ O}_2 \cdot l^{-1} \cdot h^{-1}$]

The value of η was determined by Ekama, van Haandel and Marais (1979) who compared the *average* observed experimental denitrification rate with the theoretically predicted equivalent oxygen uptake rate in "pre-aerobic" and "post-aerobic" plug flow

systems, using constants developed from *mean* aerobic system response of plants operated under cyclic flow conditions. The ratio of the slope of equivalent oxygen uptake rate in a *post-denitrification* reactor to the oxygen uptake rate in a comparable "post-aerobic" reactor was found to be $\eta = 0,38$. This ratio was also observed with the denitrification associated with slowly biodegradable substrate utilization in a *pre-denitrification* reactor. Hence the utilization rate constant of slowly biodegradable material in an anoxic reactor was made a fraction $\eta = 0,38$ of that in a comparable aerobic reactor, i.e. $K'_{mpT} = \eta \cdot K_{mpT}$. Hence, K_{2T} , associated with the utilization of slowly biodegradable material in an anoxic reactor becomes dependent on K'_{mpT} [see Eq. (2.3.24)]. Ekama *et al* (1979) assessed the temperature dependency of K_{mpT} to be

$$K_{mpT} = K_{mp20} (1,029)^{T-20} [\text{mg COD.mg VSS.}^{-1}\text{d}^{-1}].$$

The estimation of K_{mpT} was obtained by accepting $\eta = 0,38$ in calibration of a cyclically (square wave) loaded anoxic-aerobic reactor series [Ekama *et al* (1979)]. This approach has not been completely satisfactory. The uncertainty in this approach stems from the marked hydraulic effects of cyclic loading [see Section 2.2.4]. In addition, as K_{mpT} and η act in a mutually interactive manner because ($\eta \cdot K_{mpT} = K'_{mpT}$), the individual effect of either one could not be investigated. In the series system experiments η was empirically accepted as constant and the variation in the denitrification rate was ascribed solely to variations in K_{mpT} .

It is evident from the review above that in order to accurately predict the denitrification behaviour, it is necessary that constants f_{ca} , η and K_{mpT} should be accurately evaluated. Hence there is a need for development of a method that allows reliable determination of these constants. The experimental investigation that leads to a suitable method is reported in Chapter 3.

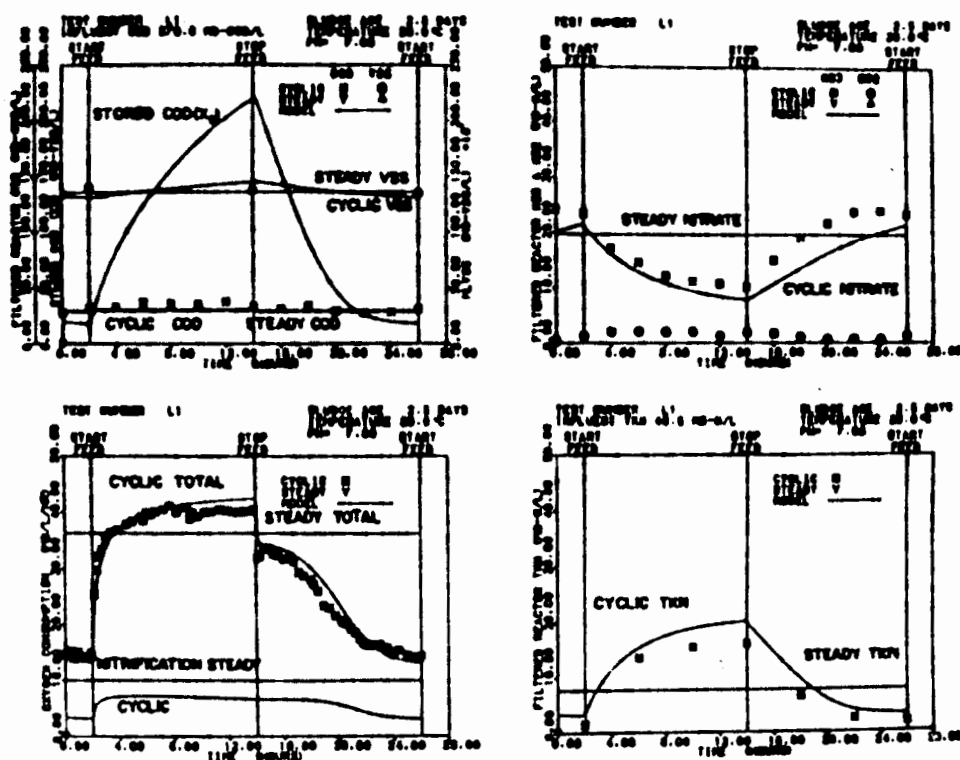


Fig.2.7: Comparison of simulated and observed response of a CMASP under cyclic flow and load with sewage feed using the bi-substrate active-site death-regeneration aerobic model of Dold, Ekama & Marais(1980).

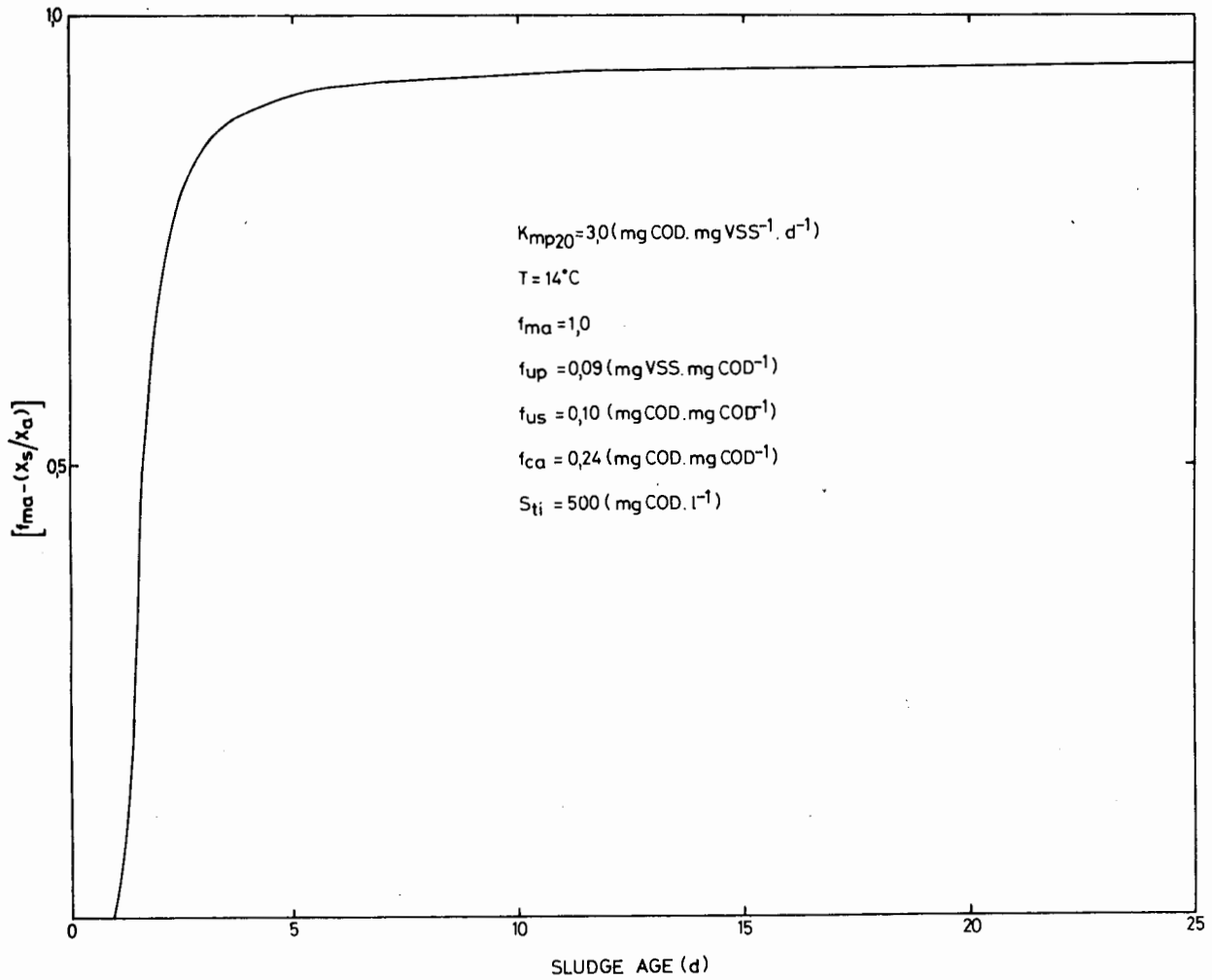


Fig.2.9: Measure of the unoccupied active sites as a function of sludge age.

CHAPTER 3

EXPERIMENTAL INVESTIGATION

3.1 Introduction

The objective of the experimental investigation was to devise experimental set ups and operational procedures which would allow accurate determination of the nitrification and denitrification constants on processes having stable response characteristics. Two approaches towards attaining these objectives were tried. In the first approach a series reactor nitrification-denitrification system was used under constant flow conditions in which the relative volumes of the reactors were chosen such that a sensitive evaluation of the constants should be possible. In the second approach the alternating anoxic-aerobic completely mixed single reactor (CMSR) system under square wave flow and load conditions was investigated.

Series reactor systems had previously been used extensively by Van Haandel, Ekama and Marais [to be published in Water Research (1981)] to determine the constants by calibration. A 3 reactor predenitrification laboratory scale unit was employed (20% predenitrification reactor, 20% first aerobic reactor, 60% second aerobic reactor) and constant and cyclic flow and load conditions, respectively, were imposed. Concurrently a five equal-sized reactor (20% each) pilot scale series predenitrification plant was tested under constant and cyclic flow and load conditions, respectively.

Under cyclic flow and load conditions the observed response on both the 3 and 5 reactor system, respectively, tended to be unstable so that accurate determination of the constants by calibration was not possible. Furthermore the response of the 5 reactor system proved to be more unstable than that of the 3 reactor system. In contrast, under constant flow and load conditions the response was stable indicating that the instability was due to the cyclic nature of the flow and load but which of these contributed the most to the instability could not be ascertained from *observations*.

The response of the system under constant flow and load,

Alternating anoxic-aerobic CMSR system had been investigated at 20°C under *constant* flow and load conditions by Van Haandel and Marais (1981). The response indicated very good stability and good potential for determining the nitrification and denitrification constants. This was confirmed also at 14°C (see section 3.3.3). However this approach required knowledge of the fraction of easily biodegradable influent COD. To determine this fraction required a separate special experimental set up and experimental investigation. The value of alternating anoxic-aerobic approach would be greatly increased if it was possible to determine the easily biodegradable fraction from the same set up as that used for the determination of the other kinetic constants. It was hypothesized that if square wave flow and load pattern was imposed on an alternating anoxic-aerobic CMSR, by suitably choosing the anoxic times within the cycle, it should be possible to determine not only the nitrification and denitrification *kinetic* constants but also the fraction of easily biodegradable influent COD. The second investigation was oriented towards this approach and is discussed in Section 3.3.

3.2 Series System Investigation

In the previous section it was stated that the instability of the response of the 3 reactor series predenitrification system under cyclic flow *and* load was attributed primarily to the cyclic *flow*. It was hypothesized that if the cyclic *flow* were replaced by constant *flow* with an imposed *cyclic TKN load*, a stable response would be obtained that would allow maximum extent of information about *both* nitrification and denitrification.

The major considerations that determined the system configuration and its operational characteristics were:

- (a) Information on denitrification can be obtained from an analysis of nitrate removal only if the nitrate entering the anoxic reactor exceeds the denitrification capacity of the reactor, i.e.: the mixed liquor and underflow recycle, respectively must be high enough such that

nitrate is always present in the stream leaving the anoxic reactor.

- (b) In order to obtain maximum sensitivity in the data from a reactor the change of the parameter in question must be the maximum possible. In denitrification, the maximum changes will be obtained by having an anoxic reactor as large as possible. However, the anoxic mass fraction in a system cannot be increased *ad lib*; if the anoxic sludge mass fraction exceeds about 40 - 50 percent, poor settling often is encountered to a degree that the system cannot be operated. This has been repeatedly observed by Marsden & Marais (1976) and Arkley & Marais (1981).
- (c) Information concerning nitrification and denitrification can be obtained from a predenitrification system. Secondary anoxic reactor has little value as the reduction in nitrate is principally due to endogenous mass loss. For this reason a post-denitrification reactor should be omitted.

Based on the considerations above and by using trial simulation the system configuration selected is shown in Fig. 3.1 and the basic process and sewage parameters listed in Table 3.1.

In Fig. 3.1 the anoxic sludge mass fraction was selected at 40 percent which is the maximum allowable to still retain good settling. To obtain maximum activity in the sludge the shortest practical sludge age was selected. With 40 percent anoxic sludge mass fraction at 14°C, 15 days sludge age was the minimum that allowed continuous and stable nitrification*. The first aerobic reactor was chosen to have a small retention time (about 20 percent of the total process volume). With

* Later 10 days sludge age was tried, but nitrification consistently declined even with repeated reinoculation of nitrifying sludge. This clearly indicates that 15 days sludge age was a good choice.

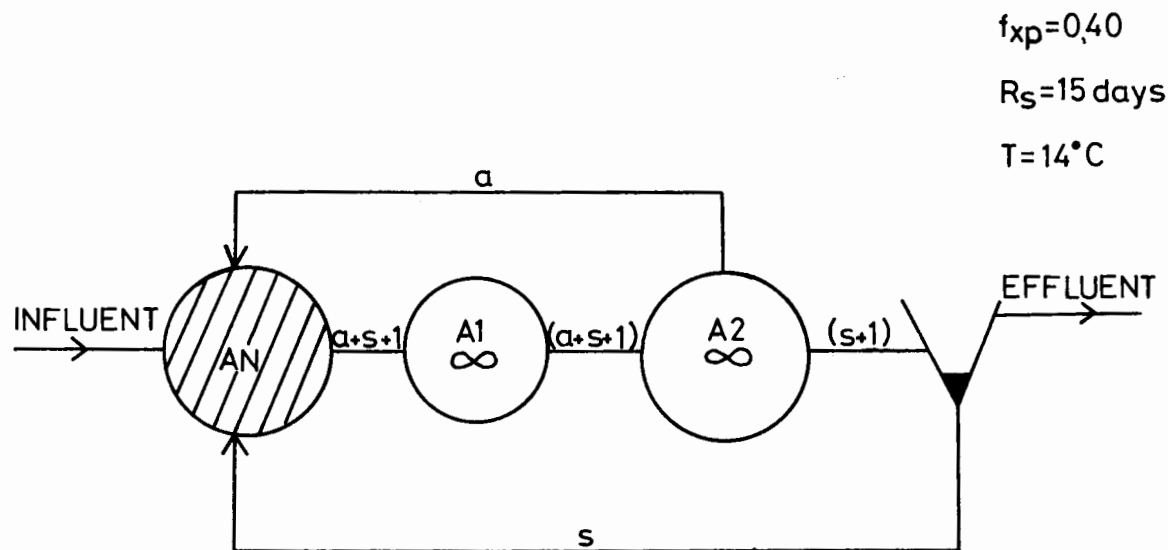


Fig.3.1: Single sludge predenitrification series reactor system employed in the determination of nitrification and denitrification constants.

this fraction it was proposed that the best conditions were available to ensure that:

- (1) The maximum nitrification rate constant would be operational and could be determined from the changes in nitrate concentrations.
- (2) Any differences in the nitrification rate between the first and the second aerobic reactors would allow investigation of possible effects of the anoxic state on subsequent nitrification.

Table 3.1 *Operational and sewage characteristics used in Fig. 3.1.*

<u>Operational Characteristics :</u>	
$T = 14^{\circ}\text{C}$	
$R_s = 15 \text{ days}$	
Anoxic reactor volume, V_{AN}	$= 4,67 \text{ [l]}$
First aerobic reactor volume, V_{A1}	$= 2,5 \text{ [l]}$
Second aerobic reactor volume, V_{A2}	$= 4,5 \text{ [l]}$
Base feed flow rate, Q_b	$= 17,2 \text{ [l.d}^{-1}\text{]}$
<u>Sewage Characteristics :</u>	
$f_{up} = 0,09 \text{ [mg VSS.mgCOD}^{-1}\text{]}$	$f_{na} = 0,75 \text{ [mgNH}_3\text{-N.mgN}^{-1}\text{]}$
$f_{us} = 0,15 \text{ [mg COD.mgCOD}^{-1}\text{]}$	$f_{nu} = 0,0 \text{ [mgN.mgN}^{-1}\text{]}$

3.2.1 Predenitrification series system under constant flow and load conditions.

The series system in Fig. 3.1 was initially run under *constant flow and load*. The objective in operating under constant flow and load was to develop an accurate experimental procedure to ensure that all the process variables were correctly measured so that when a cyclic load was imposed, any operational or analytical uncertainties would have been resolved and attention could be focused on the objective of the investigation - the determination of the magnitude of the kinetic constants in question.

Proper measurement of all process variables is indicated if satisfactory mass balances are achieved on the system operated under constant flow and load. From theoretical mass balance calculations, it was apparent that of all the measurable process variables : COD, VSS, TKN, NO_3 , Oxygen Uptake Rate (O.U.R.) and recycle flow rates, inaccurate measurement of the recycle flow rates has the most sensitive effect on the mass balance. The reason for this is that the recycle rates are quite high ($a+s=6$) so that any error in the measurement of concentration of a parameter involved in the mass balance has a multiplicative effect on the mass balance calculation. Consequently particular attention was given to measuring the recycle flows and checking them daily. The system was run for a period of about two months during which period eight batches of sewage were used and the forementioned process variables measured daily. From the measured process parameters the calculation of mass balances of nitrogen and COD were made. The procedure for calculating the mass balances is set out in detail in Appendix B together with the computer program (program B.1) based on this procedure.

A summary of the average response for each batch of sewage, and the respective nitrogen and COD balances as calculated by the mass balance program is listed in Table 3.2.

Table 3.2 *Average observed response and output of mass balance program for the system in Fig. 3.1 operated under constant flow and load.*

SEWAGE BATCH CODE** NO.	1	2	3	4	5	6	7*	8*
DATE	21/4-23/4	24/4-2/5	3/5-9/5	10/5-16/5	20/5-22/5	25/5-30/5	31/5-8/6	9/6-13/6
CODIN [mgCOD.l ⁻¹]	494,0	493,4	483,6	509,0	506,3	480,4	488,5	489,3
CODOUT[mgCOD.l ⁻¹]	37,3	68,4	51,7	71,2	54,0	56,7	52,4	55,5
TKNIN [mgN.l ⁻¹]	59,1	55,9	52,4	49,9	49,6	49,2	49,2	50,1
TKNOUT[mgN.l ⁻¹]	1,8	3,5	1,5	3,0	2,2	5,1	2,2	3,1
OUR ₁ [mgO.l ⁻¹ .h ⁻¹]	49,2	43,0	46,8	42,4	37,2	34,4	44,3	48,7
OUR ₂ [mgO.l ⁻¹ .h ⁻¹]	20,2	18,0	22,4	26,7	25,1	24,2	23,5	21,6
NO ₃ AN[mgN.l ⁻¹]	18,0	14,1	14,1	15,0	10,6	9,8	11,5	10,0
NO ₃ A2[mgN.l ⁻¹]	23,3	18,8	18,8	19,4	15,0	12,9	16,3	14,8
NO ₃ EFF[mgN.l ⁻¹]	22,8	19,7	18,8	21,9	15,8	11,2	16,4	14,9
DNIT [mgN.l ⁻¹]	23,0	23,5	22,6	21,9	21,9	14,4	17,6	19,1
NRECOV [%]	98,7	102,8	103,2	117,1	104,1	85,9	<u>98,4</u>	<u>98,1</u>
CODREC [%]	75,9	78,8	87,6	90,6	85,7	91,7	<u>92,8</u>	<u>94,2</u>

* Batch of sewage for which satisfactory mass balances were obtained on both nitrogen and COD. Mass balances were considered satisfactory when mass recoveries of 95 percent and 90 percent or higher for nitrogen and COD mass balance respectively were obtained.

** Code designation is given in the mass balance program (program B.1) in Appendix B under program output variables.

From Table 3.2 it is apparent that initially the mass balances were poor. With time, the procedures for calibrating the recycle pumps were improved and concomitantly more satisfactory balances were achieved, after six weeks of operation (Batch numbers 7 and 8). It was very evident from the mass balance evaluation that in this type of process unless extreme care is taken with the measurement of the recycles, unreliable results are the inevitable consequence.

No attempt was made to determine the nitrification and denitrification constants from the series system under constant flow and load. Two observations supported this decision:

- (1) Table 3.2(a) gives the TKN response for batches of sewage that gave satisfactory mass balances. It is apparent that the changes in TKN concentration were small and the TKN response was therefore insensitive to different values of μ_{nmT} . Also, the actual TKN concentrations in the aerobic reactors were usually low because of the dilution effect of the influent TKN by the high mixed liquor recycle, a, and underflow recycle, s, respectively and errors in the concentrations led to large errors when using differences.
- (2) The short retention time in the first aerobic reactor, A1, gave rise to small changes in nitrate concentrations entering and leaving the reactor. From experimental data (Table 3.2(b)), for batches of sewage that gave satisfactory mass balances, the average difference in nitrate concentrations entering and leaving A1 is 3,3 [mg NO₃-N.l⁻¹]; considering that the nitrate measurement is only accurate to within 1,0 [mg NO₃-N.l⁻¹], this may result in errors up to 30 percent in the estimation of μ_{nmT} .

Table 3.2(a) Observed TKN response of the system depicted in Fig.3.1 obtained under constant flow and load for batches of sewage with which satisfactory mass balances were obtained.

DATE	TKN _{AN} [mgN.ℓ ⁻¹]	TKN _{A1} [mgN.ℓ ⁻¹]	TKN _{A2} [mgN.ℓ ⁻¹]	ΔTKN _{AN-A1} [mgN.ℓ ⁻¹]	ΔTKN _{A1-A2} [mgN.ℓ ⁻¹]
31/5	4,6	2,4	1,1	2,2	1,3
3/6	7,7	6,2	2,8	1,5	3,4
5/6	7,3	6,3	1,8	1,0	4,5
6/6	6,0	2,5	1,9	3,5	0,6
7/6	5,6	3,2	2,4	2,4	0,8
12/6	6,3	3,6	1,0	2,7	2,6

Subscript AN refers to the anoxic reactor.

Subscript A1 refers to the first aerobic reactor.

Subscript A2 refers to the second aerobic reactor.

The problem generally with the series system operated under constant flow and load is that the observed response is a *net* effect of the reaction rate plus wash-out rate. The wash-out rate, however, usually has a much greater effect principally because of the requirement of high recycle flows. The masking effect of the wash-out rate makes the estimation of the reaction rate difficult to assess accurately and precisely. One way of inducing a high reaction rate is to impose high influent concentrations, but such high concentrations usually cannot be found in domestic waste waters. Alternatively, the wash-out rate could be reduced by reducing the recycle flow rates but this reduction is possible only to that degree that the effluent nitrate from the anoxic reactor, theoretically does not fall below zero. Because zero nitrate concentration arises also if the denitrification capacity exceeds the nitrate entering the reactor, the concentration must be maintained positive, say, not less than 2 [mgN.ℓ⁻¹].

Table 3.2(b) Observed nitrate response of the system depicted in Fig. 3.1 obtained under constant flow and load conditions for batches with which satisfactory mass balances were obtained.

Sewage Batch No.	DATE	NO_3AN [$\text{mgNO}_3\text{-N.l}^{-1}$]	NO_3Al [$\text{mgNO}_3\text{-N.l}^{-1}$]	ΔNO_3 [$\text{mgNO}_3\text{-N.l}^{-1}$]	State of Nitrogen and COD Mass Balance on Sewage Batch
7	31-5-80	9,9	12,4	2,5	Satisfactory N-Balance=98,4% CODBalance=92,8%
	2-6-80	11,3	14,1	2,8	
	6-6-80	10,6	14,4	3,8	
	7-6-80	12,7	16,6	3,9	
	8-6-80	12,9	16,9	4,0	
8	9-6-80	8,1	11,7	3,6	Satisfactory N-Balance=98,1% CODBalance=94,2%
	10-6-80	9,4	12,8	3,4	
	11-6-80	11,4	14,4	3,0	
	12-6-80	11,4	14,6	3,2	
	13-6-80	9,5	12,5	3,0	
Average:		10,7	14,0	3,3	

The remarks on the previous page illustrate the inadequacies and limitations of the series system under *constant flow and load* to determine kinetic effects accurately. However, the series of experiments did contribute in verifying the limitations of the system and allowed the establishment of reliable experimental procedures.

3.2.2 Predenitrification series system under constant flow and COD load and with an imposed TKN pulse.

In the previous section it was shown that the marked hydraulic effects of the recycle flows led to small relative changes in the measurable nitrification-denitrification process variables (TKN, NO_3).

It was hypothesized that if a high TKN concentration slug was introduced, transient conditions would be set up such that the relative changes of the measurable nitrification-denitrification process variables (TKN, NO_3) would be large enough to effect accurate determination of the nitrification and denitrification constants despite the marked hydraulic effects of the recycle flows. This could be achieved by using a system configuration identical to that used in the constant flow and load experiment (Fig. 3.1) and imposing a cyclic load by feeding a very small flow of high TKN concentration (TKN pulse) once a day over a fixed period of time. The TKN pulse was introduced at a constant rate directly into the anoxic reactor for a period of 4 hours at a fixed time (16.00 hours) each day.

Details of the method of feeding the TKN pulse are as follows: An ammonium chloride stock solution was prepared such that it would effect a known contribution of ammonia to the base load over the 4 hour period. The total mass of TKN entering via the base flow, $Q_b = 17,2 [\text{l.d}^{-1}]$, with a TKN concentration of approximately $50[\text{mgN.l}^{-1}]$ is $17,2 \cdot 50 = 860 [\text{mg N.d}^{-1}]$. It was suggested that if the TKN pulse should contribute an additional load of approximately a third of the daily base TKN load i.e. about $280 [\text{mg N.d}^{-1}]$, transient conditions would be set up such that the observed changes in the TKN and nitrate concentrations between the reactors would be large enough to effect accurate determination of the nitrification and denitrification constants. The contribution of the stock solution over the 4 hour pulse feeding period depends on the pulse feeding rate, V_r . The stock solution was prepared to effect a contribution of 280 mgN over the 4 hour pulse feeding period as follows: 1000 mls. of nitrogen stock solution was prepared to contain $280 \cdot 1000 / V_r = 280 \cdot 1000 / 50,20 [\text{mg N.l}^{-1}]$; $V_r =$ measured TKN pulse flow rate over the 4 hour feeding period = 50,20 [mls/4 hours]. The TKN pulse contribution per hour, ΔTKN , to the base TKN concentration was now given by

$$\Delta\text{TKN} = \frac{280}{4} \cdot \frac{1}{Q_b + \frac{V_r \cdot 4}{1000}} \quad (3.1)$$

where Q_b = Volumetric Base feed flow rate [l.h^{-1}]

but $Q_b \gg V_r \cdot 4/1000$

Hence the TKN pulse contribution *per hour* was given approximately by

$$\Delta \text{TKN} = \frac{280}{4} \cdot \frac{1}{Q_b} \quad (3.2)$$

In the first test (see Fig. 3.2a), the pulse rate was measured to be $V_r = 50,20$ [mls/4 hours] so that Eq. (3.2) could be used directly in calculating the TKN pulse contribution per hour to the base TKN concentration. In a subsequent test a week later the TKN pulse feeding rate was rated at $V_r = 60,0$ [mls/4 hours] so that Eq. (3.2) had to be multiplied by $60,0/50,20$ to account for the increased V_r .

In order to allow adaption of the process to the additional TKN feed, the system was run for a week before monitoring of process variables commenced. Thereafter monitoring was done once a week over 24 hours; each of the process variables (COD, TKN, NO_3 of all the reactors and oxygen uptake rate of each of the aerobic reactors) was measured every 2 hours. This was repeated every week until the process exhibited a stable response pattern. Tables 3.3 (a and b) show the response of two such consecutive weeks, indicating, in particular, that the autotrophic behavioural pattern had achieved an apparent dynamic steady state.

Comparison of experimentally observed results with the simulation, using the general model for a *square wave cyclic* TKN input is shown in Figs. 3.2 (a and b). The appropriate simulated results shown are the outcome of a rather tedious trial and error procedure whereby K_{mpT} , η , f_{ca} and μ_{nmT} were each changed in turn, keeping all other parameters unchanged until the closest possible agreement with experimental observations was effected. The correlation achieved however, is not good, in particular, the experimental nitrate profiles in both Figs. 3.2 (a and b) - the system did not exhibit a consistent cyclic behaviour. This anomalous behaviour

possibly can be ascribed to two effects acting in opposition caused by the TKN pulse. On the one hand the imposed TKN pulse gives rise to high TKN and nitrate concentrations which increases the reliability of their respective differences in the influent and effluent of a reactor. This would increase the reliability of the evaluation of μ_{nmT} . On the other hand, the cyclicity of, say, the nitrate is still contributory to an accumulative error effect associated with the TKN pulse: A slight error in the TKN pulse flow rate of, say, 10 mls in 4 hours (1 ml contains approximately 6 mgN) significantly affects the system's nitrate response; the slightest error in the pumping rate disproportionately distorts the effluent quality from the mean behaviour. Unfortunately the pumping system employed could not guarantee a high accuracy in the flow rate, particularly due to fluctuations in the supplied voltage to the pump.

Reviewing the approach it would appear that despite the theoretical advantages of the system, the practical problem of ensuring extremely accurate measurement of the TKN pulse feeding rate (as well as the recycles' rates) still resulted in unreliable estimates of μ_{nmT} . Furthermore, to fit the simulations to the data was so time consuming that it tended to the point of impracticability. Although feasible, the approach was discarded as not being of a simple enough practical nature for determining μ_{nmT} .

Table 3.3(a) *Observed response of the system in Fig. 3.1 under a constant COD load and a square wave TKN (pulse) load. (Test No. 1) at 14°C, $R_s = 15$ days.*

Time [h]	Anoxic reactor (AN) response		Aerobic reactor (A1) response		Aerobic reactor (A2) response		Time [h]	Ox. uptake rate in A1 [mg O. ℓ^{-1} h $^{-1}$]	Time [h]	Ox. uptake rate in A2 [mg O. ℓ^{-1} h $^{-1}$]
	TKN [mg N. ℓ^{-1}]	NO ₃ [mg N. ℓ^{-1}]	TKN [mg N. ℓ^{-1}]	NO ₃ [mg N. ℓ^{-1}]	TKN [mg N. ℓ^{-1}]	NO ₃ [mg N. ℓ^{-1}]				
14,00	7,8	22,7	4,5	26,8	2,5	28,0	14,2	54,0	14,3	21,2
*16,00	7,7	20,3	3,5	24,6	3,1	26,4	16,2	60,2	16,2	23,7
18,00	22,4	21,1	12,5	26,1	4,1	30,1	18,1	61,8	18,3	49,2
**20,00	30,2	23,3	16,9	28,3	8,1	32,8	20,1	61,7	21,2	43,0
22,00	15,1	25,2	11,8	28,8	7,4	34,4	22,1	58,3	22,7	39,1
24,00	12,3	27,4	8,7	30,2	4,8	34,4	24,1	56,0	24,5	33,3
2,00	9,4	26,5	5,5	32,0	4,1	34,8	2,1	53,7	2,2	25,7
4,00	9,5	26,7	3,6	30,6	3,2	32,8	3,8	55,5	3,8	21,0
6,00	-	25,2	-	28,6	-	30,7	5,5	55,5	5,7	24,8
8,00	7,8	23,6	2,9	27,7	2,7	29,1	7,5	58,4	7,8	22,6
10,00	6,9	21,3	3,1	27,4	3,1	26,5	9,5	49,3	9,7	19,8
12,00	8,4	-	2,7	-	2,5	-	11,5	54,0	11,7	21,2
14,00	7,0	20,4	2,4	23,6	1,3	24,5	13,5	53,3	13,7	20,2

Influent COD = 522,7 [mg COD. ℓ^{-1}] Influent (base) TKN = 55,4 [mg N. ℓ^{-1}] * begin TKN pulse ** end TKN pulse
TKN concentration during pulse feeding period = $55,4 + \frac{280}{4} \cdot \frac{24}{17,2} = 153,1$ [mg N. ℓ^{-1}]

Table 3.3(b) *Observed response of the system in Fig. 3.1 under a constant COD load and a square wave TKN (slug) load (Test No. 2) at 14°C, $R_s = 15$ days.*

Time [h]	Anoxic reactor (AN) response		Aerobic reactor (A1) response		Aerobic reactor (A2) response		Time [h]	Ox. uptake rate in A1 [mg O. ₂ l ⁻¹ h ⁻¹]	Time [h]	Ox. uptake rate in A2 [mg O. ₂ l ⁻¹ h ⁻¹]
	TKN [mg N. l ⁻¹]	NO ₃ [mg N. l ⁻¹]	TKN [mg N. l ⁻¹]	NO ₃ [mg N. l ⁻¹]	TKN [mg N. l ⁻¹]	NO ₃ [mg N. l ⁻¹]				
14,00	7,4	36,3	-	41,4	2,2	43,0	14,2	52,3	14,3	17,4
*16,00	7,8	36,8	3,7	40,5	3,1	42,3	16,1	55,1	16,3	22,1
18,00	16,4	35,7	9,1	40,8	-	43,0	18,1	56,4	18,5	40,8
**20,00	19,3	38,5	14,7	42,2	6,9	47,5	20,1	57,7	20,2	53,2
22,00	12,8	40,7	7,2	43,9	4,4	49,5	22,9	56,6	23,0	40,5
24,00	8,8	40,7	4,4	46,0	3,5	48,3	24,1	56,6	24,3	25,2
2,00	8,1	39,9	-	43,9	-	46,0	2,1	56,3	2,3	21,5
4,00	-	35,5	-	42,2	-	43,5	4,1	55,5	4,4	21,9
6,00	6,9	35,7	-	40,0	4,4	41,5	6,1	54,8	6,2	19,0
8,00	6,3	34,3	5,2	-	-	39,7	8,1	57,2	8,2	19,4
10,00	5,9	31,8	-	37,3	2,7	37,3	10,1	60,2	10,3	18,7
12,00	-	30,0	2,2	34,8	4,3	36,0	12,1	52,5	12,4	20,6
14,00	8,4	29,8	-	33,6	3,7	35,0	14,4	49,8	14,8	21,5

Influent COD = 467,6 Influent (base) TKN = 57,0 [mg N. l⁻¹] * begin TKN pulse ** end TKN pulse
TKN influent concentration during pulse feeding period = $57,0 + \frac{280}{4} \cdot \frac{60,0}{50,20} \cdot \frac{24}{17,2} = 173,7$ [mg N. l⁻¹]

Table 3.3(b) *Observed response of the system in Fig. 3.1 under a constant COD load and a square wave TKN (slug) load (Test No. 2) at 14°C, $R_s = 15$ days.*

Time [h]	Anoxic reactor (AN) response		Aerobic reactor (A1) response		Aerobic reactor (A2) response		Time [h]	Ox. uptake rate in A1 [mg O. ℓ^{-1} h $^{-1}$]	Time [h]	Ox. uptake rate in A2 [mg O. ℓ^{-1} h $^{-1}$]
	TKN [mg N. ℓ^{-1}]	NO ₃ [mg N. ℓ^{-1}]	TKN [mg N. ℓ^{-1}]	NO ₃ [mg N. ℓ^{-1}]	TKN [mg N. ℓ^{-1}]	NO ₃ [mg N. ℓ^{-1}]				
14,00	7,4	36,3	-	41,4	2,2	43,0	14,2	52,3	14,3	17,4
*16,00	7,8	36,8	3,7	40,5	3,1	42,3	16,1	55,1	16,3	22,1
18,00	16,4	35,7	9,1	40,8	-	43,0	18,1	56,4	18,5	40,8
**20,00	19,3	38,5	14,7	42,2	6,9	47,5	20,1	57,7	20,2	53,2
22,00	12,8	40,7	7,2	43,9	4,4	49,5	22,9	56,6	23,0	40,5
24,00	8,8	40,7	4,4	46,0	3,5	48,3	24,1	56,6	24,3	25,2
2,00	8,1	39,9	-	43,9	-	46,0	2,1	56,3	2,3	21,5
4,00	-	35,5	-	42,2	-	43,5	4,1	55,5	4,4	21,9
6,00	6,9	35,7	-	40,0	4,4	41,5	6,1	54,8	6,2	19,0
8,00	6,3	34,3	5,2	-	-	39,7	8,1	57,2	8,2	19,4
10,00	5,9	31,8	-	37,3	2,7	37,3	10,1	60,2	10,3	18,7
12,00	-	30,0	2,2	34,8	4,3	36,0	12,1	52,5	12,4	20,6
14,00	8,4	29,8	-	33,6	3,7	35,0	14,4	49,8	14,8	21,5

Influent COD = 467,6 Influent (base) TKN = 57,0 [mg N. ℓ^{-1}] * begin TKN pulse ** end TKN pulse
TKN influent concentration during pulse feeding period = $57,0 + \frac{280}{4} \cdot \frac{60,0}{50,20} \cdot \frac{24}{17,2} = 173,7$ [mg N. ℓ^{-1}]

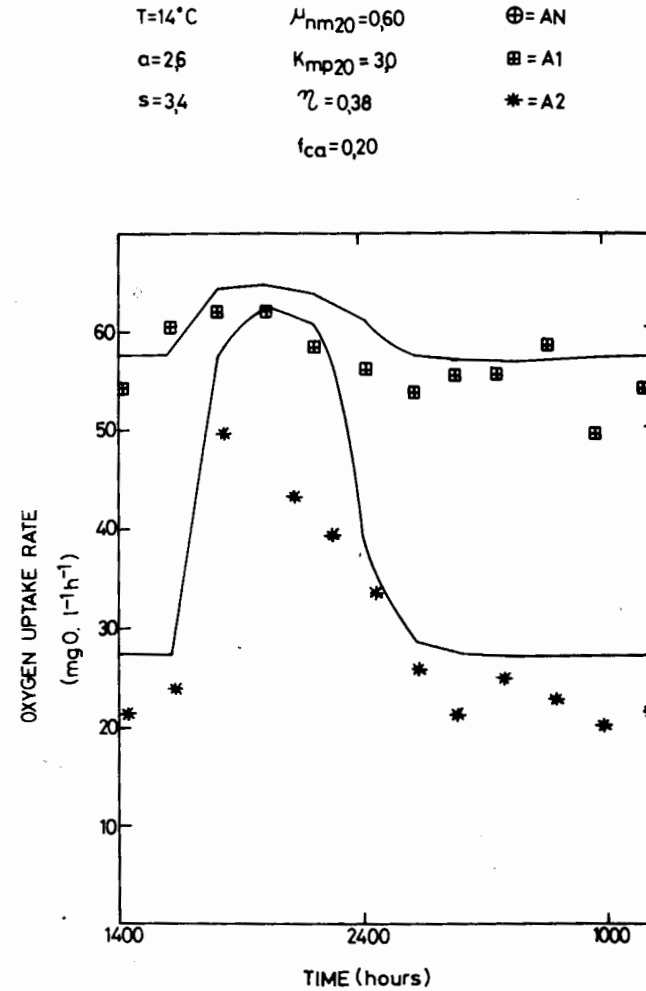
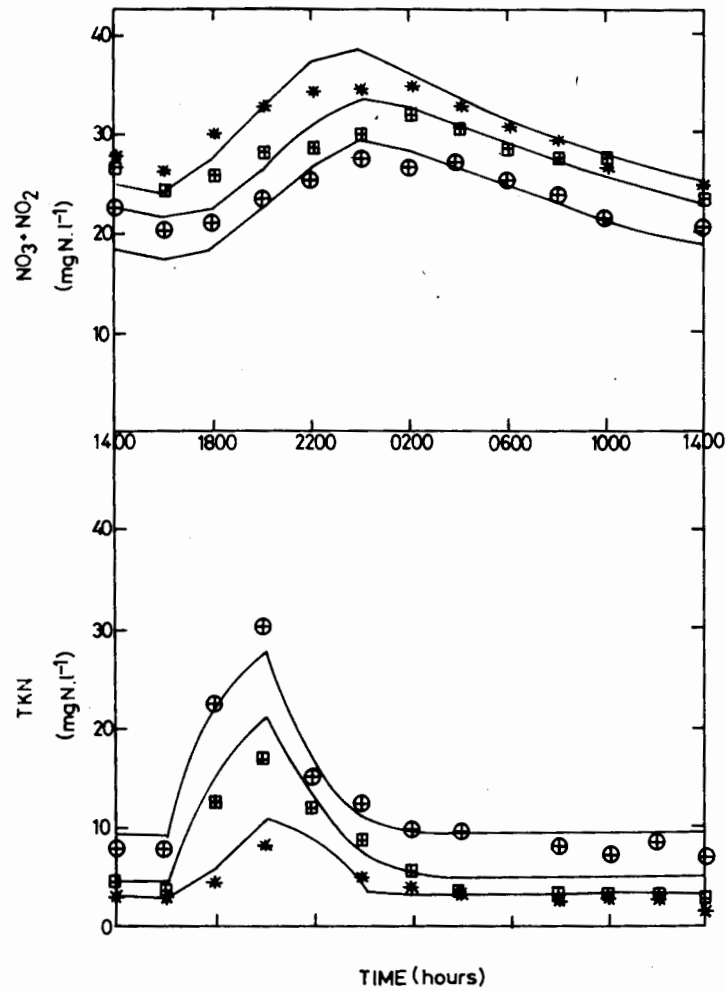


Fig.3.2a: Closest fit of the observed response of the series system in Fig.3.1 under a constant COD load and with a square wave TKN load at 14°C , $R_s = 15$ days (Experiment No. 1).

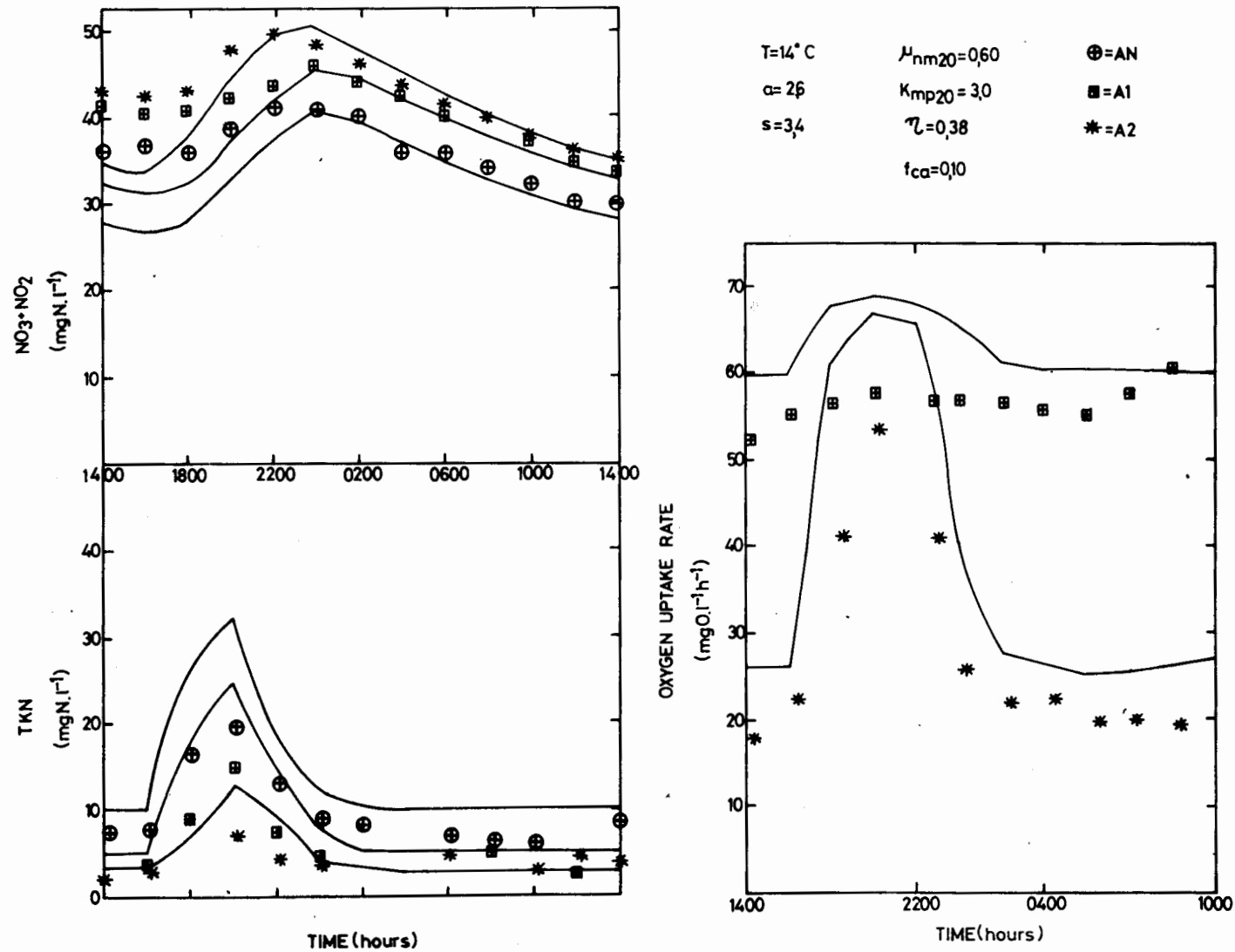


Fig.3.2b: Closest fit of the observed response of the series system in Fig.3.1 under a constant COD load and with a square wave load of TKN at 14°C, $R_s=15$ days (Experiment No. 2).

3.3. Completely Mixed Single Reactor (CMSR) System.

3.3.1 Estimation of reaction rates in CMSR system

The observed rate of change in any system is the *net* effect of the rate of reaction together with the hydraulic washout rate. In a CMSR the hydraulic effect can be easily accounted for, thereby allowing determination of the nitrification and denitrification *reaction rates* and, hence, the associated kinetic constants :

Van Haandel and Marais (1981) showed that over the interval $(t_2 - t_1)$ the average reaction rate of concentration C in a reactor j , $r_{C_{jr}}$ is given by

$$r_{C_{jr}} = (C_{jt2} - C_{jt1}) / (t_2 - t_1) - (\bar{C}_{ij} - \bar{C}_j) / R_j \quad (3.3)$$

where

C_{jt2} = Concentration of C in reactor j at time t_2

C_{jt1} = Concentration of C in reactor j at time t_1

$$\bar{C}_j = (C_{jt1} + C_{jt2}) / 2$$

\bar{C}_{ij} = Average concentration of C in the influent to reactor j over the interval $(t_2 - t_1)$.

R_j = Actual hydraulic retention time in reactor j during the interval $(t_2 - t_1)$

Considering the nitrification rate ($C = \text{NO}_3$) in a CMSR system ($j=1$), with feed that is free of nitrate ($\bar{C}_{i1} = 0$), Eq (3.3) may be reduced to

$$r[\text{NO}_3]_r = ([\text{NO}_3]_{t2} - [\text{NO}_3]_{t1}) / (t_2 - t_1) + [\bar{\text{NO}}_3] / R_h \quad (3.4)$$

where

$$[\bar{\text{NO}}_3] = ([\text{NO}_3]_{t1} + [\text{NO}_3]_{t2}) / 2$$

R_h = Actual hydraulic retention time in the CMSR during the interval $(t_2 - t_1)$

Equation (3.4) is used to evaluate the average nitrification reaction rate from the nitrate-time response curve observed on a CMSR system during an aerobic period.

Considering the denitrification rate during an anoxic period over any time interval $(t_2 - t_1)$, the average denitrification reaction rate is obtained from the nitrate-time response curve as follows:

Equation (3.4) is rewritten

$$-r[\text{NO}_3]_r = -([\text{NO}_3]_{t_2} - [\text{NO}_3]_{t_1}) / (t_2 - t_1) - [\overline{\text{NO}_3}] / R_h$$

i.e.

$$-r[\text{NO}_3]_r = ([\text{NO}_3]_{t_1} - [\text{NO}_3]_{t_2}) / (t_2 - t_1) - [\overline{\text{NO}_3}] / R_h \quad (3.5)$$

From the development above these equations are valid for any flow condition, constant or varying with time, provided that during variable flow the value of R_h over the interval $(t_2 - t_1)$ is available.

3.3.2. Alternating anoxic-aerobic CMSR (AAACMSR) system under square wave flow and load

In Section 3.1 it was hypothesized that when a square wave flow and load pattern is imposed on an alternating anoxic-aerobic CMSR (AAACMSR) system, it should be possible to determine *all* the constants affecting nitrification and denitrification i.e. μ_{nmT} , f_{ca} , K_{mpT} and η . To test this, an experimental set up identical to that given in Fig 3.3 was employed with operational and sewage characteristics as listed in Table 3.4. A daily cyclic square wave input flow (12 h feed, 12 h no feed) was imposed with a single anoxic period of 2 h duration each day during the high flow period. This was done by switching the electrical current, to an air pump, on and off. The plant was operated in this fashion until, apparently, steady state conditions were achieved. During this period the VSS and mean effluent TKN were checked daily; it was presumed that when these showed no change, steady state had been achieved.

Once steady state had been achieved, intensive testing was done over that fraction of anoxic-aerobic period during which rapid changes of concentrations of nitrate, alkalinity, TKN and NH_3 took place. Samples were taken every hour and the following process variables monitored: Oxygen uptake rate, COD, VSS, TKN, NH_3 , NO_3 and Alk.

Two experiments were carried out under square wave flow and load on the AAACMSR system using the *same* sewage batch. In the first experiment (EST 31), the 2 h anoxic period was introduced 1 hour after the beginning of the feeding period; the observed response is listed in Table 3.5a. In the second experiment (EST 32), the 2 h anoxic period was introduced 2 hours after the beginning of the feeding period; the observed response is listed in Table 3.5b. This data provided the basis for inquiring into the determination, by manual means, of the kinetic nitrification and denitrification constants.

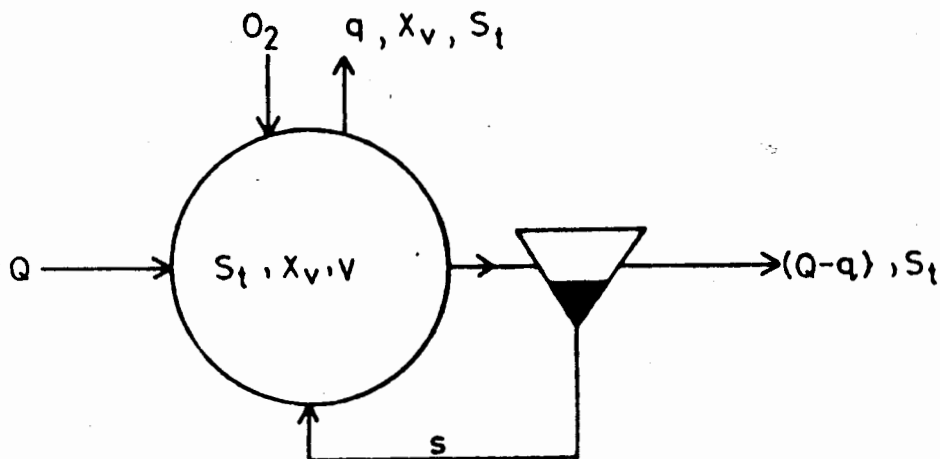


Fig.3.3: Completely mixed single reactor (CMSR) system used in the determination of nitrification and denitrification constants by imposing alternating anoxic-aerobic conditions.

Table 3.4 Operational and average sewage characteristics used in Fig. 3.3 operated under daily square wave conditions with a single anoxic period of 2 hours in 24.

<u>Operational Characteristics</u>	
$T = 14^{\circ}\text{C}$	
Reactor volume, $V = 15 \text{ [l]}$	
Feeding period, $t_f =$ Non feeding period = 12 [h]	
Hydraulic retention time during feeding period = 6 [h]	
Sludge age, $R_h = 15 \text{ [d]}$	
Duration of stable*operation before testing commenced = 14 [d]	
<u>Sewage Characteristics</u>	
	<u>Average**</u>
Influent COD = 468 to 572 [mg COD.l ⁻¹]	507 [mg COD.l ⁻¹]
Influent TKN = 48 to 51 [mg N.l ⁻¹]	49 [mg N.l ⁻¹]
$f_{ca} = 0,10 \text{ [mg COD.mg COD}^{-1}]$ (determined from an independent experimental setup)	
$f_{up} = 0,09 \text{ [mg VSS.mg COD}^{-1}]$	
$f_{us} = 0,12 \text{ [mg COD.mg COD}^{-1}]$	
$f_{na} = 0,64 \text{ [mg NH}_3\text{-N.mg N}^{-1}]$	
$f_{nu} = 0,0 \text{ [mg N.mg N}^{-1}]$	

* Stable operation characteristic to steady state conditions is indicated by a constant VSS and consistent nitrification with a corresponding constant mean effluent TKN.

** Disregarding any outliers.

Table 3.5a *Observed response of a CMSR system under square wave flow and load (experiment EST 31)*
at 14°C, $R_s = 15$ days, pH = 7,2

SAMPLE	TIME [h]	NH ₃ [mgN.ℓ ⁻¹]	TKN [mgN.ℓ ⁻¹]	NO ₃ [mgN.ℓ ⁻¹]	ALKALINITY [mgCaCO ₃ .ℓ ⁻¹]	COD [mgCOD.ℓ ⁻¹]	VSS [mgVSS.ℓ ⁻¹]	TIME [h]	O.U.R. [mgO.ℓ ⁻¹ .h ⁻¹]
Influent		27,0	51,9	0,0	413,0	503,0	-	-	-
Effluent		2,51	6,2	23,8	213,0	59,8	-	-	-
1	6,00	0,41	5,5	29,5	185,4	-	3488,0	6,1	15,3
2	8,00*	0,10	5,5	30,0	183,3	56,2	-	7,0	16,5
3	9,00***	1,1	5,3	30,0	187,5	76,6	3351,0	8,1	39,4
4	10,00	4,6	10,5	20,4	222,0	-	3346,0	9,0	46,5
5	11,00****	8,3	14,0	15,7	252,4	73,3	3394,0	11,1	47,8
6	13,00	5,1	6,4	19,2	230,2	-	3351,0	12,1	55,3
7	16,00	0,5	4,8	27,7	185,4	70,4	3395,0	13,1	53,0
8	19,00	0,4	-	28,3	181,3	-	-	14,6	52,6
9	20,00**	0,2	5,2	-	181,3	55,0	3560,0	16,1	43,8
10	21,00	0,17	3,4	28,35	180,2	68,4	3665,0	18,0	43,8
11	22,00	0,05	2,4	28,4	174,1	59,5	3644,0	19,1	43,8
								20,2	44,4
								20,3	25,1
								20,5	21,3
								20,7	19,5
								22,0	16,0

* begin feeding period
 ** end feeding period
 *** begin anoxic period
 **** end anoxic period

Table 3.5b *Observed response of a CMSR system under square wave flow and load (experiment EST 32)*
at 14°C, R_s = 15 days, pH = 7,2

SAMPLE	TIME [h]	NH ₃ [mgN.l ⁻¹]	TKN [mgN.l ⁻¹]	NO ₃ [mgN.l ⁻¹]	ALKALINITY [mgCaCO ₃ .l ⁻¹]	COD [mgCOD.l ⁻¹]	VSS [mgVSS.l ⁻¹]	TIME [h]	O.U.R. [mgO.l ⁻¹ .h ⁻¹]
Influent		32,6	51,2	0,0	412,0	492,8			
Effluent		3,4	6,7	25,0	208,1	69,0			
1	7,00	0,37	-	29,5	173,6	50,7	3642,0	7,1	13,4
2	8,00*	0,28	3,1	30,6	173,6	-	-	8,0	28,8
3	9,00	0,69	3,6	30,3	172,0	-	3393,0	8,4	38,8
4	10,00***	-	2,9	30,3	-	52,7	3482,0	8,7	46,6
5	11,00	5,82	9,2	22,8	210,1	62,9	3419,0	8,9	46,7
6	12,00****	9,42	12,6	16,3	247,2	69,0	3472,0	9,1	44,5
7	13,00	8,15	-	17,8	240,5	64,9	3366,0	9,4	42,5
8	14,00	5,63	5,6	20,7	225,1	64,9	3429,0	10,0	41,7
9	16,00	1,34	4,8	25,5	191,1	-	3451,0	12,1	50,8
10	19,00	0,79	3,5	28,8	177,2	-	-	13,1	59,4
11	20,00**	0,51	3,4	29,1	173,0	64,9	3502,0	14,1	53,3
12	21,00	1,07	3,2	29,4	172,5	-	3518,0	16,2	54,2
13	22,00	0,42	4,6	-	175,6	-	-	19,1	44,3
								20,1	40,8
								20,4	18,0
								21,2	16,5

* begin feeding period
 ** end feeding period
 *** begin anoxic period
 **** end anoxic period

3.3.2a Manual determination of μ_{nmT} , f_{ca} , K_{mpT} and η from a CMSR system under a square wave flow feeding pattern

One important object of a CMSR system operated under a square wave input pattern, and subjected to alternating anoxic-aerobic conditions during the high flow period, was to check if it is possible to estimate all the constants that significantly affect nitrification and denitrification *without* the aid of a computer. It is now the intention to describe the procedures by means of which such estimates can be made. The estimation of each of the constants from the experimental response of experiment EST 32 will be dealt with separately.

3.3.2a(I) Estimation of μ_{nmT}

The maximum nitrification rate constant, μ_{nmT} , can be readily estimated from the nitrate profile of the aeration period subsequent to the anoxic period. During this period it can be accepted that the ammonia concentration remains so high (above 2 [mg N.l⁻¹]) that the nitrification rate constant μ_{nT} will be maintained at its maximum value, μ_{nmT} , given by

$$\mu_{nmT} = Y_n (dN_n/dt)/X_n$$

To evaluate μ_{nmT} the value of X_n needs to be calculated: it was shown in Section 2.2.3 that in order to calculate X_n the values of Y_n and b_{nT} must be known. Accepting the standard values for Y_n and b_{nT} (i.e. $b_{n20} = 0,04[d^{-1}]$, $Y_n = 0,10 [mg VSS.mg N^{-1}]$), a theoretical average value for X_n may be found from

$$\bar{X}_n = Y_n R_s \Delta TKN / [(1 + b_{nT} R_s) \cdot R_h] \quad (3.6)$$

where

ΔTKN = Average nitrified TKN [mg N.l⁻¹]

i.e.

$$\Delta TKN = TKN_{IN} - TKN_{EFF} - N_s \quad (3.7)$$

where

TKN_{IN} = Average influent TKN concentration [mg N.l⁻¹]

TKN_{EFF} = Average effluent TKN concentration [mg N.l⁻¹]

N_s = Steady state heterotrophic nutrient nitrogen requirement [mg N.l⁻¹]

and

$$N_s = \{f_n[Y_h(1+f_{b_h}R_s)(1-f_{us}-P.f_{up})/(1+b_hR_s)] + f_{up}.f_n\}.S_{ti}$$

These equations formed the basis for determining the value of μ_{nmT} .

In this experimental series evaluation of μ_{nmT} was based on the response of a CMSR system under square wave flow and load (12h on, 12h off) subjected to anoxic conditions 2 hours after the beginning of the feeding period, for a period of 2 hours (experiment EST 32). The observed nitrate concentrations were plotted in Fig 3.4 and a smooth curve drawn through the data points. Values for $[NO_3]_{obs}$ were read off the smooth curve. Table 3.6 was then constructed by estimating the nitrification and denitrification rates, $r[NO_3]_r$ and $-r[NO_3]_r$, respectively from $r[NO_3]_{obs}$, and taking cognisance of the hydraulic rate, $r[NO_3]_h$, as described in Section 3.3.1. From Table 3.6, the average nitrification rate subsequent to the anoxic period, during which the ammonia concentration was above $2[mg\ N.\ell^{-1}]$, can be calculated

$$\overline{r[NO_3]}_r = (5,4+5,9+5,8)/3 = 5,7 [mg\ N.\ell^{-1}.h^{-1}]$$

From Eq (3.7),

$$\Delta TKN = TKN_{IN} - TKN_{EFF} - N_s = 49-6,5-11,3 = 31,2$$

From Eq (3.6),

$$\overline{X}_n = Y_n R_s \Delta TKN / [(1+b_n R_s).R_h] = 62 [mg\ VSS.\ell^{-1}]$$

hence,

$$\begin{aligned}\mu_{nm14} &= (\overline{r[NO_3]}_r . Y_n) / X_n \\ &= 5,7 . 0,1.24 / 62 = 0,22[d^{-1}]\end{aligned}$$

Accepting the temperature dependency equation, i.e.

$$\mu_{nmT} = \mu_{nm20} (1,123)^{T-20}$$

hence,

$$\mu_{nm20} = 0,44[d^{-1}].$$

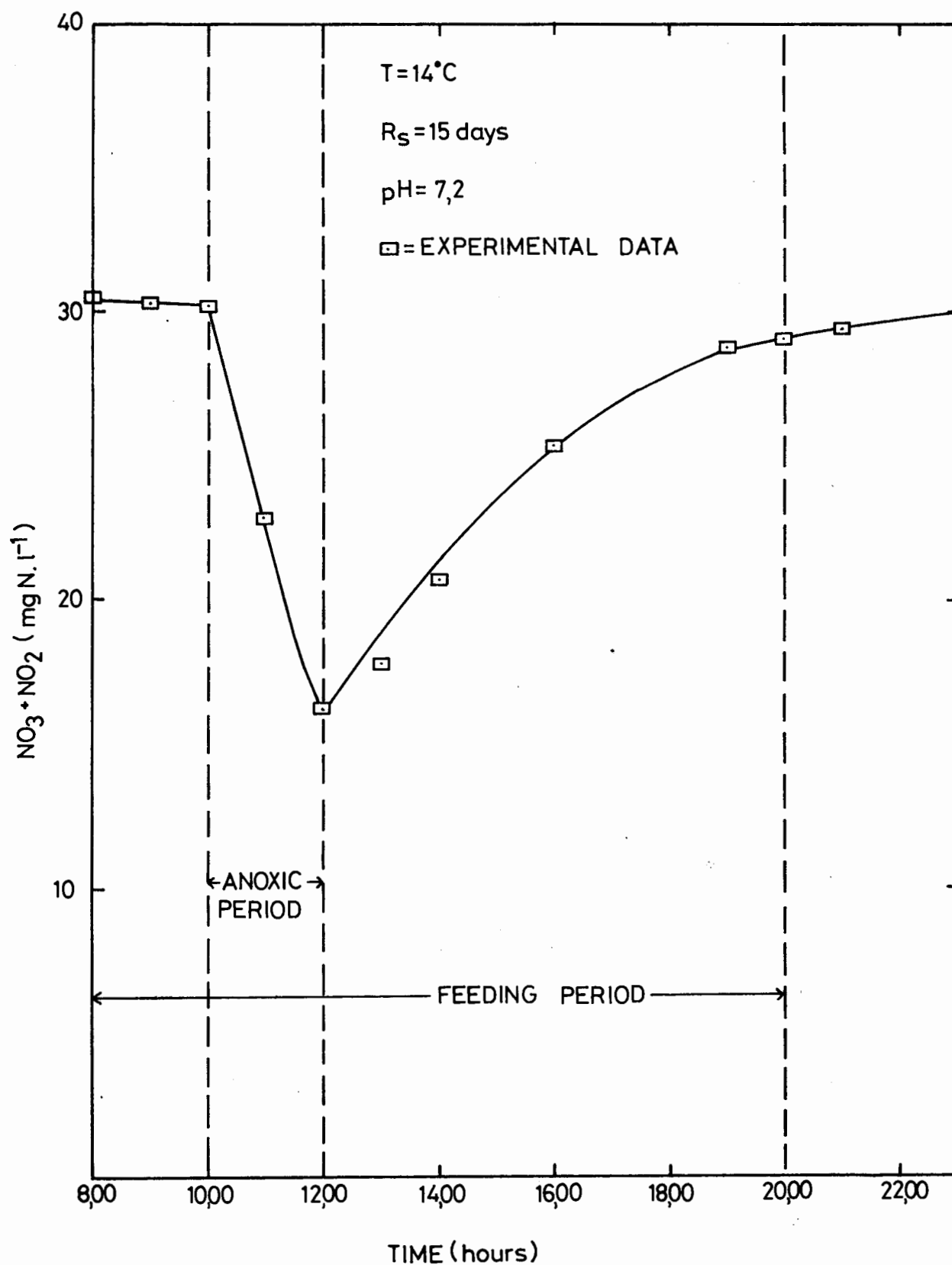


Fig.3.4: Smoothing of the observed nitrate data from experiment EST32.

Table 3.6 Calculated nitrification and denitrification reaction rates from a smooth curve of the observed nitrate response.

Time [h]	NO ₃ [mg N.ℓ ⁻¹]	Δt [h]	r[NO ₃] _{obs} [mg N.ℓ ⁻¹ h ⁻¹]	r[NO ₃] _h [mg N.ℓ ⁻¹ h ⁻¹]	r[NO ₃] _r [mg N.ℓ ⁻¹ h ⁻¹]
8,00*	30,3	1	0,0	5,07	5,1
9,00	30,3	1	0,0	5,07	5,1
10,00***	30,3	1	7,8	4,4	3,4
11,00	22,5	1	6,2	3,23	3,0
12,00****	16,3	1	2,5	2,93	5,4
13,00	18,8	1	2,6	3,35	5,9
14,00	21,4	2	1,95	3,89	5,8
16,00	25,3	3	1,1	4,5	5,6
19,00	28,6	1	0,5	4,81	5,3
20,00**	29,1	1	0,40	0,0	0,4
21,00	29,5				

*. start feed period

*** start anoxic period

** end feed period

**** end anoxic period

3.3.2a(II) Estimation of f_{ca}

The easily biodegradable COD fraction of the influent, f_{ca} , was estimated from the precipitous drop in oxygen uptake rate at feed cessation as follows:

The total drop in oxygen uptake rate during feed cessation, ΔO_t is given by

$$\Delta O_t = \Delta O_{sbs} + \Delta O_n \quad (3.8)$$

where

ΔO_n = Drop in oxygen uptake rate associated with termination of nitrogen available for nitrification entering with the feed [$\text{mg O.l}^{-1}.\text{h}^{-1}$].

ΔO_{sbs} = Drop in oxygen uptake rate associated with termination of easily biodegradable COD entering with the feed [$\text{mg O.l}^{-1}.\text{h}^{-1}$].

Theoretically f_{ca} is given by

$$f_{ca} = (\Delta O_{sbs} \cdot R_a) / [(1 - P Y_h) \cdot S_{bi}] \quad (3.9)$$

where

R_a = Actual influent hydraulic retention time during the feed period [h].

$S_{bi} = S_{ti}(1 - f_{us} - P \cdot f_{up})$ = Total biodegradable influent COD concentration on the day experiment was conducted [mg COD.l^{-1}]

From Table 3.5b the experimental values of the oxygen uptake rate (O.U.R) are plotted (in Fig 3.5) and a smooth curve is drawn through these points. The average O.U.R. before feed termination is 42,5 [$\text{mg O.l}^{-1}.\text{h}^{-1}$] and the average O.U.R. just after feed termination is 17,0 [$\text{mg O.l}^{-1}.\text{h}^{-1}$]. Hence the drop in O.U.R. is

$$\Delta O_t = 42,5 - 17,0 = 25,5 \text{ [mg O.l}^{-1}.\text{h}^{-1}\text{]}.$$

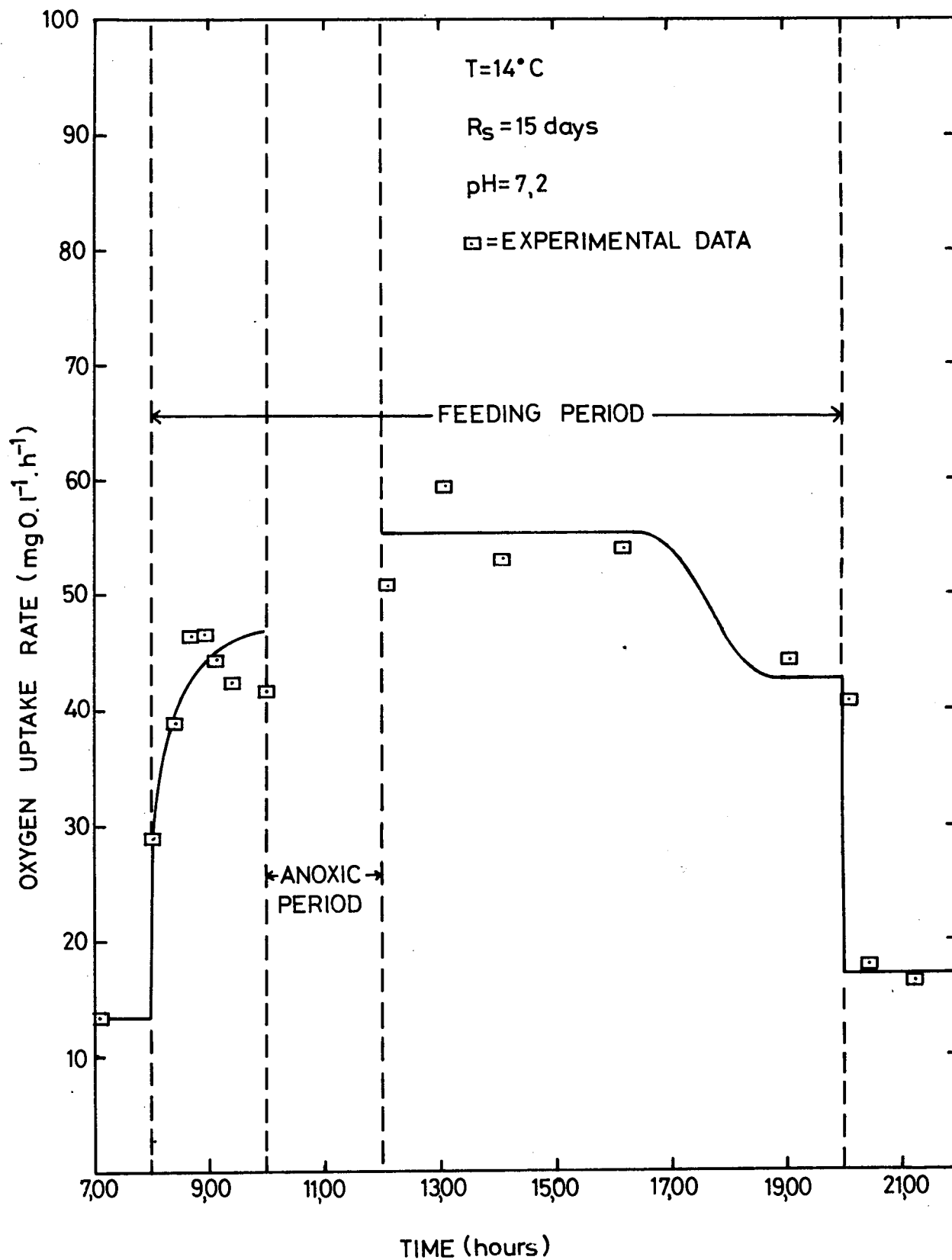


Fig. 3.5: Smoothing of the observed oxygen uptake rate data from experiment EST32.

From Section 3.3.2a(I) the average nitrification rate before feeding termination was estimated to be $5,6 \text{ [mg NO}_3\text{-N.l}^{-1}\text{.h}^{-1}\text{]}$, and just after feed termination, at $0,4 \text{ [mg NO}_3\text{-N.l}^{-1}\text{.h}^{-1}\text{]}$ (see Table 3.6). Hence the decrease in the nitrification reaction rate is $5,6 - 0,4 = 5,2 \text{ [mg N.l}^{-1}\text{.h}^{-1}\text{]}$ corresponding to a decrease in O.U.R. of

$$\Delta O_n = 5,2 \cdot 4,57 = 23,8 \text{ [mg O.l}^{-1}\text{.h}^{-1}\text{]}.$$

From Eq (3.8), the drop in O.U.R. associated with the easily biodegradable COD is

$$\Delta O_{sbs} = \Delta O_t - \Delta O_n = 25,5 - 23,8 = 1,7 \text{ [mg O.l}^{-1}\text{.h}^{-1}\text{]}.$$

Substituting in Eq (3.9), i.e:

$$f_{ca} = (1,7 \cdot 6) / [(1 - 1,54 \cdot 0,45) \cdot 493 \cdot (1 - 0,12 - 1,54 \cdot 0,09)], \text{ yielding,}$$

$$f_{ca} = 0,09 \text{ [mg COD.mg COD}^{-1}\text{]}. \text{ This value is designated the "indirect" estimation of } f_{ca}.$$

A direct, and independent, estimation of f_{ca} , using a short sludge age completely mixed aerobic reactor, as described in Section 2.3.8, yields $f_{ca} = 0,10 \text{ [mg COD.mg COD}^{-1}\text{]}$.

Comparing the values of f_{ca} found by the two procedures above, the estimates differ slightly. The direct measure of f_{ca} ($f_{ca} = 0,1$) from experience is consistent and reproducible and hence can be considered reliable. The indirect method, in contrast, is heavily dependent on the stability of nitrification and oxygen uptake rates. From Table 3.5b it is apparent that in experiment EST 32 the value of O.U.R. before feed termination fluctuated in value up to $4,3 \text{ [mg O.l}^{-1}\text{.h}^{-1}\text{]}$ from the average. From Table 3.6 it is apparent that in experiment EST 32 the value of the nitrification rate before feed termination fluctuated in value up to $0,3 \text{ [mg N.l}^{-1}\text{.h}^{-1}\text{]}$ from the average, which is equivalent to $0,3 \cdot 4,57 = 1,4 \text{ [mg O.l}^{-1}\text{.h}^{-1}\text{]}$. The fluctuations in the nitrification rate values together with the large fluctuations in the O.U.R. values, lead to uncertainty in the estimation of ΔO_{sbs} at feed cessation which ultimately leads to an uncertain estimation of f_{ca} .

O_{sbp} = Oxygen uptake rate associated with utilization of slowly biodegradable COD [$\text{mg COD.l}^{-1}.\text{h}^{-1}$]

K_{spT} = Half saturation value for slowly biodegradable COD [$\text{mg COD.mg VSS}^{-1}$]

Evaluation of K_{mpT} at a particular instance requires the evaluation of O_{sbp} , X_a and the Levenspiel ratio, i.e.

$$X_s P / (X_s P + X_a \cdot K_{spT}).$$

With the exception of O_{sbp} which can be determined from experimental observation with its associated experimental error, all the other parameters (X_a and the Levenspiel ratio) must be estimated, so that attention needs to be given to the errors induced in the estimation of K_{mpT} due to errors in the estimates of these parameters.

Determination of O_{sbp}

In the square wave loaded anoxic-aerobic alternating CMSR system the value of O_{sbp} can be found from the O.U.R. profile just after feed cessation. The O.U.R. in this region is due to two oxygen demand rates:

- (1) Rate of utilization of slowly biodegradable particulate COD,

S_{bp} ;

O.U.R. due to carbonaceous metabolism subsequent to feed termination, in terms of the death regeneration approach, is virtually solely due to the slowly biodegradable particulate COD, S_{bp} . The easily biodegradable soluble COD, S_{bs} , effectively terminates the moment the influent source of S_{bs} terminates.

- (2) Rate of nitrification of ammonia derived from ammonification of TKN released by cell death and lysis;

at long sludge ages the nitrification rate is sufficiently high that immediately subsequent to feed termination the ammonia

derived from ammonia in the influent is effectively reduced to zero. The only remaining source of ammonia is that of ammonified TKN generated by death and lysis less ammonia needed for cell regeneration.

Comparing the relative magnitudes of these rates subsequent to feed termination, that due to S_{bp} is the overwhelmingly greater. From simulated response using the general model for 15 days sludge age at 14°C , the O.U.R. due to nitrification subsequent to feed termination is of the order of 2 to 4% of that due to S_{bp} and can be neglected for all practical purposes*. Consequently the O.U.R. subsequent to feed termination can be accepted to be wholly due to S_{bp} utilization.

Estimation of X_a

The value of X_a from dynamic steady state computer simulation (see Section 3.3.2b) of square wave conditions for experiment EST 32, with $f_{ca} = 0,10$ [mg COD.mg COD $^{-1}$], $\mu_{nm20} = 0,48$ [d $^{-1}$], $K_{mp20} = 2,3$ [mg COD.mg VSS $^{-1}$.d $^{-1}$] and $\eta = 0,38$ varies from about 1262 to 1310 [mg VSS.l $^{-1}$] with an average of 1285 [mg VSS.l $^{-1}$]. The steady state value of X_a under constant flow and load is given by inserting values into the following Eq. (3.11)

$$X_a = Y_h S_{bi} R_s / [(1 + b_h R_s) \cdot R_h] \quad (3.11)$$

Inserting the relevant S_{bi} , R_s and R_h values into Eq. (3.11), the steady state constant flow and load value of X_a for the *average* loading conditions of experiment EST 32 is

$$X_a = 0,45.375,9.15 / [(1 + 0,202.15) \cdot 0,5]$$

i.e.

$$X_a = 1259 \text{ [mg VSS.l}^{-1}\text{]}$$

the error involved in accepting a steady state constant flow and load X_a (1259 [mg VSS.l $^{-1}$]), against the peak X_a (1310 [mg VSS.l $^{-1}$]),

* The errors in the experimentally measured total O.U.R. are at least of the same order of magnitude as the O.U.R. due to nitrification subsequent to feed termination, hence, an attempt to correct for this nitrification is meaningless.

or the square wave average X_a ($1285[\text{mg VSS} \cdot \text{l}^{-1}]$), is small. Hence the steady state constant flow and load X_a is assumed in all further manual calculations.

Evaluation of the Levenspiel ratio and hence K_{mpT} .

It has been shown that O_{sbp} can be experimentally determined and that a steady state constant flow and load approximation for X_a can be accepted. It follows from Eq. (3.10) that the precision in the estimation of K_{mpT} depends to a large extent on the estimation of the Levenspiel ratio ($X_s.P/[X_s.P+X_a.K_{spT}]$) at the moment when K_{mpT} is evaluated, i.e. just prior to the instance of feed cessation in a square wave cycle, which in turn depends on the estimation of X_s just prior to the instance of feed cessation. From dynamic steady state computer simulations (see Section 3.3.2b), just before feed termination X_s is at its maximum value; unfortunately this peak X_s , X_{sp} , cannot be determined manually. Guide lines on the best procedure to evaluate K_{mpT} can be obtained from a simulation based investigation of the way the peak X_s and its corresponding Levenspiel ratio changes for an expected range of K_{mpT} values.

In Table 3.7 is listed a range of Levenspiel ratios corresponding to the widest range of K_{mpT} values encountered by Ekama and Marais (1978) and Van Haandel and Marais (1981), $K_{mp20} = 2,2$ to $3,0$ [$\text{mg COD} \cdot \text{mg VSS}^{-1} \cdot \text{d}^{-1}$], for the operational conditions employed in experiment EST 32.

Table 3.7 Possible variations of the Levenspiel ratio due to possible variations of peak X_s in experiment EST 32.

K_{mp20} [$\text{mg COD} \cdot \text{mg VSS}^{-1} \cdot \text{d}^{-1}$]	X_{sp} [$\text{mg VSS} \cdot \text{l}^{-1}$]	$X_{sp}.P/[X_{sp}.P+X_a.K_{spT}]$
2,2	436	0,89
2,3	370	0,87
2,5	298	0,84
2,7	254	0,82
3,0	208	0,79

From Table 3.7, if an average Levenspiel ratio of 0.84 is accepted in estimating K_{mp20} for experiment EST 32, the maximum theoretical error in the estimation of K_{mp20} is 5 per cent, i.e. if estimates of the Levenspiel ratios under the specific operating conditions are possible (by means of a computer), good estimates for K_{mpT} can be calculated and the theoretical errors made in accepting these estimates can be determined.

In the absence of computer facilities the Levenspiel ratio can be assumed to be a unity, in which event Eq. (3.10) is reduced to

$$K_{mpT} = O_{sbp} / [(1 - PY_h) \cdot X_a] \quad (3.12)$$

This condition will be closely approximated if a high X_{sp} is ensured such that $X_{sp} \cdot P \gg X_a \cdot K_{spT}$, in which event the estimation of K_{mpT} does *not* require knowledge of the magnitude of X_s .

One way of maintaining a high X_s is to have *repeated* anoxic intervals (e.g. 2 hours anoxic period followed by 4 hours aerobic period), instead of the single 2h anoxic period employed in this experiment. If X_s is maintained high by imposing the appropriate conditions, manual estimation of K_{mpT} is possible and the reliability of its magnitude is subject only to experimental errors related to the O.U.R. measurement. The experimental design in this investigation therefore was inadequate to determine K_{mpT} reliably without the aid of computer simulations.

3.3.2a(IV) Estimation of η

The ratio of slowly biodegradable utilization rate in an anoxic environment to that in an aerobic environment, $\eta = K'_{mpT} / K_{mpT}$ can be determined as follows:

$$\eta = \frac{\Delta N_{sbp} \cdot 2.86}{(1 - PY_h) \cdot X_a \cdot K_{mpT}} \quad (3.13)$$

where

ΔN_{sbp} = Nitrate removal associated with the utilization of slowly biodegradable COD.

The magnitude of ΔN_{sbp} is given by

$$\Delta N_{sbp} = \Delta N - \Delta N_{sbs} \quad (3.14)$$

where

ΔN = Total daily nitrate removal [mg N.d⁻¹]

ΔN_{sbs} = Daily nitrate removal due to the utilization of the easily biodegradable material [mg N.d⁻¹]

Assuming complete utilization of the easily biodegradable material, ΔN_{sbs} is given by

$$\Delta N_{sbs} = Q_a \cdot t_a \cdot S_{bi} \cdot f_{ca} \cdot (1 - P_{Y_h}) / 2.86 \quad (3.15)$$

where

Q_a = Actual influent volumetric flow rate during the loading period [l.h⁻¹]

t_a = Duration of the anoxic period [h]

S_{bi} = Total biodegradable influent COD concentration on the day testing was conducted [mg COD.l⁻¹]

The certainty in the estimation of η depends primarily on the certainty with which K_{mpT} is estimated. This uncertainty is further increased by (1) any inaccuracies in the nitrate measurements during the anoxic period and (2) an inaccurate estimation of f_{ca} .

In this investigation the determination of K_{mpT} required computer simulations hence the determination of η also was not possible without this aid.

3.3.2b Determination of the values of all the nitrification and denitrification constants from an AAACMSR system under cyclic square wave flow and load feeding pattern by computer simulation

The dynamic steady state program developed by Dold, Ekama and Marais (1980) extended to include nitrification-denitrification systems by Van Haandel and Marais (1981) had to be modified so that it could be applied in determining the nitrification and denitrification kinetic constants from a cyclically square wave loaded AAACMSR system. The dynamic steady state (cyclic) program calculates the response of the plant over one daily cycle assuming the imposed cycle of flow and load

remains the same from day to day. The modification made was an extension of the program to accommodate a greater variety of operating conditions, by removing the restriction that a reactor in the process either is continuously aerated or continuously unaerated.

The general cyclic program now accommodates alternating unaerated-aerated periods in the daily cycle for each reactor, provided that, either the unaerated-aerated pair of time intervals add up to 24 hours or the unaerated-aerated pairs of intervals are a multiple of 24 hours.

The solution is found by numerical integration of the governing equations by an iterative procedure that continues until the solutions between two successive cycles differ by less than a preselected quantity for each of the parameters solved, (Van Haandel and Marais 1981). The starting values for the iterative procedure are found from the solution of a single reactor system under constant flow and load chosen such that the total flow and mass load per day are the same as under the cyclic flow and load conditions. The simulated daily cyclic response depends on the values assigned to the kinetic constants in the governing differential equations and the cyclic flow and load patterns.

The general dynamic steady state solution has been extensively used to determine the various constants in the governing equations. This is done by curve fitting; the constants are progressively changed until the best fit between observation and prediction is obtained. The difficulty with this approach is that depending on the sludge age, some of the responses become insensitive to the value selected for some constants. To obtain reliable solution of the constants it is necessary therefore to perform the tests over a range of sludge ages using the same sewage. To determine a particular constant, a transient in the response needs to be created that is governed in a large measure by the constant for which an accurate evaluation is sought.

The magnitude of a transient usually is affected by (1) the sludge age, (2) the cyclic pattern of the influent flow and/or load and/or (3) a change in the operating condition at some point in the cycle.

For the purpose of estimating μ_{nmT} , K_{mpT} , η and f_{ca} the following conditions were imposed in order to obtain the most reliable results:

- (1) To obtain maximum activity in the sludge the shortest practical sludge age that allows continuous and stable nitrification was sought; this was found to be about 15 days. (For one particular batch of sewage, when operating at $R_s = 10$ days, nitrification declined consistently even with repeated re-innoculation of nitrifying sludge).
- (2) A square wave cyclic flow and load input was selected (12 hours on 12 hours off) because at the time it was thought that the transients created at the end of the feeding period would allow the fraction of easily biodegradable COD, f_{ca} , to be determined in addition to the nitrification and denitrification kinetic constants.
- (3) A qualitative change in the operating conditions was imposed by interrupting the air supply every day for a period of two hours. The transient thus created was predicted to give rise to large changes in ammonia, nitrate and alkalinity thereby allowing reliable estimates of μ_{nmT} , K_{mpT} and η .

An alternating anoxic-aerobic CMSR system in an apparent dynamic steady state was operated as described earlier in Section 3.3.2. By choosing a two hour anoxic period during the high flow period, the parameters affected most significantly during the anoxic period and on subsequent aeration are NH_3 , TKN, NO_3 and Alk. The absolute magnitudes of the changes expected (and observed) are high and therefore should allow a reliable determination of the constants by curve fitting.

The solution procedure for determining the nitrification and denitrification constants using the dynamic steady state program is based on trial and error curve fitting. The required input data to the dynamic steady state program is:

- (1) Operational conditions
- (2) Cyclic loads
- (3) Sewage characteristics

- (4) Kinetic constants and their temperature dependence.
- (1) The operational conditions for the AAACMSR under consideration, depicted in Fig. 3.3, are listed in Table 3.4.
- (2) The cyclic loads are ideally identical. However due to experimental difficulties in maintaining an identical load for each cycle, the input load fluctuates from cycle to cycle. In Table 3.4 is given the fluctuations of cyclic COD and TKN mass inputs. In addition the *average* load, disregarding any outliers, for all the cycles is indicated. This average value is the value used in the dynamic steady state program simulations.
- (3) All sewage characteristics are listed in Table 3.4. With regard to f_{ca} , it has been shown in Section 3.3.2a(II) that the estimated value of f_{ca} from a square wave loaded AAACMSR is of the same order of magnitude as the error made in the measurement, hence any value of f_{ca} determined by curve fitting has little or no reliability. Consequently, the value of f_{ca} , given in Table 3.4, was found from a short sludge age single aerobic reactor experiment, specifically devised for this purpose, as described in Section 2.3.8.
- (4) All kinetic constants and their temperature dependence, other than those for which evaluation is sought, i.e. μ_{nmT} , K_{mpT} and η , were identical to those used by Van Haandel and Marais (1981). Preliminary estimates of μ_{nmT} , K_{mpT} and η were given by their manually determined values, and for cases where K_{mpT} and η could not be determined reliably by manual means, because of unproductive operating conditions, an accepted average value was used (see Section 3.3.2a).

Dynamic steady state simulations were carried out for experiment EST 32 and each of μ_{nmT} , K_{mpT} and η , respectively, was changed in turn, while keeping all the other constants at their preliminary estimates until the closest fit with experimentally observed response was obtained. By computer trial and error simulations for closest fit of experiment EST 32, using the general dynamic steady state program (program listing is given in Appendix A.1),

and with $f_{ca} = 0,10$, *average* influent TKN = $49 \text{ [mg N.l}^{-1}\text{]}$, *average* influent COD = $507 \text{ [mg COD.l}^{-1}\text{]}$, *average* sewage characteristics given in Table 3.4 and *actual measured* alkalinity = $412 \text{ [mg CaCO}_3\text{.l}^{-1}\text{]}$ (given in Table 3.5b), the following constants were evaluated: $K_{mp20} = 2,3 \text{ [mg COD.mg VSS}^{-1}\text{.d}^{-1}\text{]}$, $\eta = 0,38$, $\mu_{nm20} = 0,48 \text{ [d}^{-1}\text{]}$. A very good correlation between the experimental and simulated responses of experiment EST 32 is given in Fig. 3.6b.

Using the constants above, the response was simulated for experiment EST 31 with $f_{ca} = 0,10 \text{ [mg COD.mg COD}^{-1}\text{]}$, *average* influent TKN = $49 \text{ [mg N.l}^{-1}\text{]}$, *average* influent COD = $507 \text{ [mg COD.l}^{-1}\text{]}$, *average* sewage characteristics given in Table 3.4 and *actual measured* alkalinity = $413 \text{ [mg CaCO}_3\text{.l}^{-1}\text{]}$ (given in Table 3.5a). The same batch of sewage used in experiment EST 32 was also used in experiment EST 31; the difference between experiment EST 32 and EST 31 is that the anoxic period was introduced 2 and 1 hours, respectively, after the beginning of the feed period. The observed and simulated responses for experiment EST 31 are shown in Fig. 3.6a. These show close correlation, indicating that the square wave loaded AAACMSR system is a reliable method for the determination of the nitrification and denitrification constants.

The value of μ_{nm20} for the batch of sewage used in experiments (EST 31 and EST 32) was manually estimated in Section 3.3.2a(I) to be $0,44 \text{ [d}^{-1}\text{]}$. Utilizing $\mu_{nm20} = 0,48 \text{ [d}^{-1}\text{]}$ in the dynamic steady state computer program simulations for experiments (EST 31 and EST 32) gave close correlation between observed and simulated responses [Figs. 3.6 (a and b)]. This proves the method for the manual estimation of μ_{nmT} to be reliable and reasonably accurate.

3.3.3 Alternating anoxic-aerobic CMSR (AAACMSR) system under constant flow and load

Van Haandel and Marais (1981) investigated an AAACMSR system under constant flow and load at 20°C and found that the response indicated very good stability, and allowed an accurate evaluation of the nitrification and denitrification constants - provided the fraction of easily biodegradable influent COD, f_{ca} was known. To determine the fraction of easily biodegradable COD, f_{ca} , a separate special experimental set up and investigation was required, described in Section 2.3.8.

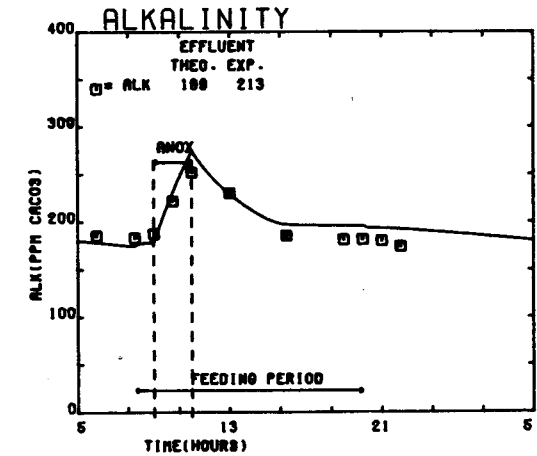
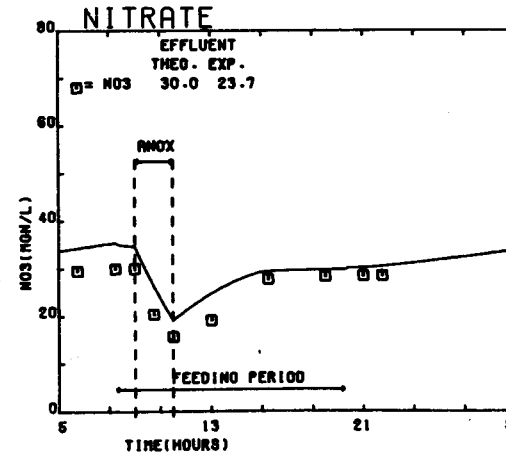
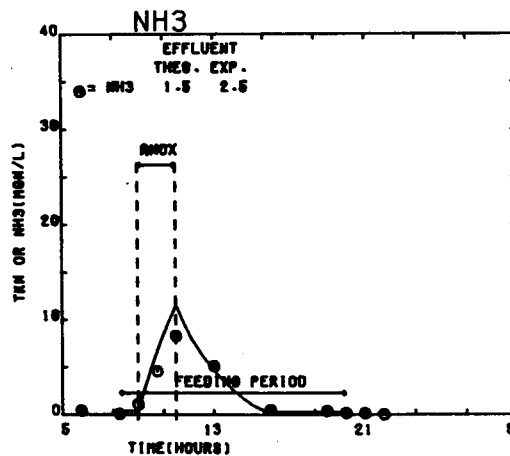
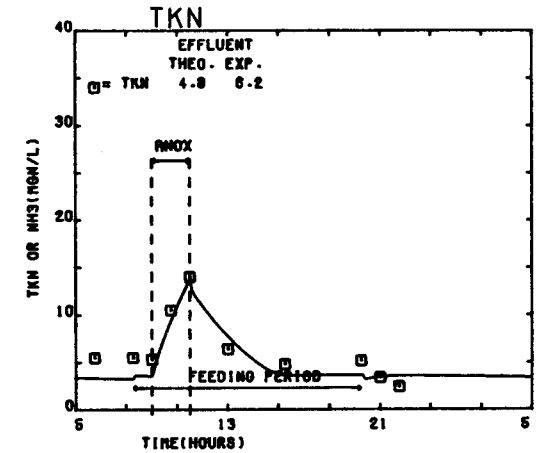
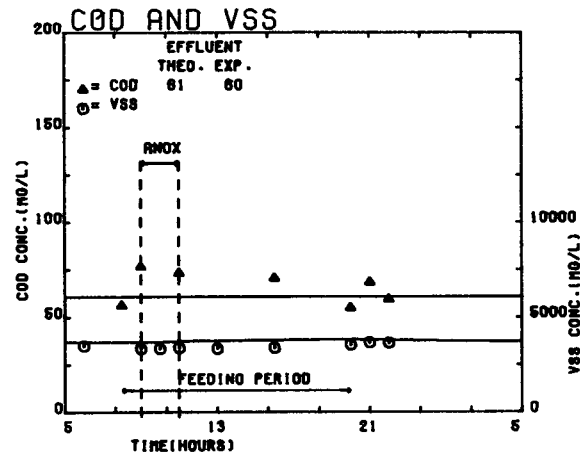
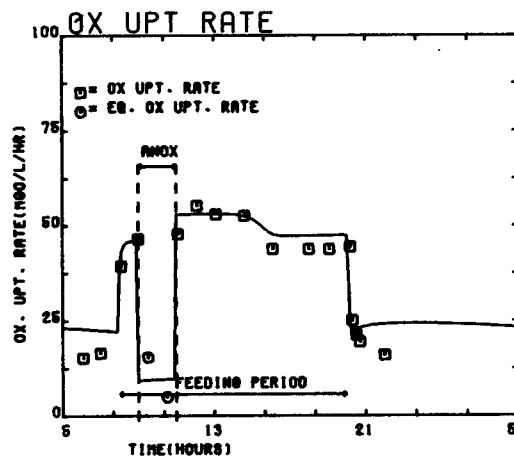


Fig.3.6a: Closest fit of the observed response of an AAACMSRS under square wave flow and load using the dynamic steady state program at 14°C, $R_s = 15$ days, pH = 7.2, with a single anoxic period of 2 hours in 24, introduced 1 hour after the beginning of the feeding period (Experiment EST 31).

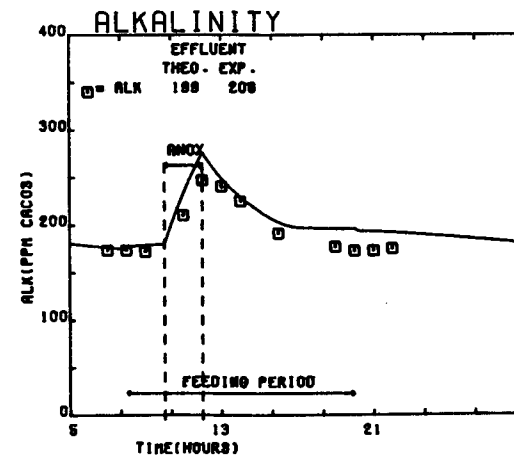
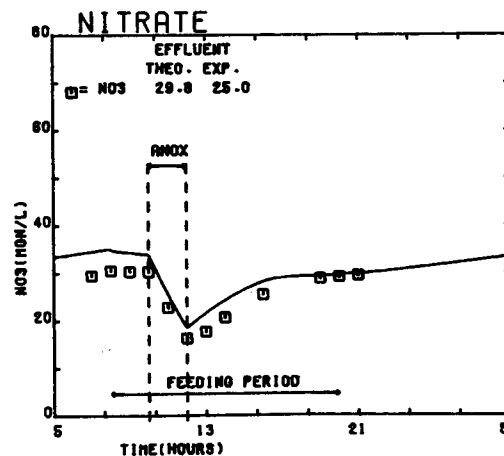
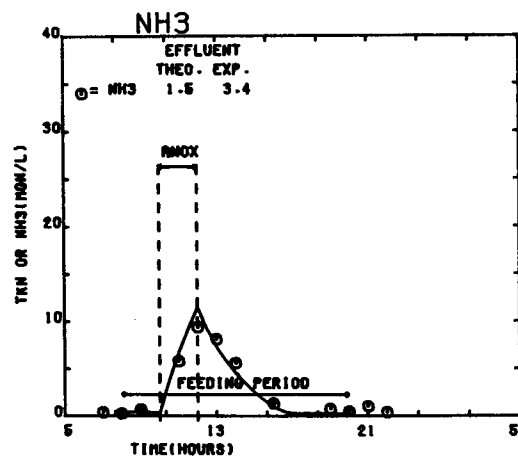
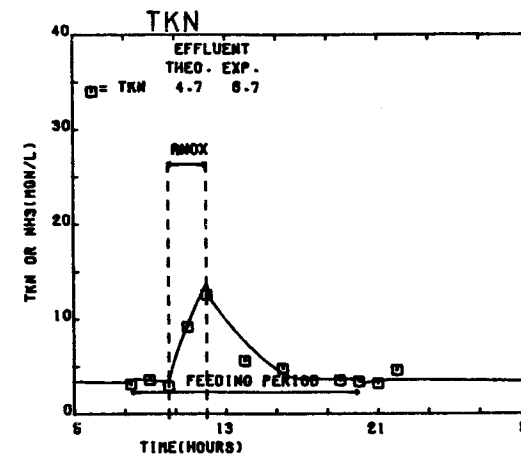
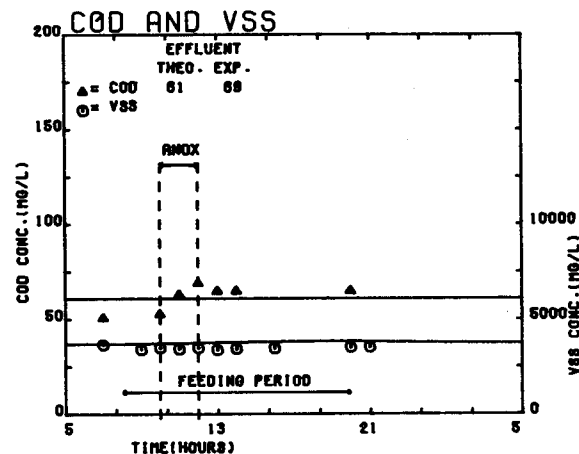
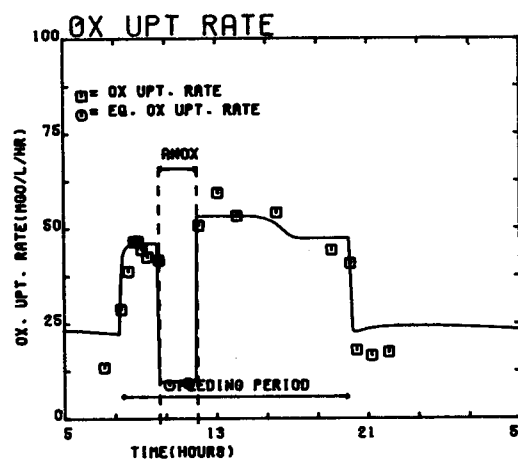


Fig.3.6b: Closest fit of the observed response of an AAACMSRS under square wave flow and load using the dynamic steady state program at 14°C, $R_s = 15$ days, pH=7.2, with a single anoxic period of 2 hours in 24, introduced 2 hours after the beginning of the feeding period (Experiment EST 32).

The alternating anoxic-aerobic CMSR system under constant flow and load has been adopted only very recently (1980) for the purpose of evaluating the nitrification and denitrification constants, and then only at 20°C. No experiments at other temperatures have been undertaken. Consequently to check the value of the method at lower temperatures, experiments were carried out at 14°C under steady state constant flow and load.

The experimental set up is shown in Fig. 3.3 and details of operational and sewage characteristics are listed in Table 3.8. Sequential aerobic and anoxic conditions were induced by switching the electrical current, to an air pump, on and off. The system was run for 2 weeks before intensive testing was undertaken, so that stable conditions could develop. During this period the VSS and mean effluent TKN were checked daily; it was presumed that when these showed no change, steady state had been achieved.

Once steady state had been achieved, intensive testing was done over that fraction of anoxic-aerobic period during which rapid changes of concentrations of nitrate, alkalinity, TKN and NH_3 took place. Samples were taken every hour and the following process variables monitored: Oxygen uptake rate, COD, VSS, TKN, NH_3 , NO_3 and Alk. Three such experiments were carried out in this fashion at approximately one week intervals.

The first two experiments (EST 11 and EST 12) were carried out in an *identical* manner (3 hour anoxic period followed by 21 hour aerobic period) on the *same* batch of sewage. The responses are listed in Tables 3.9(a and b), respectively. In order to determine the easily biodegradable COD fraction, f_{ca} , a short sludge age completely mixed aerobic reactor was run under cyclic square wave flow and f_{ca} determined in accordance with the procedure described in Section 2.3.8. The value of f_{ca} was found to be 0.10 [mg COD. mg COD⁻¹]. From data obtained in experiment EST 11, the nitrification *reaction* rates were calculated from the *observed* nitrate profile for every hour interval using the procedure as described in Section 3.3.1. Evaluation of μ_{nm20} from these rates is described earlier in Section 3.3.2a(I) and was found to be 0.31 [d⁻¹]. By computer trial and error simulations for closest fit, using the general dynamic

Table 3.8 Operational and average sewage characteristics used in Fig. 3.3 under constant flow and load with a single anoxic period of 3 hours in 24.

Operational Characteristics

$T = 14^{\circ}\text{C}$ Reactor volume, $V = 15 \text{ [l]}$

$R_s = 15 \text{ [d]}$ Hydraulic retention time, $R_h = 12 \text{ [h]}$

Duration of stable* operation before testing commenced = 14 [d]

Sewage Characteristics

Influent COD = 444 to 520 $[\text{mg COD.l}^{-1}]$ Average** 486 $[\text{mg COD.l}^{-1}]$

Influent TKN = 48,9 to 59,9 $[\text{mg N.l}^{-1}]$ 54 $[\text{mg N.l}^{-1}]$

$f_{ca} = 0,10 \text{ [mg COD.mg COD}^{-1}]$ (determined experimentally as described in Section 2.3.8)

$f_{up} = 0,09 \text{ [mg VSS.mg COD}^{-1}]$

$f_{us} = 0,15 \text{ [mg COD.mg COD}^{-1}]$

$f_{na} = 0,75 \text{ [mg NH}_3\text{-N.mg N}^{-1}]$

$f_{nu} = 0,0 \text{ [mg N.mg N}^{-1}]$

* Stable operation characteristic to steady state conditions refers to constant VSS and consistent nitrification with a corresponding constant mean effluent ammonia.

** Disregarding any outliers.

Table 3.9a *Observed response of a CMSR system under constant flow and load conditions (experiment EST 11)*
at 14°C, $R_s = 15$ days, pH = 7,25

SAMPLE	TIME [h]	NH ₃ [mgN.l ⁻¹]	TKN [mgN.l ⁻¹]	NO ₃ [mgN.l ⁻¹]	ALKALINITY [mgCaCO ₃ .l ⁻¹]	COD [mgCOD.l ⁻¹]	VSS [mgVSS.l ⁻¹]	TIME [h]	O.U.R. [mgO.l ⁻¹ .h ⁻¹]
Influent	-	34,35	53,6	0,0	439,0	515,1	-	-	-
Effluent	-	2,48	4,0	35,0	187,0	75,8	-	-	-
1	15,00	0,17	1,3	36,6	169,0	-	-	15,12	25,0
2	16,00	0,27	2,1	36,2	166,0	71,7	3566,0	16,08	29,2
3	17,00*	0,35	1,6	36,4	162,0	-	-	16,75	27,6
4	18,00	2,90	6,1	32,3	193,0	75,8	3540,0	-	-
5	19,00	5,77	9,0	26,8	217,0	-	-	-	-
6	20,00**	7,73	9,3	21,9	242,0	79,9	3564,0	20,18	42,4
7	21,00	6,18	8,5	25,3	230,0	-	-	21,07	43,5
8	22,00	4,36	7,2	28,1	214,0	79,9	3586,0	22,10	43,2
9	23,00	2,93	5,2	30,5	198,0	-	-	23,17	45,0
10	24,00	0,57	2,8	33,2	178,0	63,5	3621,0	24,13	38,5

* begin anoxic period

** end anoxic period

Table 3.9b *Observed response of a CMSR system under constant flow and load conditions (experiment EST 12)*
at 14°C, $R_s = 15$ days, pH = 7,2

SAMPLE	TIME [h]	NH ₃ [mgN.ℓ ⁻¹]	TKN [mgN.ℓ ⁻¹]	NO ₃ [mgN.ℓ ⁻¹]	ALKALINITY [mgCaCO ₃ .ℓ ⁻¹]	COD [mgCOD.ℓ ⁻¹]	VSS [mgVSS.ℓ ⁻¹]	TIME [h]	O.U.R. [mgO.ℓ ⁻¹ .h ⁻¹]
Influent		43,4	54,9	0,0	427,0	524,3	-	-	-
Effluent		2,35	4,6	35,1	183,0	67,2	-	-	-
1	15,00	0,70	2,5	36,6	166,0	-	-	15,28	31,7
2	16,00	0,22	-	37,9	163,0	75,8	3734,0	16,33	31,4
3	17,00*	-	3,1	37,9	-	-	-	16,82	30,6
4	18,00	3,53	7,3	33,7	190,0	75,8	3720,0	-	-
5	19,00	6,48	10,3	28,0	218,0	-	-	-	-
6	20,00**	9,02	11,0	23,1	238,0	73,7	3684,0	20,10	45,0
7	21,00	7,82	10,3	25,3	229,0	-	-	21,10	47,5
8	22,00	5,75	8,2	27,6	216,0	69,6	3731,0	22,17	45,2
9	23,00	2,57	5,1	32,3	191,0	-	-	23,10	45,3
10	24,00	0,32	3,6	34,8	171,0	71,7	3631,0	24,08	35,0
11	1,00	0,27	2,8	35,1	168,0	-	-	1,13	32,3
12	2,00	0,19	3,3	35,9	167,0	-	-	2,05	30,8

* begin anoxic period

** end anoxic period

steady state program, and with $f_{ca} = 0,10$, *average* influent COD = 486,0 [mg COD. ℓ^{-1}], *average* influent TKN = 54,0 [mg N. ℓ^{-1}], *average* sewage characteristics given in Table 3.8 and *actual measured* alkalinity = 439,0 [mg CaCO₃. ℓ^{-1}] (given in Table 3.9a), the following constants were evaluated: $K_{mp20} = 2,3$ [mg COD.mg VSS⁻¹.d⁻¹], $\eta = 0,38$, $\mu_{nm20} = 0,33$ [d⁻¹]. Excellent correlation was observed between the experimental and simulated responses[see Fig 3.7a] so that it is very likely that the constants found above were reliable.

Using the constants derived above, the response was simulated for the second experiment (EST 12), using the general dynamic steady state program with $f_{ca} = 0,10$, *average* influent COD = 486,0 [mg COD. ℓ^{-1}], *average* influent TKN = 54,0 [mg N. ℓ^{-1}], *average* sewage characteristics given in Table 3.8 and *actual measured* alkalinity = 427,0 [mg CaCO₃. ℓ^{-1}] (given in Table 3.9b). The observed and simulated responses are shown in Fig. 3.7b. These show close correlation, indicating that the experimental procedure gave rise to remarkably reproducible data; this verifies the findings of Van Haandel and Marais (1981) that the constant flow and load AAACMSR system is a reliable method for the determination of the nitrification and denitrification constants.

In the two experiments the observed and simulated responses showed very good correlation for oxygen uptake rate, COD, VSS, TKN, ammonia (NH₃), nitrate (NO₃) and alkalinity (Alk), [Figs. 3.7(a and b)], and lend support to the adequacy of the general dynamic steady state model under time varying input conditions. In particular it verifies the reliability of the alkalinity parameter, recently incorporated into the general model, as an additional check on the nitrification and denitrification behaviour of a single sludge system.

To investigate the effect of the length of anoxic period on the *nitrification* response, a third experiment (EST 21) was carried out with a relatively long anoxic period i.e. 4 hour anoxic period (instead of 2 hour) followed by a 20 hour aerobic period. It was hypothesized that if the anoxic period affects the nitrifiers, the nitrification rate may show a decline for a certain period of time immediately after aerobic conditions are reimposed. By computer trial and error simulations for closest fit of experiment EST 21,

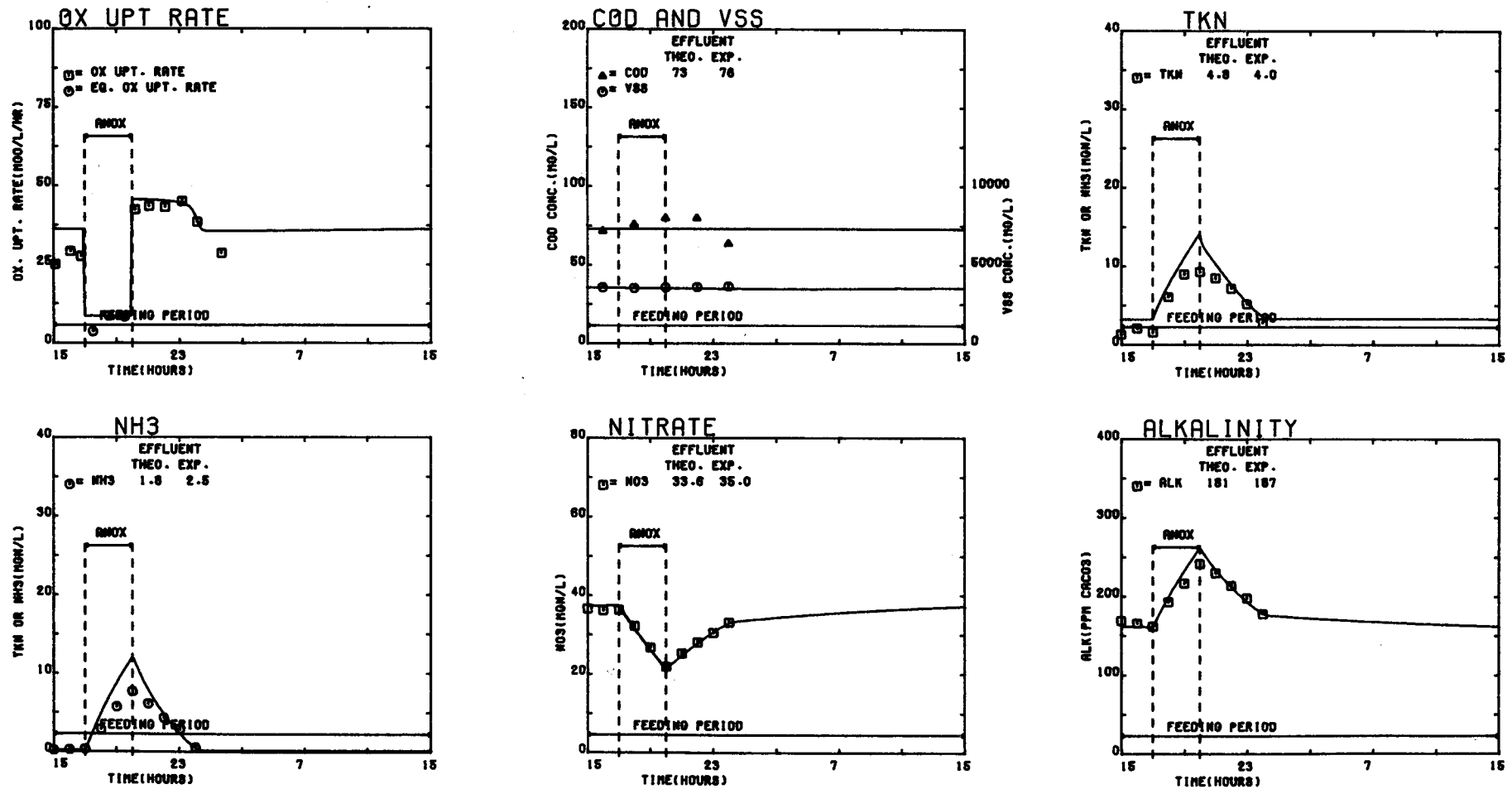


Fig.3.7a: Closest fit of the observed response of an alternating anoxic-aerobic completely mixed single reactor system (AAACMSRS) under constant flow and load using the dynamic steady state program at 14°C, $R_s = 15$ days, pH=7.25, $f_{ca}=0.10[\text{mgCOD}.\text{mgCOD}^{-1}]$, with a single anoxic period of 3 hours in 24 (Experiment EST 11).

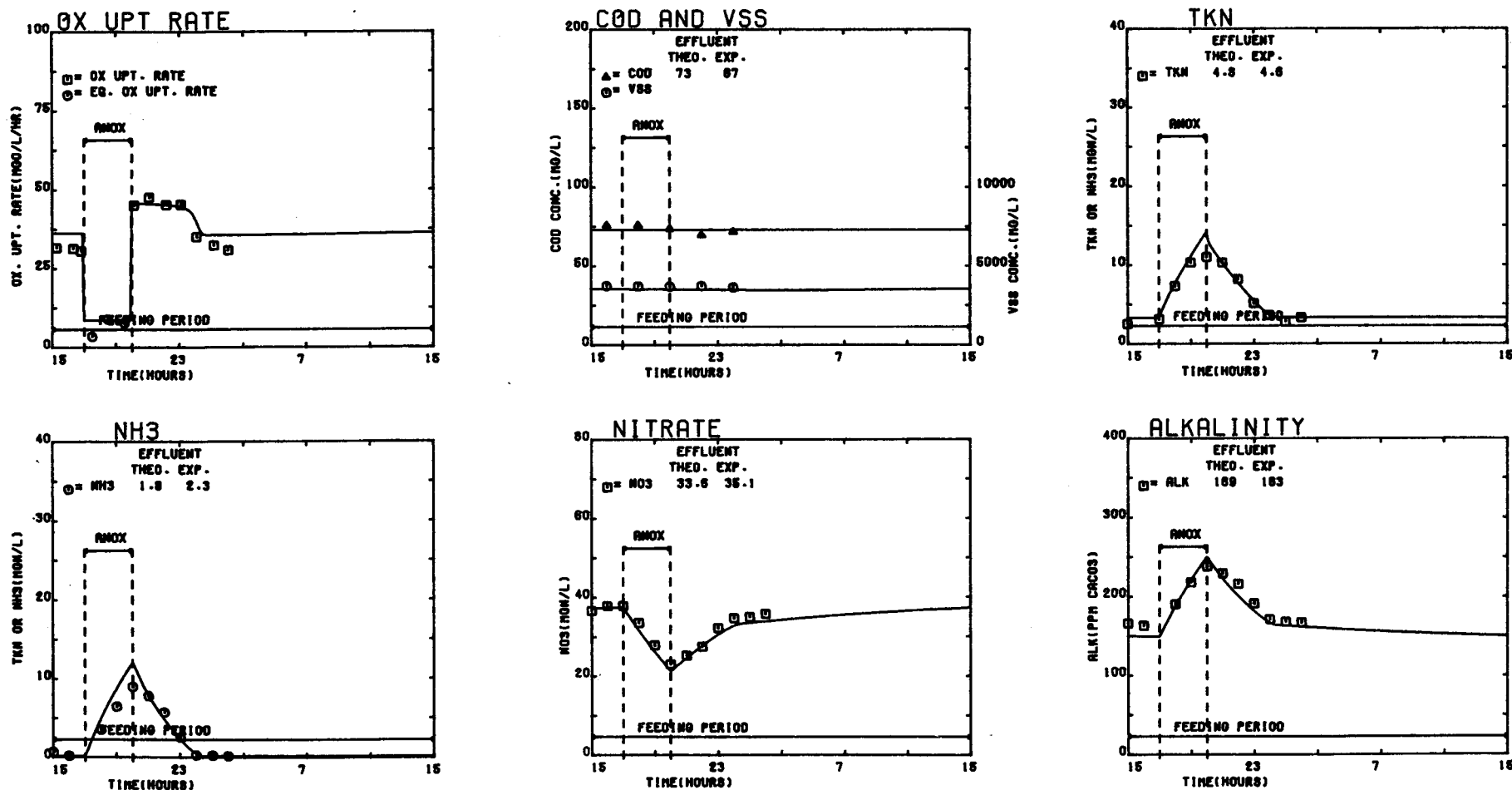


Fig.3.7b: Closest fit of the observed response of an AAACMSRS under constant flow and load using the dynamic steady state program at 14°C, $R_s = 15$ days, $f_{ca} = 0.10$ [mgCOD.mgCOD⁻¹], pH=7.2, with a single anoxic period of 3 hours in 24 (Experiment EST 12).

using the general dynamic steady state program and with $f_{ca} = 0,10$ [mg COD.mg COD⁻¹], *average* influent COD = 489,0 [mg COD.l⁻¹], *average* influent TKN = 46,0 [mg N.l⁻¹], *average* sewage characteristics given in Table 3.10 and *actual measured* alkalinity = 401,7 [mg CaCO₃.l⁻¹] (given in Table 3.11), the following constants were evaluated: $K_{mp20} = 3,0$ [mg COD.mg VSS⁻¹.d⁻¹], $\eta = 0,38$, $\mu_{nm20} = 0,33$ [d⁻¹]. A consistent* correlation between the experimental and simulated response of experiment EST 21 over *the whole* period monitored (Fig.3.8a) indicates unequivocally that the μ_{nmT} observed did not show evidence of change in the period immediately after termination of the anoxic period. If it did, it would not have been possible to consistently* fit the nitrate response curve by simulation during this period with a constant value for μ_{nmT} . Thus, for an anoxic period of 4 hours every 24 hours, the system nitrification appeared to be unaffected. This observation does not imply that such inhibition might not occur in other tests with different sources of sewage and different operational conditions.

Attention is now given to Fig. 3.8a where closest fit of experiment EST 21 does not satisfactorily describe the nitrate and alkalinity responses. Fig. 3.8a clearly indicates that when the *average* influent TKN load is used as an input data to the dynamic steady state program *for experiment EST 21*, the nitrate and alkalinity responses over the *whole* period monitored are *consistently* over-estimated. In all other cases where the *average* influent TKN load was used as an input data to the dynamic steady state program, Figs. [3.6(a and b), and 3.7(a and b)], very good correlations between experimental and observed responses were obtained.

It is hypothesized that the failure of the dynamic *steady state* model to adequately describe the nitrate and alkalinity responses (i.e. the nitrification behaviour) in experiment EST 21 stems from *unsteady* operating conditions that prevailed on the day testing was conducted. Table 3.12 gives a comparative study of the deviation

* Although correlation of say the nitrate response in Fig. 3.8a is not excellent, it is *consistently* over predicted. It will be shown later that this problem arises due to limitations of the dynamic steady state model to describe the response governed by the experimental conditions employed in experiment EST 21, and it can be resolved without affecting the value of any of the kinetic constants.

Table 3.10 Operational and average sewage characteristics used in Fig. 3.3 under constant flow and load with a single anoxic period of 4 hours in 24.

<u>Operational Characteristics</u>	
$T = 14,7^{\circ}\text{C}$	Reactor volume, $V = 15 \text{ [}\ell\text{]}$
$R_s = 15 \text{ [d]}$	Hydraulic retention time, $R_h = 12 \text{ [h]}$
Duration of stable* operation before testing commenced = 5 [d]	
<u>Sewage Characteristics</u>	
<u>Average**</u>	
Influent COD = 461,7 to 561,5 [mg COD. ℓ^{-1}]	489,0 [mg COD. ℓ^{-1}]
Influent TKN = 45,4 to 53,1 [mg N. ℓ^{-1}]	46,0 [mg N. ℓ^{-1}]
$f_{ca} = 0,10 \text{ [mg COD.mg COD}^{-1}\text{]}$	
$f_{up} = 0,09 \text{ [mg VSS.mg COD}^{-1}\text{]}$	
$f_{us} = 0,15 \text{ [mg COD.mg COD}^{-1}\text{]}$	
$f_{na} = 0,60 \text{ [mg NH}_3\text{-N.mg N}^{-1}\text{]}$	
$f_{nu} = 0,0 \text{ [mg N.mg N}^{-1}\text{]}$	

* Stable operation characteristic to steady state conditions refers to constant VSS and consistent nitrification with a corresponding constant mean effluent ammonia.

** Disregarding any outliers.

Table 3.11 *Observed response of a CMSR system under constant flow and load conditions (experiment EST 21)*
at 14,7°C, $R_s = 15$ days, pH = 7,2

SAMPLE	TIME [h]	NH ₃ [mgN.l ⁻¹]	TKN [mgN.l ⁻¹]	NO ₃ [mgN.l ⁻¹]	ALKALINITY [mgCaCO ₃ .l ⁻¹]	COD [mgCOD.l ⁻¹]	VSS [mgVSS.l ⁻¹]	TIME [h]	O.U.R. [mgO.l ⁻¹ .h ⁻¹]
Influent		33,28	52,9	0,0	401,7	561,5	-	-	-
Effluent		2,1	6,0	21,0	211,7	79,6	-	-	-
1	14,00	0,32	4,0	24,3	185,7	-	-	14,42	33,7
2	15,00	0,04	5,2	24,7	180,8	59,5	3788,0	15,15	33,3
3	16,00*	0,15	5,4	26,1	176,4	-	-	15,88	33,9
4	17,00	2,84	8,8	20,0	197,5	-	-	-	-
5	18,00	4,95	11,6	14,4	237,9	-	-	-	-
6	19,00	7,40	12,8	8,6	263,2	-	-	-	-
7	20,00**	9,04	17,5	2,6	283,8	109,9	3744,0	20,17	51,7
8	21,00	6,59	12,4	7,6	263,2	-	-	21,27	59,0
9	22,00	0,0	7,5	13,1	233,0	67,5	3749,0	22,15	44,8
10	23,00	0,11	5,8	17,7	206,5	-	-	23,02	36,0
11	24,00	0,0	5,4	19,2	203,4	65,5	3762,0	24,22	33,7
12	1,00	0,04	4,6	20,2	196,7	-	-	-	-
13	2,00	0,04	4,6	21,3	197,2	65,5	3743,0	2,75	33,0

* begin anoxic period

** end anoxic period

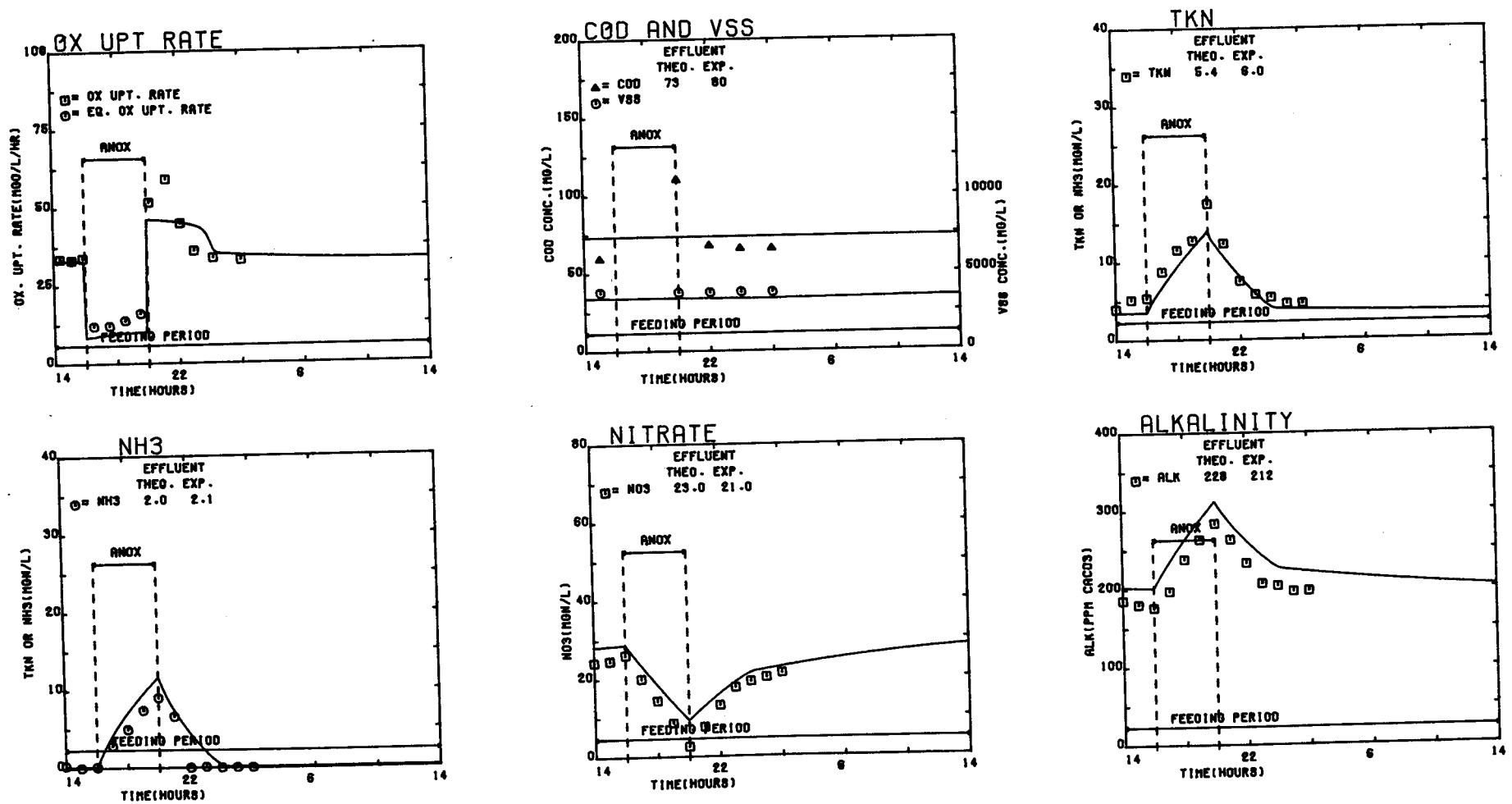


Fig.3.8a Closest fit of the observed response of an AAACMSRS under constant flow and load using the dynamic steady state program at 14,7°C, $R_s = 15$ days, $f_{ca}=0.10$ [mgCOD.mgCOD⁻¹], pH=7,2, with a single anoxic period of 4 hours in 24 (Experiment EST 21).

of nitrification behaviour in AAACMSR systems from dynamic steady state by comparing for all the AAACMSR experiments,

- (1) the *average* influent TKN concentration used in the dynamic steady state simulations to the *actual measured* TKN concentration on the day testing was conducted, and
- (2) the correlations obtained between experimental and observed nitrate and alkalinity responses.

Table 3.12 indicates that there exists a very strong correlation between the extent of deviation of *actual measured* influent TKN from the *average* influent TKN to the goodness of fit of nitrate and alkalinity responses; as the deviation from average influent TKN load decreases so the *actual dynamic steady state* nitrification behaviour is more closely predicted by the general *dynamic steady state program*, with concomitant closer correlations between the observed and predicted responses of nitrate and alkalinity. In principle the same reasoning can also be applied to the extent of deviation of the *actual measured* from the *average* influent COD but in practice, unless the magnitude of this deviation is large, the measurable parameters governed by COD utilization are either insensitive to the deviation or can not be directly used; the VSS test is insensitive to small deviations from dynamic steady state carbonaceous metabolic activities while the total oxygen uptake rate test includes dynamic steady state oxygen requirements for nitrification, which makes the estimation of dynamic steady state *carbonaceous* oxygen requirements heavily dependent on the validity of the nitrification description.

From the above discussion it seems that the dynamic steady state model is suitable for accurate estimations of the kinetic constants governing nitrification and denitrification from an AAACMSR system *only if* the operating conditions are such that the deviation from (average) dynamic steady state behaviour is small. Fortunately this was the case in experiments (EST 31, EST 32, EST 11 and EST 12) each of which was operated under stable conditions for a period of 2 weeks or more before testing commenced. In experiment EST 21 however stable operation was maintained for 5 days only before testing commenced and the *actual measured* influent TKN load on the day testing

Table 3.12 *Comparative study of deviation of nitrification behaviour in AACMSR systems from dynamic steady state.*

Experiment number	Stable * Operation [d]	Average ** influent TKN [mgN.l ⁻¹]	Actual *** measured influent TKN [mgN.l ⁻¹]	Deviation of average from actual measured influent TKN	Dynamic steady state closest fit Fig.number	Correlation between experimental and observed nitrate and alkalinity responses
EST 31	14	49,0	51,9	5,9%	3.6a	Satisfactory
EST 32	14	49,0	51,2	4,5%	3.6b	Satisfactory
EST 11	14	54,0	53,6	0,7%	3.7a	Excellent
EST 12	14	54,0	54,9	1,7%	3.7b	Very good
EST 21	5	46,0	52,9	15 %	3.8a	Unsatisfactory

* Stable operation is the period before testing commenced during which constant VSS and effluent ammonia were measured.

** Average influent TKN concentration disregarding any outliers.

*** Actual measured influent TKN on the day testing was conducted.

was conducted deviated by 15% from the *average* influent TKN load. This, *inter alia*, led to unsatisfactory description of the nitrification behaviour when using the dynamic steady state model, hence unsatisfactory predictions of nitrate and alkalinity responses were obtained [Fig. 3.8a].

Resolution of this problem necessitated a model that can accommodate deviations from ideal dynamic steady state (cyclicity). Such a flexible program is the unsteady state (transient) program developed by Van Haandel and Marais (1981). The application of the unsteady state program to determine the nitrification and denitrification constants from an alternating anoxic-aerobic CMSR system is as follows:

In simulations using the unsteady state program of Van Haandel and Marais (1981) the response is calculated by a non iterative straightforward integration procedure. In the unsteady state program the starting values of the process variables at the beginning of the experiment must be fed in as input data; the starting values of the measurable soluble parameters (S_u , NH_3 , NO_3 , Alk) are obtained from experimental measurements while the variables that cannot be measured (X_a , X_s , X_e , X_i , S_{bp} , X_n) are estimated from theory, (e.g. by using the general dynamic steady state program).

In experiment EST 21 the *dynamic steady state* values of the sludge parameters, i.e. X_a , X_s , X_e , X_i , S_{bp} and X_n can be accepted. This is valid because the fluctuations in loads and sewage characteristics affect the values of the sludge parameters to a much lesser extent than the values of the soluble parameters; the sludge age is 15 days and this means that if approximate dynamic steady state conditions are achieved, every day about 6 per cent of the sludge is withdrawn (by wastage) and is concurrently replaced by metabolic activities and enmeshment with a zero net increase in sludge mass. In contrast, all soluble parameters are directly affected by the hydraulics of the system, i.e. total daily flow is put through the system in 24 hours every day. This means that if for instance the soluble ammonia concentration doubles, i.e. NH_3 increases by one hundred per cent, X_n will at the most be affected by a net increase of six per cent.

The determination of μ_{nmT} , K_{mpT} and η is done by using initial estimates for these constants, e.g. by using the kinetic constants that effected closest fit with the dynamic steady state model and then changing each one of them in turn keeping the others unchanged until the best fit with the experimental response is obtained. The magnitude of f_{ca} is determined from a short sludge age single aerobic completely mixed reactor as described in Section 2.3.8.

From the discussion above it seems that the unsteady state program should be suitable for the determination of reliable nitrification and denitrification constants from an AAACMSR that approximately achieved a dynamic steady state on the day testing commenced.

Using the unsteady state program with the following input data: $f_{ca} = 0,10$ [mg COD.mg COD⁻¹], *average* sewage characteristics given in Table 3.10, *actual measured* influent COD = 561,5 [mg COD.l⁻¹], *actual measured* influent TKN = 52,9 [mg N.l⁻¹], *actual measured* alkalinity = 401,7 [mg CaCO₃.l⁻¹], nitrification and denitrification kinetic constants that effected closest fit between experimental and simulated responses by using the dynamic steady state program with *average* influent load as input data, i.e. $K_{mp20} = 3,0$ [mg COD.mg VSS⁻¹.d⁻¹], $\eta = 0,38$, $\mu_{nm20} = 0,33$ [d⁻¹] and, with the following dynamic steady state starting values taken from the dynamic steady state computer simulation for experiment EST 21: $S_{bp} = 6,264$ [mg COD.l⁻¹], $X_s = 73,34$ [mg VSS.l⁻¹], $X_a = 1227,0$ [mg VSS.l⁻¹], $X_e = 776,25$ [mg VSS.l⁻¹], $X_i = 1348,9$ [mg VSS.l⁻¹] and $X_n = 63,08$ [mg VSS.l⁻¹], together with the *actual measured* starting values for NO₃, NON, NH₃ and Alk. (given in Table 3.11), very good correlation was obtained between the predicted and observed responses given in Fig. 3.8b.

Comparing the predictive power of the dynamic steady state model with that of the unsteady model for experiment EST 21, Figs. 3.8 (a and b), respectively, the nitrate and alkalinity responses are *much* more closely predicted by the unsteady model. This can be attributed to the inadequacy of the dynamic steady state model to accommodate a combination of deviations, the one from ideal dynamic steady state *average influent TKN load* and the other from dynamic

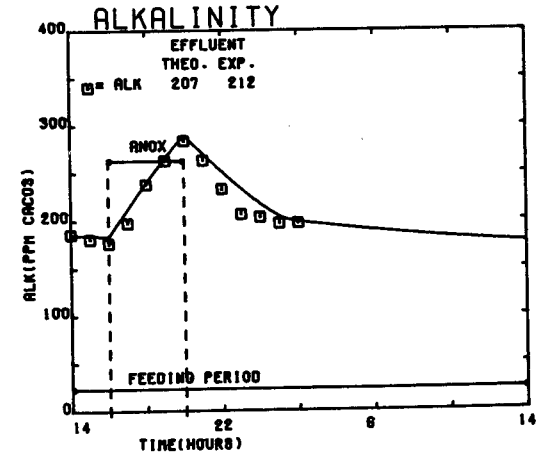
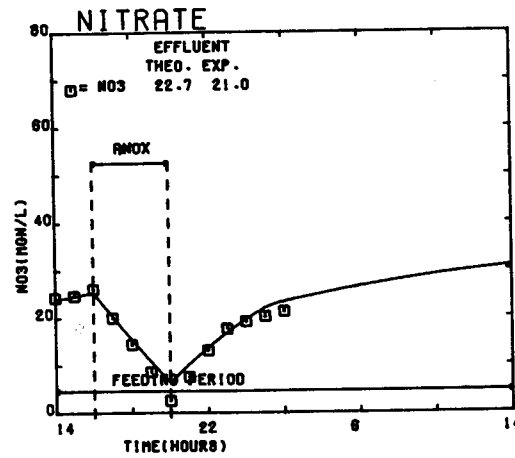
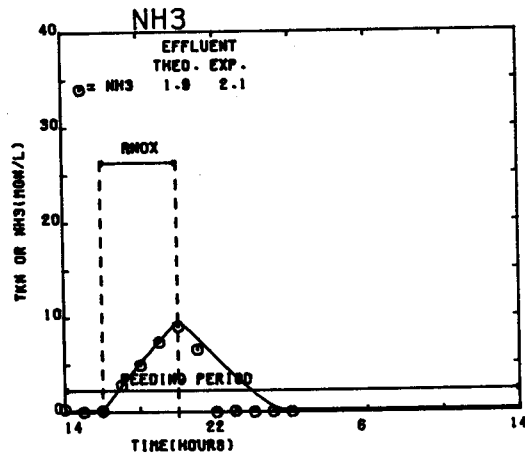
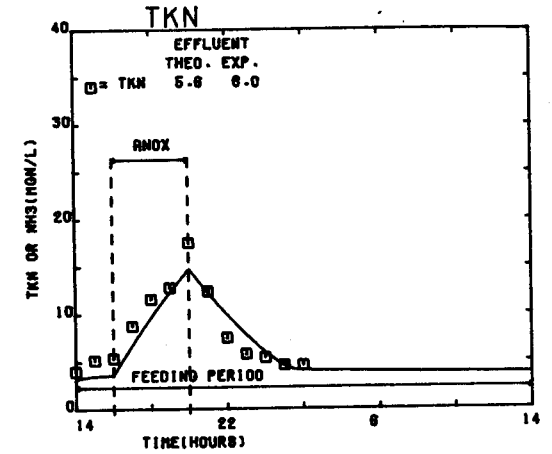
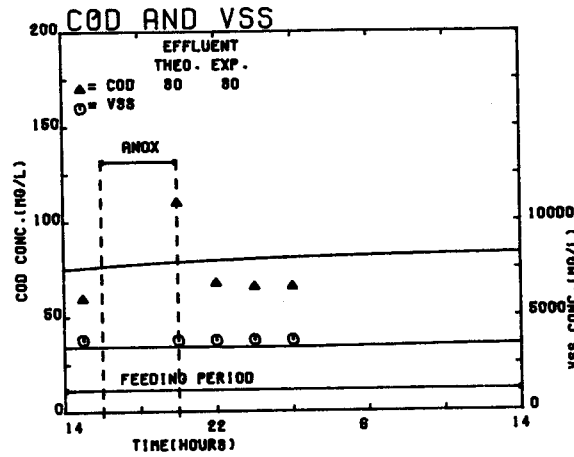
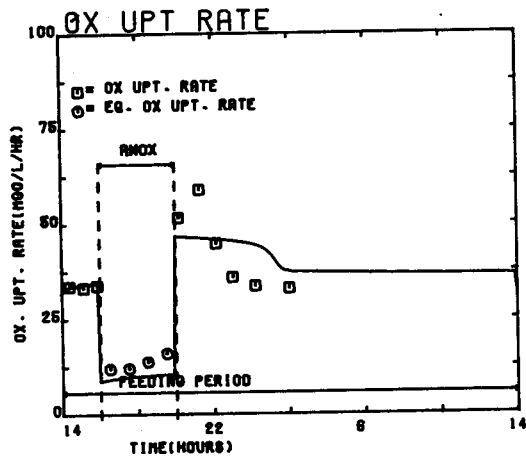


Fig.3.8b Closest fit of the observed response of an AAACMSRS under constant flow and load using the unsteady state program at 14,7°C, $R_s = 15$ days, $f_{ca} = 0,10$ [mgCOD.mgCOD⁻¹], pH=7,2, with a single anoxic period of 4 hours in 24 (Experiment EST 21).

steady state *starting values* for nitrate and alkalinity. It is important to note that even though the unsteady state program gave a better correlation with observed response than the dynamic steady state program, it made use of steady state starting values for S_{bp} , X_s , X_a , X_e , X_i and X_n which can *not* be *directly* measured.

In simulations of experiment EST 21 the kinetic constants that effected closest fit with the dynamic steady state model also effected closest fit with the unsteady model, however the nitrate and alkalinity responses were much more closely fitted when using the unsteady model. Hence when operating under steady state or close to steady state conditions, depending on how closely ideal steady state has been attained, either the dynamic steady state model is sufficiently accurate in predicting the nitrification and denitrification constants from an AAACMSR system or the unsteady state model must be used *in conjunction with* its dynamic steady state counterpart.

3.3.4 Application of a CMSR to the determination of the empirical denitrification constants for a pre-denitrification system

Design of any predenitrification system is normally based on the empirical approach [see Eq.(2a)]. Hence, from a practical point of view it is advantageous to know the values of the denitrification constants. These are: (1) f_{ca} that determines the nitrate removal due to utilization of the easily biodegradable substrate, and (2) K_{2T} that determines the nitrate removal due to utilization of slowly biodegradable substrate.

Considering experiment EST 32, it has been shown in Section 2.3.8 that f_{ca} could be determined by employing a short sludge age completely mixed aerobic reactor under cyclic square wave flow and load. Following the procedure described in Section 2.3.8 the magnitude of f_{ca} was estimated as $f_{ca} = 0,10$ [mg COD.mg COD⁻¹] without the aid of a computer. In Section 3.3.2a(IV) the daily nitrate removal due to utilization of slowly biodegradable COD, ΔN_{sbp} , is given by Eq. (3.14) which requires knowledge of the total daily nitrate removal, ΔN and the daily nitrate removal due to the utilization of easily biodegradable COD, ΔN_{sbs} .

The value of ΔN_{sbs} is evaluated using Eq. (3.15), i.e.

$$\Delta N_{sbs} = 2,5 \cdot 2.365,5 \cdot 0,10 \cdot (1 - 1,54 \cdot 0,45) / 2,86 = 19,6 \text{ [mg N.d}^{-1}\text{]}.$$

The value of ΔN is calculated from the estimated denitrification reaction rate during the anoxic period given in Table 3.6. From Table 3.6,

$$\Delta N = 3,4 \cdot 1 \cdot 15 + 3,0 \cdot 1 \cdot 15 = 96 \text{ [mg N.d}^{-1}\text{]}.$$

Hence from Eq. (3.14)

$$\Delta N_{sbp} = \Delta N - \Delta N_{sbs} = 96 - 19,6 = 76,4 \text{ [mg N.d}^{-1}\text{]}.$$

The rate of nitrate removal due to utilization of slowly biodegradable COD over the 2 hour anoxic period, $r_{N,sbp}$ is

$$r_{N,sbp} = \Delta N_{sbp} / (t_a \cdot V) = 76,4 / (2 \cdot 15) = 2,55 \text{ [mg N.l}^{-1}\text{.h}^{-1}\text{]}.$$

Comparing $r_{N,sbp}$ with Eq. (2a) it follows that

$$r_{N,sbp} = \Delta N_2 / R_1 = K_{2T} \cdot X_a$$

Hence, with the previously accepted

$$X_a = 1259 \text{ [mg VSS.l}^{-1}\text{]},$$

K_{2T} can be evaluated at 14°C as

$$K_{2_{14}} = r_{N,sbp} / X_a = 2,55 / 1259 = 2,03 \cdot 10^{-3} \text{ [mg N.mg VSS}^{-1}\text{.h}^{-1}\text{]}$$

i.e.,

$$K_{2_{14}} = 0,049 \text{ [mg N.mg VSS}^{-1}\text{.d}^{-1}\text{]}.$$

It is important to note that K_{2T} can always be determined manually regardless of the operational conditions as it does not require knowledge of K_{mpT} , K'_{mpT} or X_s . However extrapolation of experimental results to other operational conditions is only possible by computer simulations.

CHAPTER 4

CONCLUSIONS

This investigation was concerned with the development of different experimental set-ups and operational procedures in search for accurate and reliable evaluation of the kinetic constants and sewage characteristics that influence nitrification (μ_{nmT}) and denitrification (K'_{mpT} and f_{ca}).

Two different set-ups were investigated:

- (1) Series completely mixed three reactor configuration (Fig 3.1), with the first reactor anoxic, the second and third aerobic, operated under (a) constant flow and load and (b) constant flow and COD load with a varying TKN load.
- (2) Single completely mixed reactor system under sequential anoxic-aerobic conditions with two modes of operation, (a) constant flow and load and (b) square wave flow and load.

Three reactor system:

The three reactor series system was operated under *constant* flow and load in an effort to eliminate the instability normally encountered when operating under cyclic flow and load. It was hypothesized that if the retention time in the first aerobic reactor was made so short that the reaction of interest was not complete, a "transient" type condition would be induced and the need for a cyclic flow and load to achieve transients could be eliminated.

Analysis of the experimental response of the system indicated that the transient condition was ill defined - estimation of the constants by calibration could be obtained only within a range of values.

To intensify the transient state it was hypothesized that if a high TKN concentration slug was introduced over a short period of time each day, theoretically a high intensity transient could be induced and because the TKN pulse flow was small, constant flow and COD load could still be maintained. This did not prove successful principally because of the difficulties encountered in ensuring

accurate metering of the TKN slug. The cyclic nature of the TKN load again was influenced by the recycle rates *as if* the TKN input was in effect under cyclic flow and load, and this resulted again in unreliable estimates of the constants. Furthermore, it was found that from a normal practice point of view to fit the simulations to the data was time consuming to the point of impracticability.

Single reactor system:

In search for alternative set-ups and operating procedures, the single reactor system subjected to alternating aerobic and anoxic conditions (by appropriate control of aeration) under constant or cyclic flow and load was investigated. This proved a more reliable set-up than the series system:

- (1) The single reactor system considerably simplified operation; the need for accurate metering of the recycle flows in the series system fell away because the single recycle in the single reactor system is in fact intra-reactor and as such does not affect the kinetic response as long as there is no accumulation of sludge in the settler.
- (2) Large variations of ammonia and nitrate with time could be induced by simply selecting the appropriate anoxic-aerobic time sequence.
- (3) The experimental response from a single reactor system was found to be stable and reproducible.
- (4) The general dynamic steady state model could be readily adapted to simulate the response of the single reactor process to any selection sequence of anoxic and aerobic periods under either constant or cyclic flow and load conditions and the constants μ_{nmT} , K_{mpT} and η could be readily determined by calibration.
- (5) In the single reactor system (in contrast to the series reactor system) the whole mass of sludge is exposed to either anoxic or aerobic conditions and thereby provides a means to test the effect of the anoxic environment on nitrification organisms in the subsequent aerobic state. This showed that for anoxic and aerobic sequence of up to 4 and 20 hours respectively, under constant flow and load, no apparent effect was observed. This however

may not be the case under different operating conditions and with different sewage sources.

- (6) In experiments where on the day of the test the input varied from the mean inputs for the period prior to the test, the calibration procedure was modified as follows. The process was simulated using the mean input data and the resulting output response was used as starting values for simulation of the output response on the day testing was done. The starting values of the independent parameters were either the experimentally measured values (S_u , NH_3 , NON , NO_3 , Alk) or, where direct measurements were not possible (for S_{bp} , K_s , K_a , K_e , K_i , K_n), the corresponding either constant or cyclic steady state values of the general dynamic steady state model. This approach led to very successful calibration (as indicated by the close fits obtained) and hence reliable determination of the constants.
- (7) It was shown possible to manually determine good estimates for μ_{nmT} from the nitrate response of a single reactor system. Manual determination was also possible for K_{mpT} and η provided the easily biodegradable COD fraction in the influent was available. Manual estimations should be of particular usefulness in cases where computer facilities are not available.

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APPENDIX A

A.1 Listing of the computer program for the dynamic steady state model.

A.2 Listing of the computer program for the unsteady state model.

A.3 Listing of the computer program used for plotting experimental and predicted responses of an alternating anoxic aerobic completely mixed single reactor system.

PROGRAM A.1

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'THIS PROGRAMME IS THE JOINT PROPERTY OF THE
UNIVERSITY OF CAPE TOWN AND THE WATER RESEARCH
COMMISSION OF SOUTH AFRICA. BEFORE USER MAY
LEGALLY UTILIZE IT, HE MUST OBTAIN THE NECESSARY
AUTHORIZATION, I.E.:

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CAPE TOWN WHO WILL USE THE PROGRAMME FOR
EDUCATIONAL PURPOSES ONLY, THIS AUTHORIZATION
CAN BE OBTAINED FROM PROF. G. VAN R. MARAIS
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COMMISSION (P.O. BOX 824, PRETORIA, 0001,
SOUTH AFRICA)''

FOR A DETAILED GUIDE DESCRIBING THE USE OF THIS
PROGRAM REFER TO:

' MANUAL FOR USING THE GENERAL ACTIVATED SLUDGE
SIMULATION PROGRAMS', DEPT. OF CIVIL ENG.
UNIVERSITY OF CAPE TOWN.

26 C*****

27 C DYNAMIC STATE MODEL OF ACTIVATED SLUDGE PROCESS FOR SERIES REACTORS

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(INCLUDING STORAGE OF COD, NITRIFICATION AND
DENITRIFICATION
CALCULATION CHARACTERISTICS

THIS PROGRAMME REQUIRES THE FOLLOWING CORE STORAGE CAPACITY:-
FOR ND=241 AND NF=4*(ND-1)+1 AND NEE=NE+1
STORAGE=10*NE+23 KILOWORDS
DOUBLING ND INCREASES STORAGE REQUIREMENT BY 1.75
APPROX. CALCULATION CPU TIME ON A UNIVAC 1106 COMPUTER
FOR 100 ITERATIONS:
CPU TIME (SECONDS) =50*NE+30

KO = TIME BASE VARIABLE
M = CYCLE NUMBER MONITOR
D = NUMBER OF INTERGRATION STEPS PER DAY
DT = INTERGRATION STEP LENGTH
DD = NUMBER OF INTERGRATION STEPS PER DAY FOR NITRIFICATION
DDT = INTERGRATION STEP LENGTH FOR NITRIFICATION
RDT = RATIO OF INTERGRATION STEP LENGTHS
KH = NUMBER OF TIME STEPS PER HOUR = D/24
HK = NO. OF INTERVALS PER DAY OF SLUDGE WASTAGE

NOTE

NITROGEN CONCENTRATIONS MEASURED IN MG-N/L WITH TKN TEST
COD CONCENTRATIONS MEASURED IN MG/L OF COD
VSS CONCENTRATIONS MEASURED IN MG/L OF VSS
OXYGEN CONSUMPTIONS IN MG/L/HOUR
UNIT OF MASS IS THE MG
UNIT OF VOLUME IS THE L
UNIT OF TIME IS THE DAY
SYMBOL S REFERS TO COD
SYMBOL X REFERS TO VSS
SYMBOL N REFERS TO NITROGEN
SYMBOL O REFERS TO OXYGEN


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64      C
65      C      DIMENSION STATEMENTS
66      C      -----
67      C
68      PARAMETER NE=1
69      PARAMETER NEE=2
70      PARAMETER ND=241
71      PARAMETER NDD=242
72      COMMON/EXTC/SBS(NE,ND),SU(NE,ND),XA(NE,ND),XE(NE,ND),XI(NE,ND),
73      1XV(NE,ND),XS(NE,ND),NU(NE,ND),NON(NE,ND),XN(NE,ND),NH3(NE,ND),
74      2NO3(NE,ND),SBP(NE,ND),ALK(NE,ND)
75      COMMON/BLOCKC/Q(NEE,NDD),V(NE),SBI(ND),SUI(ND),NIT(NE,ND),
76      1NUI(ND),XII(ND),NONI(ND),NH3I(ND),NO3I(ND),SBPR(ND),ALKI(ND),
77      2XAR(ND),XSR(ND),XER(ND),XIR(ND),XNR(ND),SYNTH(NE,ND),QAV(NEE),
78      3ANPOT(NE,ND),AMON(NE,ND),DSFCTR(NE)
79      CHARACTER IMFT*19/'((20X,I4,(E13.5)))'/,CH*1
80      REAL KVT,KMPT,KSPT,KMST,KSST,KRT,KNT
81      REAL NUI,NOI,NH3I,NO3I,NIT
82      REAL NU,NON,NH3,NO3
83      INTEGER D,DD,HK,TYPE,RESULT
84      DOUBLE PRECISION DT,DDT,RDT
85      DIMENSION STI(ND),STQ(ND),
86      &INIT(NE),ANOX(NE),AROB(NE),IFREQ(NE),INTDN(NE),FANOX(NE)
87      REAL NTI(ND),NTQ(ND),NO3Q(ND),NXII(ND)
88      REAL NTIAV,NUIAV,NOIAV,NO3IAV,NH3IAV,NXIIAV
89      REAL NTQAV,NO3QAV
90      REAL NTIMAX,NO3IAX,NTQMAX,NO3QAX
91      REAL NTI0,NTI00,NO3I0,NO3I00,LFP
92      DIMENSION RA(NE),RN(NE),LRSD(NE),NTDN(NE,ND),ALKQ(ND)
93      REAL KV20,KMP20,KSP20,KMS20,KSS20,KNMAX,KN20,KR20
94      DIMENSION STOSBP(NE,ND),STOXV(NE,ND),STNO3(NE,ND),STOXN(NE,ND)
95      DIMENSION KO(ND),WQ(NE,ND),STOXS(NE,ND),
96      1OC(NE,ND),ON(NE,ND),OT(NE,ND),ST(NE,ND),RXS(NE,ND)
97      REAL NT(NE,ND)
98      DIMENSION SBSAV(NE),ODAV(NE),OCAV(NE),RNO3(NE),ALKAV(NE),AMAV(NE),
99      1ONAV(NE),OTAV(NE),STAV(NE),SUAV(NE),SBPAV(NE),
100     2XAAV(NE),XEAV(NE),XSAV(NE),XIAV(NE),XVAV(NE),
101     3XNAV(NE),RXSAV(NE),NAME(NE),LNAME(3),APTAV(NE),APTNR(NE)
102     REAL NONAV(NE),NTAV(NE),NO3AV(NE),NH3AV(NE),NUAV(NE),NSWAV(NE)
103     REAL NONSS,NUSS,NO3SS,NH3SS
104     C
105     C      INPUT OF DATA
106     C      -----
107     C      TYPE = LOGICAL NUMBER DESCRIBING INFLUENT LOAD PATTERN
108     C      IF = 1   SINE WAVE LOAD PATTERN
109     C      IF = 2   SQUARE WAVE LOAD PATTERN
110     C      IF = 3   GENERAL LOADING PATTERN
111     C      RESULT = LOGICAL NUMBER TO DIRECT OUTPUT
112     C      IF = 1   PRINT RESULTS
113     C      IF = 2   PLOT RESULTS
114     C      IF = 0   PLOT AND PRINT RESULTS
115     C      KOM = TIME AT WHICH TIME BASE COUNTER COMMENCES
116     C      END = NUMBER OF HOURS FOR WHICH PLOTTING IS REQUIRED
117     C      NDH = NUMBER OF NODES OF THE 'KH' INTERVALS REQUIRED
118     C      FOR PLOTTING E.G.: IF PLOTTING IS REQUIRED
119     C      FOR A PERIOD OF 18 HOURS, NDH=18*KH+1
120     C      CRIT = MAXIMUM ERROR ALLOWED BETWEEN CONSECUTIVE DAYS'
121     C      VALUES OF PROCESS VARIABLES TO TEST FOR CONVERGENCE
122     100 FORMAT ( )
123     READ(8,100)TYPE,RESULT,KOM,NDH,CRIT
124     WRITE(5,100)TYPE,RESULT,KOM,NDH,CRIT
125     IF(TYPE.EQ.1) GO TO 111
126     IF(TYPE.EQ.2) GO TO 112
127     IF(TYPE.EQ.3) GO TO 113

```

```

128      111 READ(8,100)PI
129      WRITE(5,100)PI
130      C      VF = VOLUME OF FEED PER DAY
131      C      QO = AVERAGE INFLUENT FLOW FOR SINE WAVE PATTERN
132      C      STIO = INFLUENT COD CONCENTRATION DURING PEAK FLOW PERIOD FOR
133      C      SQUARE WAVE PATTERN OR AVERAGE FOR SINE WAVE PATTERN
134      C      NTIO = INFLUENT TKN CONCENTRATION DURING PEAK FLOW PERIOD FOR
135      C      SQUARE WAVE PATTERN OR AVERAGE FOR SINE WAVE PATTERN
136      C      NO3IO = INFLUENT NO3 CONCENTRATION DURING PEAK FLOW PERIOD FOR
137      C      SQUARE WAVE PATTERN OR AVERAGE FOR SINE WAVE PATTERN
138      C      ALKIO = INFLUENT ALK CONCENTRATION DURING PEAK FLOW PERIOD FOR
139      C      SQUARE WAVE PATTERN OR AVERAGE FOR SINE WAVE PATTERN
140      C      STIOO = INFLUENT COD CONCENTRATION DURING BASE FLOW PERIOD
141      C      NTIOO = INFLUENT TKN CONCENTRATION DURING BASE FLOW PERIOD
142      C      NO3IOO = INFLUENT NO3 CONCENTRATION DURING BASE FLOW PERIOD
143      C      ALKIOO = INFLUENT ALK CONCENTRATION DURING BASE FLOW PERIOD
144      C      LFP = LENGTH OF FEED PERIOD (SQUARE WAVE)
145      C      BQ = BASE FLOW RATE IN SQUARE WAVE PATTERN
146      C      START = INTERVAL BETWEEN 'KOM' AND THE BEGINNING OF THE
147      C      FEEDING PERIOD (HOURS).
148      C      AROB = LENGTH OF AERATED PERIOD (HOURS)
149      C      ANOX = LENGTH OF UNAERATED PERIOD (HOURS)
150      C      INIT = INTERVAL BETWEEN 'KOM' AND 'ANOX' DURING WHICH THE REACTO
151      C      WAS AERATED (HOURS)
152      C      IFREQ = NUMBER OF PAIRS OF UNAERATED-AERATED INTERVALS
153      C
154      READ(8,100)QO,STIO,NTIO,NO3IO,ALKIO,VARQ,VARSTI,VARNTI,VNO3I,VALKI
155      WRITE(5,100)QO,STIO,NTIO,NO3IO,ALKIO,VARQ,VARSTI,VARNTI,VNO3I,VALK
156      GO TO 113
157      112 READ(8,100)VF,STIO,NTIO,NO3IO,ALKIO,BQ,STIOO,NTIOO,NO3IOO,ALKIOO,
158      &LFP,START
159      WRITE(5,100)VF,STIO,NTIO,NO3IO,ALKIO,BQ,STIOO,NTIOO,NO3IOO,ALKIOO,
160      &LFP,START
161      C      KINETIC CONSTANTS
162      C      -----
163      C      AN EXTRA T OR 20 INDICATES VALUES AT T OR 20 DEG. C
164      C      KV = MAX. RATE OF COD TRANSFER FROM SEWAGE INTO STORAGE
165      C      KR = RATE OF CONVERSION OF ORGANIC N TO SALINE NH3
166      C      KM = MAX. RATE OF ORGANISM SYNTHESIS FROM STORED COD
167      C      ('KMS' AND 'KMP' REFER TO EASILY BIODEG.(SOLUBLE) AND
168      C      SLOWLY BIODEG.(PARTICULATE) COD FRACTIONS, RESPECTIVELY)
169      C      KS = SATURATION COEF. FOR EASILY BIODEG. COD UTILIZATION
170      C      UM = MAXIMUM GROWTH RATE OF NITRIFYING ORGANISMS
171      C      KN = SATURATION COEF. FOR NH3 UTILIZATION
172      C      BH = DEATH RATE CONSTANT
173      C      BN = ENDOGENOUS RESPIRATION RATE FOR NITRIFICATION
174      C      THEA = TEMPERATURE DEPENDENCY CONSTANT FOR EASILY BIODEG. COD
175      C      THES = TEMPERATURE DEPENDENCY CONSTANT FOR SYNTHESIS
176      C      THEE = TEMPERATURE DEPENDENCY CONSTANT FOR DEATH
177      C      PHIN = PH DEPENDENCY CONSTANT FOR NITRIFICATION
178      C      THEN = TEMPERATURE DEPENDENCY CONSTANT FOR NITRIFICATION
179      C      THEZ = CONSTANT FOR NO TEMPERATURE DEPENDENCY
180      C      YH = YIELD OF ORGANISMS FROM COD UTILIZED
181      C      YN = ACTIVE MASS YIELD OF NITRIFYING ORGANISMS FROM AMMONIA
182      C      F = UNBIODEG. FRACTION OF ORGANISM
183      C      FR = MAXIMUM FRACTION OF STORED COD (AS VSS) TO ACTIVE
184      C      ORGANISM MASS
185      C      FN = FRACTION OF CARBONACEOUS CELL MASS AS N
186      C      FNS = FRACTION OF NITROGEN ASSOCIATED WITH STORED COD
187      C      P = COD EQUIVALENT OF VSS
188      C      FBS = FRACTION EASILY BIODEG./TOTAL BIODEG. INFL.COD
189      C      FOE = FRACTION OF N RELEASED BY ENDOGENOUS RESPIRATION
190      C      AS ORGANIC NITROGEN
191      C      FOS = FRACTION OF NITROGEN REQUIRED FOR CELL SYNTHESIS

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192 C          AS ORGANIC NITROGEN
193 C          FACT = RATIO BETWEEN KMP IN ANOX. AND AROB. ENVIRONMENT
194 C          FCS = FRACTION OF LYSSED MATERIAL RELEASED AS SBS
195 C
196 C
197 113 READ(8,100) KV20,KMP20,KSP20,KMS20,KSS20,UMAX,KNMAX,BH20,BN20,
198 &KR20,FOS,FBS,FR,YH,YN,F,P,FN,FOE,FACT,FCS
199 READ(8,100)RS,NR,D,NJ,FUS,FUP,UN,SN
200 READ(8,100)THEA,THEM,THES,THEE,PHIN,THEN,THEZ,TEMP,PH
201 READ(8,100) (V(J),LRSD(J),J=1,NR,1)
202 WRITE(5,100) (V(J),LRSD(J),J=1,NR,1)
203 READ(8,100) ICSD,ISSD,NRSD,LINI,LSRI,LARI,LARO,LBRI,LBRO,SR,AR,BR
204 WRITE(5,100) ICSD,ISSD,NRSD,LINI,LSRI,LARI,LARO,LBRI,LBRO,SR,AR,BR
205 READ(8,100) (INIT(J),ANOX(J),AROB(J),IFREQ(J),J=1,NR)
206 KH=D/24
207 NL=D+1
208 NLL=D+2
209 DD=NJ*D
210 NRR=NR+1
211 C
212 C          SEWAGE CHARACTERISTICS
213 C          -----
214 C          NM,NN = LIMITS OF PEAK FLOW PERIOD
215 C          QMAX = MAXIMUM FLOW VALUE OF THE DAY
216 C          STIMAX = MAXIMUM COD CONCENTRATION VALUE OF THE DAY
217 C          NTIMAX = MAXIMUM TKN CONCENTRATION VALUE OF THE DAY
218 C          NO3IAX = MAXIMUM NO3 CONCENTRATION VALUE OF THE DAY
219 C          ALKIAx = MAXIMUM ALK CONCENTRATION VALUE OF THE DAY
220 C          Q = INFLUENT FLOW
221 C          STI = TOTAL INFLUENT COD
222 C          NTI = TOTAL INFLUENT TKN
223 C          STQ = TOTAL INFLUENT COD LOAD
224 C          NTQ = TOTAL INFLUENT TKN LOAD
225 C          NO3Q = TOTAL INFLUENT NO3 LOAD
226 C          ALKQ = TOTAL INFLUENT ALK LOAD
227 C          NONI = INFLUENT ORGANIC NITROGEN CONCENTRATION
228 C          NH3I = INFLUENT AMMONIA CONCENTRATION
229 C          NO3I = INFLUENT NITRATE CONCENTRATION
230 C          ALKI = INFLUENT ALKALINITY CONCENTRATION
231 C          NUI = UNBIODEGRADABLE FRACTION OF SEWAGE N
232 C          NXII = UNSABLE N IN INERT MATERIAL IN INFLUENT
233 C          XII = UNBIODEG. SOLID FRACTION OF SEWAGE COD AS VSS
234 C          SBI = BIODEG. FRACTION OF SEWAGE COD
235 C          SUI = UNBIODEG. SOLUBLE FRACTION OF SEWAGE COD
236 C          FUS = FRACTION OF SOLUBLE UNBIODEG. COD
237 C          FUP = FRACTION OF SOLID UNBIODEG. COD AS VSS
238 C          UN = UNBIODEGRADABLE FRACTION OF N
239 C          SN = FRACTION OF TKN AS AMMONIA
240 C          STQMAX = MAXIMUM COD LOAD VALUE OF THE DAY
241 C          NTQMAX = MAXIMUM TKN LOAD VALUE OF THE DAY
242 C          NO3QAX = MAXIMUM NO3 LOAD VALUE OF THE DAY
243 C          ALKQAX = MAXIMUM ALK LOAD VALUE OF THE DAY
244 C          PQ = PEAK FLOW RATE IN SQUARE WAVE PATTERN
245 C          VARSTI = AMPLITUDE OF INPUT COD CONC. WAVE (SINE WAVE)
246 C          VARNTI = AMPLITUDE OF INPUT TKN CONC. WAVE (SINE WAVE)
247 C          VNO3I = AMPLITUDE OF INPUT NO3 CONC. WAVE (SINE WAVE)
248 C          VALKI = AMPLITUDE OF INPUT ALK CONC. WAVE (SINE WAVE)
249 C          VARQ = AMPLITUDE OF INFLUENT FLOW WAVE (SINE WAVE)
250 C          VARSTL = AMPLITUDE OF INFLUENT COD LOAD WAVE (SINE WAVE)
251 C          VARNTL = AMPLITUDE OF INFLUENT TKN LOAD WAVE (SINE WAVE)
252 C          VRNO3L = AMPLITUDE OF INFLUENT NO3 LOAD WAVE (SINE WAVE)
253 C          VRALKL = AMPLITUDE OF INFLUENT ALK LOAD WAVE (SINE WAVE)
254 C
255 C          PROCESS CHARACTERISTICS

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256 C -----
257 C
258 C TEMP = OPERATING TEMPERATURE OF MIXED LIQUOR
259 C PH = OPERATING PH OF MIXED LIQUOR
260 C RSM = MINIMUM SLUDGE AGE FOR NITRIFICATION
261 C RN = NOMINAL HYDRAULIC RETENTION TIME IN REACTORS
262 C RA = ACTUAL HYDRAULIC RETENTION TIME IN REACTORS
263 C TRN = TOTAL HYDRAULIC RETENTION TIME OF PROCESS
264 C SR = SLUDGE UNDERFLOW RECYCLE RATIO TO AVERAGE INFLUENT FLOW
265 C QSR = SLUDGE UNDERFLOW RECYCLE RATE
266 C AR = MIXED LIQUOR RECYCLE RATIO TO AVERAGE INFLUENT FLOW
267 C QAR = MIXED LIQUOR RECYCLE RATE
268 C NR = NUMBER OF REACTORS IN SERIES
269 C NRR = NUMBER OF REACTORS IN SERIES INCLUDING CLARIFIER
270 C NRSD = NUMBER OF REACTORS FROM WHICH SLUDGE IS WASTED
271 C LRSD = LOGICAL NUMBER DESCRIBING SLUDGE WASTAGE
272 C FROM REACTORS
273 C IF = 1 SLUDGE TO BE WASTED FROM REACTOR
274 C IF = 0 NO SLUDGE TO BE WASTED FROM REACTOR
275 C ICSD = TIME AT WHICH SLUDGE WASTAGE COMMENCES
276 C ISSD = TIME AT WHICH SLUDGE WASTAGE TERMINATES
277 C V = VOLUME OF REACTOR
278 C VP = TOTAL VOLUME OF PROCESS
279 C WQ = SLUDGE WASTE FLOW
280 C N.B. THE SLUDGE WASTE FLOW MAY NEVER BE GREATER THAN
281 C THE INFLUENT FLOW AT ANY INTERVAL OF THE DAY
282 C I.E. THE EFFLUENT FLOW FROM A REACTOR MAY NEVER BE LESS
283 C THAN ZERO AT ANY INTERVAL OF THE DAY
284 C
285 C IF(TYPE.EQ.1) GO TO 115
286 C IF(TYPE.EQ.2) GO TO 116
287 C READ(8,100) (Q(1,I),I=1,NL,KH)
288 C WRITE(5,100) (Q(1,I),I=1,NL,KH)
289 C READ(8,100) (STI(I),I=1,NL,KH)
290 C WRITE(5,100) (STI(I),I=1,NL,KH)
291 C READ(8,100) (NTI(I),I=1,NL,KH)
292 C WRITE(5,100) (NTI(I),I=1,NL,KH)
293 C READ(8,100) (NO3I(I),I=1,NL,KH)
294 C WRITE(5,100) (NO3I(I),I=1,NL,KH)
295 C READ(8,100) (ALKI(I),I=1,NL,KH)
296 C WRITE(5,100) (ALKI(I),I=1,NL,KH)
297 C IF(TYPE.EQ.3) GO TO 117
298 C 115 DO 8 I=1,NL,KH
299 C KL=(I-1)/KH
300 C DUM=SIN(2*PI*(KL-8)/24)
301 C Q(1,I)=Q0*(1+VARQ*DUM)
302 C STI(I)=STI0*(1+VARSTI*DUM)
303 C NTI(I)=NTI0*(1+VARNTI*DUM)
304 C NO3I(I)=NO3I0*(1+VNO3I*DUM)
305 C ALKI(I)=ALKI0*(1+VALKI*DUM)
306 C 8 CONTINUE
307 C
308 C INTERPOLATION OF STI, NO3I, Q AND NTI AT REQUIRED INTERVALS
309 C -----
310 C
311 C 117 DO 25 I=1,24
312 C KI=KH*I+1
313 C KJ=KH*(I-1)+1
314 C DIFSTI=(STI(KI)-STI(KJ))/KH
315 C DIFFQ=(Q(1,KI)-Q(1,KJ))/KH
316 C DIFNTI=(NTI(KI)-NTI(KJ))/KH
317 C DNO3I=(NO3I(KI)-NO3I(KJ))/KH
318 C DALKI=(ALKI(KI)-ALKI(KJ))/KH
319 C DO 25 J=2,KH,1

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320      KK=KJ+J-1
321      STI(KK)=STI(KJ)+(J-1)*DIFSTI
322      Q(1,KK)=Q(1,KJ)+(J-1)*DIFFQ
323      NTI(KK)=NTI(KJ)+(J-1)*DIFNTI
324      NO3I(KK)=NO3I(KJ)+(J-1)*DNO3I
325      ALKI(KK)=ALKI(KJ)+(J-1)*DALKI
326      25 CONTINUE
327      GO TO 118
328      116 PQ=BQ+(VF-BQ)*24.0/LFP
329      DO 24 I=1,NL,1
330      NTI(I)=NTI00
331      STI(I)=STI00
332      Q(1,I)=BQ
333      NO3I(I)=NO3I00
334      ALKI(I)=ALKI00
335      24 CONTINUE
336      NM=INT(START)*KH+1
337      END=START+LFP
338      NN=NM+LFP*KH
339      DO 23 I=NM,NN,1
340      STI(I)=STI0
341      Q(1,I)=PQ
342      NTI(I)=NTI0
343      NO3I(I)=NO3I0
344      ALKI(I)=ALKI0
345      23 CONTINUE
346      118 DO 1 I=1,NL
347      STQ(I)=STI(I)*Q(1,I)
348      NTQ(I)=NTI(I)*Q(1,I)
349      NO3Q(I)=NO3I(I)*Q(1,I)
350      ALKQ(I)=ALKI(I)*Q(1,I)
351      SUI(I)=FUS*STI(I)
352      NUI(I)=UN*NTI(I)
353      XII(I)=FUP*STI(I)
354      NXII(I)=XII(I)*FN
355      NH3I(I)=SN*NTI(I)
356      NONI(I)=NTI(I)-NUI(I)-NXII(I)-NH3I(I)
357      1 SBI(I)=STI(I)*(1.0-FUS-FUP*P)
358      C
359      C      CALCULATION OF AVERAGE SEWAGE CHARACTERISTICS
360      C      -----
361      C
362      SUMQ=0
363      SUMSTI=0
364      SUMNTI=0
365      SNO3I=0.0
366      SALKI=0.0
367      QMAX=0.0
368      STIMAX=0.0
369      SUMSTQ=0.0
370      STQMAX=0.0
371      SUMNTQ=0.0
372      NTQMAX=0.0
373      NTIMAX=0.0
374      SNO3Q=0.0
375      NO3QAX=0.0
376      NO3IAX=0.0
377      SALKQ=0.0
378      ALKQAX=0.0
379      ALKIAX=0.0
380      DO 2 I=1,D,1
381      SUMNTI=SUMNTI+NTI(I)
382      SNO3I=SNO3I+NO3I(I)
383      SALKI=SALKI+ALKI(I)

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384      SUMSTI=SUMSTI+STI(I)
385      SUMSTQ=SUMSTQ+STQ(I)
386      SUMNTQ=SUMNTQ+NTQ(I)
387      SNO3Q=SNO3Q+NO3Q(I)
388      SALKQ=SALKQ+ALKQ(I)
389      IF(STQ(I).GT.STQMAX) STQMAX=STQ(I)
390      IF(NTQ(I).GT.NTQMAX) NTQMAX=NTQ(I)
391      IF(NO3Q(I).GT.NO3QAX) NO3QAX=NO3Q(I)
392      IF(ALKQ(I).GT.ALKQAX) ALKQAX=ALKQ(I)
393      IF(Q(1,I).GT.QMAX) QMAX=Q(1,I)
394      IF(STI(I).GT.STIMAX) STIMAX=STI(I)
395      IF(NTI(I).GT.NTIMAX) NTIMAX=NTI(I)
396      IF(NO3I(I).GT.NO3IAX) NO3IAX=NO3I(I)
397      IF(ALKI(I).GT.ALKIAX) ALKIAX=ALKI(I)
398      2 SUMQ=SUMQ+Q(1,I)
399      QAV(1)=SUMQ/D
400      STQAV=SUMSTQ/D
401      NTQAV=SUMNTQ/D
402      NO3QAV=SNO3Q/D
403      ALKQAV=SALKQ/D
404      IF(TYPE.EQ.2) GO TO 119
405      NTIAV=SUMNTI/D
406      STIAV=SUMSTI/D
407      NO3IAV=SNO3I/D
408      ALKIAV=SALKI/D
409      GO TO 120
410      119 STLAV=STQAV/VF
411      NTIAV=NTQAV/VF
412      NO3IAV=NO3QAV/VF
413      ALKIAV=ALKQAV/VF
414      120 SBIAV=STIAV*(1.0-FUS-FUP*P)
415      SUIAV=FUS*STIAV
416      NUIAV=NTIAV*UN
417      XIIAV=FUP*STIAV
418      NXIIAV=FN*XIIAV
419      NH3IAV=SN*NTIAV
420      NONIAV=NTIAV-NXIIAV-NUIAV-NH3IAV
421      QSR=SR*QAV(1)
422      QAR=AR*QAV(1)
423      QBR=BR*QAV(1)
424      IF(TYPE.EQ.2) GO TO 121
425      IF(TYPE.EQ.3) GO TO 121
426      VARSTL=STQMAX/STQAV-1.00
427      VARNTL=NTQMAX/NTQAV-1.00
428      VRNO3L=NO3QAX/NO3QAV-1.00
429      VRALKL=ALKQAX/ALKQAV-1.00
430      IF(VRNO3L.LT.0.0) VRNO3L=0.0
431      IF(VRALKL.LT.0.0) VRALKL=0.0
432      IF(TYPE.EQ.1) GO TO 121
433      VARQ=QMAX/QAV(1)-1.00
434      VARSTI=STIMAX/STIAV-1.00
435      VARNTI=NTIMAX/NTIAV-1.00
436      VNO3I=NO3IAX/NO3IAV-1.00
437      VALKI=ALKIAX/ALKIAV-1.00
438      IF(VNO3I.LT.0.0) VNO3I=0.0
439      IF(VALKI.LT.0.0) VALKI=0.0
440      121 CONTINUE
441      C
442      C          CALCULATE TOTAL AND AEROBIC VOLUMES
443      C          -----
444      C
445      DO 37 J=1,NR,1
446      DO 37 I=1,NL,1
447      NTDN(J,I)=1

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448 37 CONTINUE
449 VP=0.0
450 DO 133 J=1,NR,1
451 NAMBA=IFREQ(J)
452 VP=VP+V(J)
453 DO 133 I=1,NL,1
454 DO 133 II=1,NAMBA,1
455 IB=(INIT(J)+(II-1)*(ANOX(J)+AROB(J)))*KH+1
456 IE=(INIT(J)+(II-1)*(ANOX(J)+AROB(J))+ANOX(J))*KH+1
457 IF(I.GE.IB.AND.I.LT.IE) NTDN(J,I)=0
458 NTDN(J,NL)=NTDN(J,1)
459 133 CONTINUE
460 JJ=3
461 DO 77 J=1,NR,1
462 INTDN(J)=0
463 DO 77 I=1,NL,1
464 INTDN(J)=INTDN(J)+NTDN(J,I)
465 77 CONTINUE
466 LNAME(1)='AROB'
467 LNAME(2)='ANOX'
468 LNAME(3)='ONOF'
469 SANOX=0.0
470 DO 78 J=1,NR,1
471 IF(INTDN(J).EQ.0) JJ=2
472 IF(INTDN(J).EQ.NL) JJ=1
473 NAME(J)=LNAME(JJ)
474 FANOX(J)=1.0-(FLOAT(INTDN(J)))/(FLOAT(NL))
475 SANOX=SANOX+FANOX(J)*V(J)/VP
476 78 CONTINUE
477 C
478 C ADJUST TEMPERATURE AND PH DEPENDENT CONSTANTS
479 C -----
480 C
481 IF(PH.LT.8.0) UM20=UMAX
482 IF(PH.LT.7.2) UM20=UMAX*(PHIN)**(PH-7.20)
483 IF(PH.LT.8.0) KN20=KNMAX
484 IF(PH.LT.7.2) KN20=KNMAX*(PHIN)**(7.20-PH)
485 IF(TYPE.NE.1) VARNTL=0.0
486 UMT=UM20*(THEN)**(TEMP-20.0)
487 KNT=KN20*(THEN)**(TEMP-20.0)
488 KSPT=KSP20*(THESE)**(20.0-TEMP)
489 KMPT=KMP20*(THESE)**(TEMP-20.0)
490 KSST=KSS20*(THESE)**(TEMP-20.0)
491 DO 17 J=1,NR,1
492 DO 17 I=1,NL,1
493 IF(NTDN(J,I).EQ.0) GO TO 19
494 17 CONTINUE
495 KMST=10.0
496 GO TO 20
497 19 KMST=KMS20*(THEA)**(TEMP-20.0)
498 20 KRT=KR20*(THEE)**(TEMP-20.0)
499 KVT=KV20*(THEE)**(TEMP-20.0)
500 BHT=BH20*(THEE)**(TEMP-20.0)
501 BNT=BN20*(THEE)**(TEMP-20.0)
502 RSM=1.0/((UMT/(1.0+(KNT/(SN*NTIAV))))-BNT)
503 RSA=RS*(1.0-SANOX)
504 TRN=VP/QAV(1)
505 FNS=NONIAV/SBIAV
506 CC=(1.0-F)*BHT*YH*RS/(1.0+BHT*(1.0-YH*P*(1.0-F))*RS)
507 FNS=(FNS*(1.0-FBS)+FN*CC)/((1.0-FBS)+P*CC)
508 C
509 C CALCULATE INITIAL CONDITIONS (PARTICULAR INTEGRAL)
510 C *****
511 CALL PARMOD(YH,FBS,RS,RSA,BHT,KVT,TRN,KMPT,KSPT,KMST,KSST,

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512      1P,F,FN,FNS,KRT,KNT,BNT,UMT,YN,SR,AR,BR,FACT,FR,
513      2SBI,AV,SUI,AV,XII,AV,NUI,AV,NO,NI,AV,NTI,AV,NO,3I,AV,STQ,AV,ALKI,AV,
514      3X,ASS,X,SSS,X,ESS,X,ISS,X,VSS,X,NSS,SBP,SS,SB,SSS,SU,SS,
515      4N,ON,SS,NH,3SS,NO,3SS,NU,SS,ALK,SS,VAR,NTL)
516  C*****
517      FACTC=STQAV/(STI,AV*QAV(1))
518      FACTN=NTQAV/(NTI,AV*QAV(1))
519      XASS=FACTC*XASS
520      XSSS=FACTC*XSSS
521      XESS=FACTC*XESS
522      XISS=FACTC*XISS
523      XVSS=FACTC*XVSS
524      XNSS=FACTN*XNSS
525      CRITXV=CRIT*XVSS
526      IF(CRITXV.LT.CRIT) CRITXV=CRIT
527      CRITXS=CRIT*XSSS
528      IF(CRITXS.LT.CRIT) CRITXS=CRIT
529      CRISBP=CRIT*SBPSS
530      IF(CRISBP.LT.CRIT) CRISBP=CRIT
531      CRINO3=CRIT*NO3SS
532      IF(CRINO3.LT.CRIT) CRINO3=CRIT
533      CRITXN=CRIT*XNSS
534      IF(CRITXN.LT.CRIT) CRITXN=CRIT
535  C
536  C      GENERAL NOTATION FOR PROCESS VARIABLES IN REACTOR
537  C      -----
538  C
539  C      SBP = BIODEGRADABLE PARTICULATE COD
540  C      SBS = BIODEGRADABLE SOLUBLE COD
541  C      SU = UNBIODEGRADABLE COD
542  C      ST = TOTAL COD
543  C      XS = STORED COD
544  C      XA = ACTIVE ORGANISMS
545  C      XE = INERT RESIDUE FROM DECAYING ORGANISMS
546  C      XI = INERT MATERIAL FROM SEWAGE
547  C      XN = ACTIVE NITROSOMONAS
548  C      XV = TOTAL VSS
549  C      OA = OXYGEN CONSUMPTION RATE FOR ADSORPTION OF COD
550  C      OS = OXYGEN CONSUMPTION RATE FOR SYNTHESIS
551  C      OE = OXYGEN CONSUMPTION RATE FOR ENDOGENOUS RESPIRATION
552  C      OC = TOTAL CARBONACEOUS OXYGEN CONSUMPTION RATE
553  C      ON = OXYGEN CONSUMPTION FOR NITRIFICATION
554  C      OT = TOTAL OXYGEN CONSUMPTION RATE
555  C      NSW = NITROGEN REMOVED IN SLUDGE WASTAGE PER DAY
556  C      NT = TOTAL TKN IN REACTOR
557  C      NU = UNBIODEGRADABLE TKN IN REACTOR
558  C      NON = ORGANIC NITROGEN CONCENTRATION IN REACTOR
559  C      NH3 = AMMONIA CONCENTRATION IN REACTOR
560  C      NO3 = NITRATES CONCENTRATION IN REACTOR
561  C      ALK = ALKALINITY CONCENTRATION IN REACTOR
562  C      RXS = RATE OF CHANGE OF STORED COD (AS MG-VSS/L/D)
563  C      SCI = ENERGY ENTERING STORAGE FROM LIQUID PHASE (MG-COD/L)
564  C      SCVO = ENERGY REMOVED FROM STORAGE DUE TO CELL SYNTHESIS (MG-V
565  C
566  C      THE SUFFIX 'AV' AFTER A SYMBOL INDICATES DAILY AVERAGE
567  C      VALUES FOR VARIABLES IN INFLUENT OR REACTORS
568  C      THE SUFFIX 'R' AFTER A SYMBOL INDICATES
569  C      VALUES FOR VARIABLES IN RECYCLE
570  C      THE SUFFIX 'P' AFTER A SYMBOL INDICATES VALUES
571  C      FOR VARIABLES CARRIED OVER TO PLOTTER ROUTINES
572  C      THE SUFFIX 'MAV' AFTER A SYMBOL INDICATES DAILY AVERAGE
573  C      VALUES OF VARIABLES FOR PROCESS
574  C
575  C      BALC = PERCENTAGE COD RECOVERY

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576 C      TCODI = TOTAL COD MASS INPUT
577 C      TCODO = TOTAL COD MASS OUTPUT
578 C      TWCOD = TOTAL COD MASS IN WASTE FLOWS FROM REACTORS
579 C      WCOD = COD MASS IN WASTE FLOW FROM REACTOR
580 C      ECOD = TOTAL COD MASS IN EFFLUENT
581 C      BALN = PERCENTAGE NITROGEN RECOVERY
582 C      TNTI = TOTAL NITROGEN MASS INPUT
583 C      TNTO = TOTAL NITROGEN MASS OUTPUT
584 C      TWN = TOTAL NITROGEN MASS IN WASTE FLOWS FROM REACTORS
585 C      WN = NITROGEN MASS IN WASTE FLOW FROM REACTOR
586 C      EN = TOTAL NITROGEN MASS IN EFFLUENT
587 C
588 DT=1./D
589 DDT=1./DD
590 RDT=DDT/DT
591 C*****
592 CALL DSTRCY(CRIT,VP,XVSS,NR,LINI,LSRI,LARI,LARO,LBRI,LBRO,
593 1QSR,QAR,QBR,SR,AR,BR,DT,DDT,RDT,NLL)
594 C*****
595 C
596 C      RELATE STEP INTERVALS TO THE TIME BASE
597 C      -----
598 C
599 II=0
600 KN=1440/D
601 KHL=KH+1
602 DO 27 I=1,NL,1
603 JE=I-1
604 IF(JE.EQ.0) GO TO 3
605 KO(I)=KO(I-1)+KN
606 GO TO 4
607 3 KO(I)=KOM
608 4 II=II+1
609 IF (II.EQ.KHL) KO(I)=KO(I)+40
610 IF (II.EQ.KHL) II=1
611 IF (KO(I).EQ.2400) KO(I)=0
612 IF(KO(I).EQ.ICSD) IC=I-1
613 IF(KOM.EQ.ICSD) IC=1
614 IF(KO(I).EQ.ISSD) IS=I
615 27 CONTINUE
616 HK=IS-IC
617 DO 9 J=1,NR,1
618 DO 29 I=1,NL,1
619 WQ(J,I)=0.0
620 29 CONTINUE
621 IF(LRSD(J).EQ.0) GO TO 32
622 DO 28 I=IC,IS,1
623 WQ(J,I)=VP/RS*D/HK*1.0/NRSD/DSFCTR(J)
624 28 CONTINUE
625 32 DO 48 I=1,NL,1
626 Q(J+1,I)=Q(J,I)-WQ(J,I)
627 IF(Q(J+1,I).GT.0.00) GO TO 48
628 Q(J+1,I)=0.00
629 WQ(J,I)=Q(J,I)
630 48 CONTINUE
631 9 CONTINUE
632 C
633 C      SET INITIAL CONDITIONS
634 C      -----
635 C
636 DUM=(1.0+SR)/SR
637 DO 7 J=1,NR,1
638 DO 7 I=1,NL,1
639 SBP(J,I)=SBPSS*DSFCTR(J)

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640      SBS(J,I)=SBSSS
641      XS(J,I)=XSSS*DSFCTR(J)
642      XA(J,I)=XASS*DSFCTR(J)
643      XE(J,I)=XESS*DSFCTR(J)
644      XI(J,I)=XISS*DSFCTR(J)
645      XV(J,I)=XVSS*DSFCTR(J)
646      SU(J,I)=SUSS
647      NU(J,I)=NUSS
648      NON(J,I)=NONSS
649      NH3(J,I)=NH3SS
650      XN(J,I)=XNSS*DSFCTR(J)
651      NO3(J,I)=NO3SS
652      ALK(J,I)=ALKSS
653      IF(J.NE.NR) GO TO 7
654      SBPR(I)=SBP(J,I)*DUM
655      XSR(I)=XS(J,I)*DUM
656      XAR(I)=XA(J,I)*DUM
657      XER(I)=XE(J,I)*DUM
658      XIR(I)=XI(J,I)*DUM
659      XNR(I)=XN(J,I)*DUM
660  7 CONTINUE
661      DO 10 J=1,NR,1
662      DO 10 I=1,NL
663      STOSBP(J,I)=SBP(J,I)
664      STNO3(J,I)=NO3(J,I)
665      STOXV(J,I)=XV(J,I)
666      STOXs(J,I)=XS(J,I)
667      STOXN(J,I)=XN(J,I)
668  10 CONTINUE
669      DO 5 J=1,NR,1
670      WRITE(5,200) SBS(J,1),SBP(J,1),XS(J,1),XA(J,1),XE(J,1),XI(J,1),
671      1XV(J,1),XN(J,1),NO3(J,1),ALK(J,1),SU(J,1),NON(J,1),NH3(J,1)
672      5 CONTINUE
673  200 FORMAT(1H ,5X,13F7.2)
674      M=0
675  190 J=0
676  191 J=J+1
677      DO 199 I=1,D,1
678  C*****
679      IF(NTDN(J,I).EQ.1) CALL AROMOD(D,DT,J,I,FR,FBS,FCS,KVT,KMPT,KSPT,
680      1KMST,KSST,P,NR,YH,BHT,F,KRT,FOE,FN,FOS,FNS,NJ,DDT,UMT,YN,KNT,
681      2RDT,BNT,FACT,LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR)
682  C*****
683      IF(NTDN(J,I).EQ.0) CALL ANOMOD(D,DT,J,I,FR,FBS,FCS,KVT,KMPT,KSPT,
684      1KMST,KSST,P,NR,YH,BHT,F,KRT,FOE,FN,FOS,FNS,NJ,DDT,UMT,YN,KNT,
685      2RDT,BNT,FACT,LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR)
686  C*****
687  199 CONTINUE
688      IF(J.EQ.NR) GO TO 192
689      GO TO 191
690  192 CONTINUE
691  C*****
692      CALL SETLCY(NL,J,QSR)
693  C*****
694      DO 18 J=1,NR,1
695      DO 18 I=1,NL
696      IF(DABS(SBP(J,I)-STOSBP(J,I)).GE.CRISBP) GO TO 14
697      IF(DABS(NO3(J,I)-STNO3(J,I)).GE.CRINO3) GO TO 14
698      IF(DABS(XV(J,I)-STOXV(J,I)).GE.CRITXV) GO TO 14
699      IF(DABS(XS(J,I)-STOXs(J,I)).GE.CRITXS) GO TO 14
700      IF(DABS(XN(J,I)-STOXN(J,I)).GE.CRITXN) GO TO 14
701  18 CONTINUE
702      GO TO 15
703  14 M=M+1

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704      IF (M.GE.200) GO TO 15
705      DO 16 J=1,NR,1
706      DO 16 I=1,NL
707      STOSBP(J,I)=SBP(J,I)
708      STNO3(J,I)=NO3(J,I)
709      STOXV(J,I)=XV(J,I)
710      STOXS(J,I)=XS(J,I)
711      STOXN(J,I)=XN(J,I)
712 16 CONTINUE
713      DO 6 J=1,NR,1
714      SBP(J,1)=SBP(J,NL)
715      SBS(J,1)=SBS(J,NL)
716      XS(J,1)=XS(J,NL)
717      XA(J,1)=XA(J,NL)
718      XE(J,1)=XE(J,NL)
719      XI(J,1)=XI(J,NL)
720      SU(J,1)=SU(J,NL)
721      NU(J,1)=NU(J,NL)
722      NON(J,1)=NON(J,NL)
723      NH3(J,1)=NH3(J,NL)
724      XN(J,1)=XN(J,NL)
725      NO3(J,1)=NO3(J,NL)
726      ALK(J,1)=ALK(J,NL)
727      XV(J,1)=XV(J,NL)
728 6 CONTINUE
729      GO TO 190
730 15 CONTINUE
731      DO 26 J=1,NR,1
732      DO 26 I=1,NL
733      JA=I-1
734      IF (JA.EQ.0) JA=D
735      JB=I+1
736      IF (JB.EQ.NLL) JB=2
737      JD=I
738      IF (JD.EQ.NL) JD=1
739      RXS(J,I)=(XS(J,JB)-XS(J,JA))/(DT*2.0)
740      ST(J,I)=SU(J,I)+SBS(J,I)
741      NT(J,I)=NON(J,I)+NU(J,I)+NH3(J,I)
742      OC(J,I)=(1.0-P*YH)*SYNTH(J,JD)/(DT*24.0)
743      ANPOT(J,I)=ANPOT(J,JD)/(DT*24.0)
744      ON(J,I)=4.57*NIT(J,JD)/(DT*24.0)
745      OT(J,I)=OC(J,I)+ON(J,I)
746      AMON(J,I)=AMON(J,JD)/(DT*24.0)
747 26 CONTINUE
748 C
749 C      CALCULATION OF AVERAGE RESULTS FROM DYNAMIC MODEL
750 C      -----
751 C      EXAMPLE:  XVAV = AVERAGE XV
752 C
753      TRN=0.0
754      SUMXVT=0.0
755      SUMWST=0.0
756      SUMXAM=0.0
757      SUMXEM=0.0
758      SUMXIM=0.0
759      SUMXSM=0.0
760      SUMXNM=0.0
761      SUMODM=0.0
762      SMAPTM=0.0
763      APTMNR=0.0
764      RNO3AV=0.0
765      SUMOCM=0.0
766      SUMONM=0.0
767      SUMOTM=0.0

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768      SUMAMM=0.0
769      SUNSWM=0.0
770      TWCOD=0.0
771      TWN=0.0
772      EN=0.0
773      EALK=0.0
774      ECOD=0.0
775      DO 31 J=2,NRR,1
776      SUMQ=0.0
777      DO 30 I=1,D,1
778 30 SUMQ=SUMQ+Q(J,I)
779 31 QAV(J)=SUMQ/D
780      DO 22 J=1,NR,1
781      SUMWS=0.0
782      SUMNSW=0.0
783      SUMNU=0.0
784      SUMXA=0.0
785      SUMXE=0.0
786      SUMXI=0.0
787      SUMSBP=0.0
788      SUMSBS=0.0
789      SUMXV=0.0
790      SUMXS=0.0
791      SUMXN=0.0
792      SUMOC=0.0
793      SUMOD=0.0
794      SUMAPT=0.0
795      SUMON=0.0
796      SUMSU=0.0
797      SUMRXS=0.0
798      SUMNON=0.0
799      SUMNH3=0.0
800      SUMNT=0.0
801      SUMNO3=0.0
802      SUMALK=0.0
803      SUMOT=0.0
804      SUMAM=0.0
805      WCOD=0.0
806      WN=0.0
807      WALK=0.0
808      TWALK=0.0
809      DO 21 I=1,D
810      SUMWS=SUMWS+XV(J,I)*WQ(J,I)*DT
811      SUMNSW=SUMNSW+(FN*(XA(J,I)+XE(J,I)+XI(J,I))+FNS*XS(J,I))
812      1*WQ(J,I)*DT
813      IF(J.NE.NR) GO TO 11
814      ECOD=ECOD+Q(J+1,I)*ST(J,I)*DT
815      EN=EN+Q(J+1,I)*(NT(J,I)+NO3(J,I))*DT
816      EALK=EALK+Q(J+1,I)*ALK(J,I)*DT
817 11 SUMNU=SUMNU+NU(J,I)
818      SUMXA=SUMXA+XA(J,I)
819      SUMXE=SUMXE+XE(J,I)
820      SUMXI=SUMXI+XI(J,I)
821      SUMSBP=SUMSBP+SBP(J,I)
822      SUMSBS=SUMSBS+SBS(J,I)
823      SUMXV=SUMXV+XV(J,I)
824      SUMXN=SUMXN+XN(J,I)
825      SUMXS=SUMXS+XS(J,I)
826      SUMOC=SUMOC+OC(J,I)
827      SUMAPT=SUMAPT+ANPOT(J,I)
828      IF(NTDN(J,I).EQ.1) GO TO 12
829      SUMOD=SUMOD+OC(J,I)
830 12 SUMON=SUMON+ON(J,I)
831      SUMSU=SUMSU+SU(J,I)

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832      SUMRXS=SUMRXS+RXS(J,I)
833      SUMNO3=SUMNO3+NO3(J,I)
834      SUMALK=SUMALK+ALK(J,I)
835      SUMNON=SUMNON+NON(J,I)
836      SUMNH3=SUMNH3+NH3(J,I)
837      SUMNT=SUMNT+NT(J,I)
838      WCOD=WCOD+WQ(J,I)*(ST(J,I)+SBP(J,I))*DT
839      WN=WN+WQ(J,I)*(NT(J,I)+NO3(J,I))*DT
840      WALK=WALK+WQ(J,I)*ALK(J,I)*DT
841      SUMOT=SUMOT+OT(J,I)
842      SUMAM=SUMAM+AMON(J,I)
843      21 CONTINUE
844      NUAV(J)=SUMNU/D
845      XAAV(J)=SUMXA/D
846      XEAV(J)=SUMXE/D
847      XIAV(J)=SUMXI/D
848      SBPAV(J)=SUMSBP/D
849      SBSAV(J)=SUMSBS/D
850      XVAV(J)=SUMXV/D
851      XSAV(J)=SUMXS/D
852      XNAV(J)=SUMXN/D
853      OCAV(J)=SUMOC/D
854      ODAV(J)=SUMOD/D
855      APTAV(J)=SUMAPT/D
856      APTNR(J)=APTAV(J)*24.0*V(J)/QAV(1)
857      RNO3(J)=ODAV(J)*24.0*V(J)/(QAV(1)*2.85)
858      ONAV(J)=SUMON/D
859      SUAV(J)=SUMSU/D
860      RXSAV(J)=SUMRXS/D
861      STAV(J)=SUAV(J)+SBSAV(J)
862      NSWAV(J)=SUMNSW/QAV(1)
863      SUNSWM=SUNSWM+NSWAV(J)
864      OTAV(J)=SUMOT/D
865      AMAV(J)=SUMAM/D
866      NO3AV(J)=SUMNO3/D
867      ALKAV(J)=SUMALK/D
868      NONAV(J)=SUMNON/D
869      NH3AV(J)=SUMNH3/D
870      NTAV(J)=SUMNT/D
871      SUMWST=SUMWST+SUMWS
872      SUMXVT=SUMXVT+XVAV(J)*V(J)
873      SUMXAM=SUMXAM+XAAV(J)*V(J)
874      SUMXEM=SUMXEM+XEAV(J)*V(J)
875      SUMXIM=SUMXIM+XIAV(J)*V(J)
876      SUMXSM=SUMXSM+XSAV(J)*V(J)
877      SUMXNM=SUMXNM+XNAV(J)*V(J)
878      SUMODM=SUMODM+ODAV(J)*V(J)
879      SMAPTM=SMAPTM+APTAV(J)*V(J)
880      APTMNR=APTMNR+APTNR(J)
881      RNO3AV=RNO3AV+RNO3(J)
882      SUMOCM=SUMOCM+OCAV(J)*V(J)
883      SUMONM=SUMONM+ONAV(J)*V(J)
884      SUMOTM=SUMOTM+OTAV(J)*V(J)
885      SUMAMM=SUMAMM+AMAV(J)*V(J)
886      I=NLL
887      Q(J,I)=QAV(J)
888      C*****
889      CALL DUMVCY(J,I,DUMIF,DUMS,DUMA,DUMB,DUMIR,DUMO,
890      1LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR,
891      1DT,DDT,RDT)
892      C*****
893      RA(J)=24.0*DDT/DUMO
894      RN(J)=V(J)/QAV(J)*24.0
895      TWCOD=TWCOD+WCOD

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896      TWN=TWN+WN
897      TWALK=TWALK+WALK
898      TRN=TRN+RN (J)
899  22  CONTINUE
900      RSAV=SUMXVT/SUMWST
901      XAMAV=SUMXAM/VP
902      XEMAV=SUMXEM/VP
903      XIMAV=SUMXIM/VP
904      XSMVAV=SUMXSM/VP
905      XVMVAV=SUMXVT/VP
906      XNMVAV=SUMXNM/VP
907      ODMVAV=SUMODM/VP
908      APTMAV=SMAPTM/VP
909      OCMVAV=SUMOCM/VP
910      ONMAV=SUMONM/VP
911      OTMAV=SUMOTM/VP
912      AMMAV=SUMAMM/VP
913  C
914  C      CHECK MASS BALANCES ON COD , NITROGEN AND ALKALINITY
915  C      -----
916  C  1.  COD
917  C  -----
918      TCODI=STQAV
919      TCODO=P*SUMWST+SUMOCM*24.0+TWCOD+ECOD
920      BALC=TCODO*100.0/TCODI
921  C  2.  NITROGEN
922  C  -----
923      TNTI=NTQAV
924      TNTO=SUNSWM*QAV(1)+TWN+EN+SUMODM*24/2.85
925      BALN=TNTO*100.0/TNTI
926  C  3.  ALKALINITY
927  C  -----
928      TALKI=ALKQAV
929      TALKO=TWALK+EALK
930      &+3.57*24.0*(2.0*SUMONM/4.57-SUMODM/2.86-SUMAMM)
931      BALALK=TALKO/TALKI*100.0
932  C
933  C      OUTPUT AND FORMAT STATEMENTS
934  C      -----
935  C
936      IF(RESULT.EQ.2) GO TO 126
937      WRITE(5,201)TEMP,PH,YH,BHT,KVT,YN,THEN,F,FBS,FN,KNT,PHIN,
938      1THEE,P,FOE,UMT,BNT,FOS,KRT,KSPT,THEZ,KMPT,THEA,THES,
939      2KMST,KSST,FR,FNS,RSM,RS,RSA
940  201  FORMAT(1H1,20X,22(1H*),/,1H ,20X,22(1H*),/,
941      11H ,20X,22(1H*),/,
942      11H0,10X,'KINETIC CONSTANTS',/,1H ,10X,17(1H-),/,
943      21H ,5X,'PROCESS OPERATING TEMPERATURE=',F5.1,1X,'DEG. C',/,
944      31H ,28X,'AND PH=',F5.2,/,1H0,5X,'HETEROTROPHS',28X,'AUTOTROPHS',/,
945      41H ,5X,12(1H-),28X,10(1H-),/,1H ,6X,'YH=',F5.3,4X,'BH=',F5.3,4X,
946      5'KV=',F5.3,8X,'YN=',F5.3,6X,'THEN=',F5.3,/,1H ,7X,'F=',F5.3,3X,
947      6'FBS=',F5.3,4X,'FN=',F5.3,8X,'KN=',F5.2,6X,'PHIN=',F5.3,/,
948      75X,'THEE=',F5.3,5X,'P=',F5.3,3X,'FOE=',F5.3,8X,'UM=',F5.3,
949      28X,'BN=',F5.3,/,
950      86X,'FOS=',F5.3,4X,'KR=',F5.3,3X,'KSP=',F5.3,6X,'THEZ=',F5.3,/,
951      16X,'KMP=',F5.3,2X,'THEA=',F5.3,2X,'THES=',F5.3,/,
952      26X,'KMS=',F5.2,3X,'KSS=',F5.1,/,
953      27X,'FR=',F5.3,3X,'FNS=',F5.3,/,
954      91H0,5X,'MINIMUM SLUDGE AGE FOR NITRIFICATION=',F5.2,1X,'DAYS',/,
955      11H ,5X,'OPERATING SLUDGE AGE OF PROCESS=',F5.2,1X,'DAYS',/,
956      11H ,5X,'AEROBIC SLUDGE AGE OF PROCESS=',F5.2,1X,'DAYS')
957      WRITE(5,202)FUP,FUS,SN,UN,QAV(1),STQAV,NTQAV,STIAV,SBIAV,SUIAV,
958      1XIIAV,NTIAV,NH3IAV,NONIAV,NUIAV,NO3IAV,ALKIAV
959  202  FORMAT(1H0,10X,'AVERAGE COMPOSITION OF SEWAGE',/,

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960      11H ,10X,29(1H-),/,1H0,5X,'SOLID INERT FRACTION OF INFLUENT COD=',
961      2F5.3,/,1H ,5X,'SOLUBLE INERT FRACTION OF INFLUENT COD=',F5.3,/,
962      31H ,5X,'AMMONIA TO TKN FRACTION=',F5.3,/,
963      41H ,5X,'INERT TKN FRACTION=',F5.3,/,1H0,4X,'QAV=',F9.5,
964      15X,'STQAV=',E13.5,5X,'NTQAV=',E13.5,/,
965      51H ,5X,'STI=',F7.2,5X,'SBI=',F7.2,4X,'SUI=',F6.2,
966      64X,'XII=',F6.2,/,1H ,5X,'NTI=',F6.2,4X,'NH3I=',F6.2,
967      74X,'NONI=',F5.2,5X,'NUI=',F5.2,5X,'NO3I=',F5.2,/,
968      &4X,'ALKI=',F7.2)
969      IF(TYPE.EQ.1) GO TO 122
970      IF(TYPE.EQ.2) GO TO 123
971      IF(TYPE.EQ.3) GO TO 122
972 122 WRITE(5,205)VARQ,VARSTI,VARNTI,VNO3I,VALKI,VARSTL,VARNTL,VNO3L,
973      &VRALKL
974 205 FORMAT(1H0,10X,'SINE WAVE INFLUENT FLOW AND FEED PATTERN',/,
975      11H ,10X,40(1H-),/,
976      41H0,10X,'AMPLITUDE OF FLOW WAVE=',F5.2,4X,'(FRAC OF AVE)',/,
977      51H ,10X,'AMPLITUDE OF COD WAVE=',F5.2,4X,'(FRAC OF AVE)',/,
978      61H ,10X,'AMPLITUDE OF TKN WAVE=',F5.2,4X,'(FRAC OF AVE)',/,
979      71H ,10X,'AMPLITUDE OF NO3 WAVE=',F5.2,4X,'(FRAC OF AVE)',/,
980      71H ,10X,'AMPLITUDE OF ALK WAVE=',F5.2,4X,'(FRAC OF AVE)',/,
981      81H ,10X,'AMPLITUDE OF COD LOAD WAVE=',F5.2,4X,'(FRAC OF AVE)',/,
982      81H ,10X,'AMPLITUDE OF TKN LOAD WAVE=',F5.2,4X,'(FRAC OF AVE)',/,
983      81H ,10X,'AMPLITUDE OF NO3 LOAD WAVE=',F5.2,4X,'(FRAC OF AVE)',/,
984      81H ,10X,'AMPLITUDE OF ALK LOAD WAVE=',F5.2,4X,'(FRAC OF AVE)')
985      WRITE(5,204) NR,LINI,LSRI,LARO,LARI,LBRO,LBRI,SR,AR,BR
986 204 FORMAT(1H0,10X,'PROCESS CONFIGURATION DATA',/,1H ,5X,26(1H-),/,
987      11H0,5X,'NUMBER OF TANKS IN SERIES',I2,/,
988      11H ,5X,'INFLUENT FLOW INTO TANK',I2,/,
989      11H ,5X,'S-RECYCLE FROM SETTLER TO TANK ',I2,/,
990      11H ,5X,'A-RECYCLE FROM TANK',I2,1X,'INTO TANK',I2,/,
991      11H ,5X,'B-RECYCLE FROM TANK',I2,1X,'INTO TANK',I2,/,
992      11H ,5X,'S RECYCLE RATIO=',F4.1,/,
993      11H ,5X,'A RECYCLE RATIO=',F4.1,/,
994      11H ,5X,'B RECYCLE RATIO=',F4.1)
995      GO TO 124
996 123 WRITE(5,206)LFP,PQ,BQ,VF
997 206 FORMAT(1H0,10X,'SQUARE WAVE INFLUENT FLOW AND FEED PATTERN',/,
998      11H ,10X,42(1H-),/,
999      11H ,10X,'LENGTH OF FEED PERIOD (HRS)=',F5.2,/,
1000     21H ,10X,'PEAK FLOW (L/D)=',F9.5,/,
1001     31H ,10X,'BASE FLOW (L/D)=',F9.5,/,
1002     41H ,10X,'VOLUME OF FEED PER DAY (LITRES)=' F9.5)
1003 124 WRITE(5,203) TRN,VP,SANOX,M,RSAB
1004 203 FORMAT(1H1,10X,21(1H-),/,1H ,10X,'DAILY AVERAGE RESULTS',/,
1005     11H ,10X,21(1H-),/,
1006     11H0,5X,'TOTAL HYDRAULIC RETENTION TIME=',F8.3,1X,'HOURS',/,
1007     11H ,5X,'TOTAL VOLUME=',E13.5,1X,'LITRES',/,
1008     11H ,5X,'ANOXIC FRACTION=',F6.2,1X,/,
1009     21H ,5X,'NO. OF DAYS TO REACH DYNAMIC STEADY STATE=',I3,/,
1010     21H ,5X,'SLUDGE AGE =' ,F5.2,1X,'DAYS')
1011     WRITE(5,244)TCODI,TCODO,BALC,TNTI,TNTO,BALN,TALKI,TALKO,BALALK
1012 244 FORMAT(1H0,10X,'MASS BALANCES ON COD AND NITROGEN',/,
1013     11H ,10X,33(1H-),/,
1014     21H0,5X,'TOTAL COD MASS INPUT=',1X,E13.6,1X,'(MG-COD/D)',/,
1015     21H ,5X,'TOTAL COD MASS OUTPUT=',1X,E13.6,1X,'(MG-COD/D)',/,
1016     31H ,5X,'PERCENTAGE COD RECOVERY=',1X,F7.3,1X,'(%)',/,
1017     21H ,5X,'TOTAL NITROGEN MASS INPUT=',1X,E13.6,1X,'(MG-N/D)',/,
1018     21H ,5X,'TOTAL NITROGEN MASS OUTPUT=',1X,E13.6,1X,'(MG-N/D)',/,
1019     31H ,5X,'PERCENTAGE NITROGEN RECOVERY=',1X,F7.3,1X,'(%)',/,
1020     21H ,5X,'TOTAL ALKALINITY MASS INPUT=',1X,E13.6,1X,'(CACO3/D)',/,
1021     21H ,5X,'TOTAL ALKALINITY MASS OUTPUT=',1X,E13.6,1X,'(CACO3/D)',/,
1022     31H ,5X,'PERCENTAGE ALKALINITY RECOVERY=',1X,F7.3,1X,'(%)')
1023     WRITE(5,300)

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1024 300 FORMAT(1H0,30X,'TANK 1',6X,'TANK 2',
1025      26X,'TANK 3',6X,'TANK 4',6X,'TANK 5',6X,'TANK 6',
1026      36X,'MEAN',/,1H,30X,7(1H-),6X,7(1H-),6X,7(1H-),6X,7(1H-),
1027      46X,7(1H-),6X,7(1H-),6X,4(1H-))
1028      WRITE(5,327) (NAME(J),J=1,NR,1)
1029 327 FORMAT(1H,23X,6A13)
1030      WRITE(5,328)
1031 328 FORMAT(1H,30X,6(1H-),7X,6(1H-),7X,6(1H-),
1032      17X,6(1H-),7X,6(1H-),7X,6(1H-))
1033      WRITE(5,301) (XAAV(J),J=1,NR,1),XAMAV
1034      WRITE(5,302) (XSAV(J),J=1,NR,1),XSMAY
1035      WRITE(5,303) (XEAV(J),J=1,NR,1),XEMAV
1036      WRITE(5,304) (XIAV(J),J=1,NR,1),XIMAV
1037      WRITE(5,305) (XVAV(J),J=1,NR,1),XVMAV
1038      WRITE(5,319) (XNAV(J),J=1,NR,1),XNMAV
1039      WRITE(5,313) (ODAV(J),J=1,NR,1),ODMAV
1040      WRITE(5,314) (OCAV(J),J=1,NR,1),OCMAV
1041      WRITE(5,315) (ONAV(J),J=1,NR,1),ONMAV
1042      WRITE(5,316) (OTAV(J),J=1,NR,1),OTMAV
1043      WRITE(5,3160) (AMAV(J),J=1,NR,1),AMMAV
1044      WRITE(5,309) (NSWAV(J),J=1,NR,1),SUNSWM
1045      WRITE(5,321) (SBPAV(J),J=1,NR,1)
1046      WRITE(5,320) (SBSAV(J),J=1,NR,1)
1047      WRITE(5,324) (SUAV(J),J=1,NR,1)
1048      WRITE(5,311) (STAV(J),J=1,NR,1)
1049      WRITE(5,306) (NONAV(J),J=1,NR,1)
1050      WRITE(5,310) (NH3AV(J),J=1,NR,1)
1051      WRITE(5,323) (NUAV(J),J=1,NR,1)
1052      WRITE(5,312) (NTAV(J),J=1,NR,1)
1053      WRITE(5,307) (NO3AV(J),J=1,NR,1)
1054      WRITE(5,703) (ALKAV(J),J=1,NR,1)
1055      WRITE(5,308) (RNO3(J),J=1,NR,1),RNO3AV
1056      WRITE(5,326) (APTAV(J),J=1,NR,1),APTMAY
1057      WRITE(5,329) (APTNR(J),J=1,NR,1),APTMNR
1058      WRITE(5,3329) (FANOX(J),J=1,NR,1),SANOX
1059      WRITE(5,322) (V(J),J=1,NR,1)
1060      WRITE(5,325) (QAV(J),J=1,NRR,1)
1061      WRITE(5,317) (RA(J),J=1,NR,1)
1062      WRITE(5,318) (RN(J),J=1,NR,1)
1063 301 FORMAT(1H,8X,'XA (MG-VSS/L)',2X,7(E13.5))
1064 302 FORMAT(1H,8X,'XS (MG-VSS/L)',2X,7(E13.5))
1065 303 FORMAT(1H,8X,'XE (MG-VSS/L)',2X,7(E13.5))
1066 304 FORMAT(1H,8X,'XI (MG-VSS/L)',2X,7(E13.5))
1067 305 FORMAT(1H,8X,'XV (MG-VSS/L)',2X,7(E13.5))
1068 319 FORMAT(1H,8X,'XN (MG-VSS/L)',2X,7(E13.5))
1069 313 FORMAT(1H,8X,'OD (MG-O/L/HR)',2X,7(E13.5))
1070 314 FORMAT(1H,8X,'OC (MG-O/L/HR)',2X,7(E13.5))
1071 315 FORMAT(1H,8X,'ON (MG-O/L/HR)',2X,7(E13.5))
1072 316 FORMAT(1H,8X,'OT (MG-O/L/HR)',2X,7(E13.5))
1073 3160 FORMAT(1H,8X,'AM (MG-N/L/HR)',2X,7(E13.5))
1074 309 FORMAT(1H,8X,'NSW (MG-N/L/D)',2X,7(E13.5))
1075 321 FORMAT(1H,8X,'SBP (MG-COD/L)',2X,6(E13.5))
1076 320 FORMAT(1H,8X,'SBS (MG-COD/L)',2X,6(E13.5))
1077 324 FORMAT(1H,8X,'SU (MG-COD/L)',2X,6(E13.5))
1078 311 FORMAT(1H,8X,'ST (MG-COD/L)',2X,6(E13.5))
1079 306 FORMAT(1H,8X,'NON (MG-N/L)',2X,6(E13.5))
1080 310 FORMAT(1H,8X,'NH3 (MG-N/L)',2X,6(E13.5))
1081 323 FORMAT(1H,8X,'NU (MG-N/L)',2X,6(E13.5))
1082 312 FORMAT(1H,8X,'NT (MG-N/L)',2X,6(E13.5))
1083 307 FORMAT(1H,8X,'NO3 (MG-N/L)',2X,6(E13.5))
1084 703 FORMAT(1H,8X,'ALK (MG-CACO3/L)',1X,6(E13.5))
1085 326 FORMAT(1H,8X,'APT (MG-N/L/HR)',2X,7(E13.5))
1086 329 FORMAT(1H,8X,'APTNR (MG-N/L)',2X,7(E13.5))
1087 3329 FORMAT(1H,8X,'ANOX. FRACTION',2X,7(E13.5))

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1088 308 FORMAT(1H ,8X,'RNO3      (MG-N/L)',2X,7(E13.5))
1089 322 FORMAT(1H ,8X,'VOL      (LITRES)',2X,6(E13.5))
1090 325 FORMAT(1H ,8X,'FLOW      (L/D)',2X,7(E13.5))
1091 317 FORMAT(1H ,8X,'AHRT      (HOURS)',2X,6(E13.5))
1092 318 FORMAT(1H ,8X,'NHRT      (HOURS)',2X,6(E13.5))
1093 WRITE(CH,'(I1)')NR
1094 IMFT(10:10)=CH
1095 WRITE(5,399) (KO(I),STQ(I),NTQ(I),Q(1,I),SBI(I),STI(I),
1096 INTI(I),SUI(I),NUI(I),XII(I),NH3I(I),NONI(I),I=1,NL,KH)
1097 399 FORMAT(1H1,10X,'DYNAMIC INPUTS',/,1H ,10X,14(1H-),/,
1098 11H ,2X,'TIME',2X,'COD LOAD',2X,'TKN LOAD',4X,'FLOW',7X,'SBI',7X,
1099 2'STI',7X,'NTI',7X,'SUI',7X,'NUI',7X,'XII',6X,'NH3I',6X,'NONI',/,
1100 31H ,2X,4(1H-),2X,8(1H-),3X,8(1H-),4X,4(1H-),7X,3(1H-),7X,3(1H-),
1101 47X,3(1H-),7X,3(1H-),7X,3(1H-),7X,3(1H-),6X,4(1H-),6X,4(1H-),/,
1102 5(1H ,2X,I4,11(E10.4)))
1103 WRITE(5,400)
1104 400 FORMAT(1H1,10X,21(1H-),/,1H ,10X,'DAILY DYNAMIC RESULTS',/,
1105 11H ,10X,21(1H-),/,
1106 11H0,20X,'TIME',5X,'TANK 1',6X,'TANK 2',
1107 26X,'TANK 3',6X,'TANK 4',6X,'TANK 5',6X,'TANK 6',/,
1108 31H ,20X,4(1H-),5X,7(1H-),6X,7(1H-),6X,7(1H-),6X,7(1H-),
1109 46X,7(1H-),6X,7(1H-))
1110 WRITE(5,401)
1111 401 FORMAT(1H0,3X,'XA      (MG-VSS/L)')
1112 WRITE(5,IMFT) (KO(I),(XA(J,I),J=1,NR,1),I=1,NL,KH)
1113 WRITE(5,402)
1114 402 FORMAT(1H0,3X,'XS      (MG-VSS/L)')
1115 WRITE(5,IMFT) (KO(I),(XS(J,I),J=1,NR,1),I=1,NL,KH)
1116 WRITE(5,403)
1117 403 FORMAT(1H0,3X,'XE      (MG-VSS/L)')
1118 WRITE(5,IMFT) (KO(I),(XE(J,I),J=1,NR,1),I=1,NL,KH)
1119 WRITE(5,404)
1120 404 FORMAT(1H0,3X,'XI      (MG-VSS/L)')
1121 WRITE(5,IMFT) (KO(I),(XI(J,I),J=1,NR,1),I=1,NL,KH)
1122 WRITE(5,405)
1123 405 FORMAT(1H0,3X,'XV      (MG-VSS/L)')
1124 WRITE(5,IMFT) (KO(I),(XV(J,I),J=1,NR,1),I=1,NL,KH)
1125 WRITE(5,419)
1126 419 FORMAT(1H0,3X,'XN      (MG-VSS/L)')
1127 WRITE(5,IMFT) (KO(I),(XN(J,I),J=1,NR,1),I=1,NL,KH)
1128 WRITE(5,407)
1129 407 FORMAT(1H0,3X,'OC      (MG-O/L/HR)')
1130 WRITE(5,IMFT) (KO(I),(OC(J,I),J=1,NR,1),I=1,NL,KH)
1131 WRITE(5,408)
1132 408 FORMAT(1H0,3X,'ON      (MG-O/L/HR)')
1133 WRITE(5,IMFT) (KO(I),(ON(J,I),J=1,NR,1),I=1,NL,KH)
1134 WRITE(5,409)
1135 409 FORMAT(1H0,3X,'OT      (MG-O/L/HR)')
1136 WRITE(5,IMFT) (KO(I),(OT(J,I),J=1,NR,1),I=1,NL,KH)
1137 WRITE(5,4090)
1138 4090 FORMAT(1H0,3X,'AM      (MG-N/L/HR)')
1139 WRITE(5,IMFT) (KO(I),(AMON(J,I),J=1,NR,1),I=1,NL,KH)
1140 WRITE(5,417)
1141 417 FORMAT(1H0,3X,'WQ      (L/DAY)')
1142 WRITE(5,IMFT) (KO(I),(WQ(J,I),J=1,NR,1),I=1,NL,KH)
1143 WRITE(5,411)
1144 411 FORMAT(1H0,3X,'SBP      (MG-COD/L)')
1145 WRITE(5,IMFT) (KO(I),(SBP(J,I),J=1,NR,1),I=1,NL,KH)
1146 WRITE(5,418)
1147 418 FORMAT(1H0,3X,'SBS      (MG-COD/L)')
1148 WRITE(5,IMFT) (KO(I),(SBS(J,I),J=1,NR,1),I=1,NL,KH)
1149 WRITE(5,420)
1150 420 FORMAT(1H0,3X,'SU      (MG-COD/L)')
1151 WRITE(5,IMFT) (KO(I),(SU(J,I),J=1,NR,1),I=1,NL,KH)

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1152      WRITE(5,412)
1153      412 FORMAT(1H0,3X,'ST      (MG-COD/L)')
1154      WRITE(5,IMFT) (KO(I),(ST(J,I),J=1,NR,1),I=1,NL,KH)
1155      WRITE(5,413)
1156      413 FORMAT(1H0,3X,'NON      (MG-N/L)')
1157      WRITE(5,IMFT) (KO(I),(NON(J,I),J=1,NR,1),I=1,NL,KH)
1158      WRITE(5,421)
1159      421 FORMAT(1H0,3X,'NU      (MG-N/L)')
1160      WRITE(5,IMFT) (KO(I),(NU(J,I),J=1,NR,1),I=1,NL,KH)
1161      WRITE(5,415)
1162      415 FORMAT(1H0,3X,'NH3      (MG-N/L)')
1163      WRITE(5,IMFT) (KO(I),(NH3(J,I),J=1,NR,1),I=1,NL,KH)
1164      WRITE(5,414)
1165      414 FORMAT(1H0,3X,'NT      (MG-N/L)')
1166      WRITE(5,IMFT) (KO(I),(NT(J,I),J=1,NR,1),I=1,NL,KH)
1167      WRITE(5,422)
1168      422 FORMAT(1H0,3X,'APT (MG-N/L/HR)')
1169      WRITE(5,IMFT) (KO(I),(ANPOT(J,I),J=1,NR,1),I=1,NL,KH)
1170      WRITE(5,416)
1171      416 FORMAT(1H0,3X,'NO3      (MG-N/L)')
1172      WRITE(5,IMFT) (KO(I),(NO3(J,I),J=1,NR,1),I=1,NL,KH)
1173      WRITE(5,461)
1174      461 FORMAT(1H0,3X,'ALK      (MG-CACO3/L)')
1175      WRITE(5,IMFT) (KO(I),(ALK(J,I),J=1,NR,1),I=1,NL,KH)
1176      IF(RESULT.NE.0) GO TO 127
1177      C -----
1178      C      SET THE SOLUBLE EFFLUENT CONCENTRATIONS THAT ARE
1179      C
1180      C      TO BE WRITTEN TO THE PLOT FILE
1181      C -----
1182      126 ECOD=STAV(NR)
1183      ENT=NTAV(NR)
1184      ENH3=NH3AV(NR)
1185      ENO3=NO3AV(NR)
1186      EALK=ALKAV(NR)
1187      C -----
1188      C
1189      C      WRITE ALL THE PARAMETERS USED IN PLOTTING OF
1190      C
1191      C      SIMULATED RESPONSE TO THE PRINT FILE ( NO. 18)
1192      C -----
1193      WRITE(18,522)ECOD,ENT,ENH3,ENO3,EALK
1194      WRITE(18,522)KOM,D,INIT(1),ANOX(1),AROB(1),IFREQ,NDH
1195      522 FORMAT()
1196      WRITE(18,45)(OT(1,I),XV(1,I),ST(1,I),NT(1,I),NH3(1,I),NO3(1,I),
1197      &ALK(1,I),I=1,NDH)
1198      WRITE(18,522) START,END
1199      45  FORMAT(7E13.5)
1200      ENDFILE 18
1201      127 STOP
1202      END

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1 C*****
2 SUBROUTINE PARMOD(YH,FBS,RS,RSA,BHT,KVT,TRN,KMPT,KSPT,KMST,KSST,
3 1P,F,FN,FNS,KRT,KNT,BNT,UMT,YN,SR,AR,BR,FACT,FR,
4 2SBIAV,SUIAV,XIIAV,NUIAV,NONIAV,NTIAV,NO3IAV,STQAV,ALKIAV,
5 3XASS,XSSS,XESS,XISS,XVSS,XNSS,SBPSS,SBSSS,SUSS,
6 4NONSS,NH3SS,NO3SS,NUSS,ALKSS,VARNTL)
7 C*****
8 C THIS SUBROUTINE CALCULATES THE STEADY STATE
9 C SOLUTION ( PARTICULAR INTEGRAL )
10 C*****
11 PARAMETER NE=1
12 PARAMETER NEE=2
13 PARAMETER ND=241
14 PARAMETER NDD=242
15 REAL NUIAV,NONIAV,NTIAV,NO3IAV
16 REAL NH3MAX,NSW
17 COMMON/EXTC/SBS(NE,ND),SU(NE,ND),XA(NE,ND),XE(NE,ND),XI(NE,ND),
18 1XV(NE,ND),XS(NE,ND),NU(NE,ND),NON(NE,ND),XN(NE,ND),NH3(NE,ND),
19 2NO3(NE,ND),SBP(NE,ND),ALK(NE,ND)
20 COMMON/BLOCKC/Q(NEE,NDD),V(NE),SBI(ND),SUI(ND),NIT(NE,ND),
21 1NUI(ND),XII(ND),NONI(ND),NH3I(ND),NO3I(ND),SBPR(ND),ALKI(ND),
22 2XAR(ND),XSR(ND),XER(ND),XIR(ND),XNR(ND),SYNTH(NE,ND),QAV(NEE),
23 3ANPOT(NE,ND),AMON(NE,ND),DSFCTR(NE)
24 REAL KVT,KMPT,KSPT,KMST,KSST,KRT,KNT,KMPTEF
25 REAL NUI,NONI,NH3I,NO3I,NIT
26 REAL NU,NON,NH3,NO3
27 REAL NUSS,NONSS,NH3SS,NO3SS
28 KMPTEF=KMPT*(FACT*(1.0-RSA/RS)+RSA/RS)
29 XA0=YH*RS/(TRN*(1.+BHT*RS*(1.-P*YH*(1.-F))))*SBIAV
30 SBSI=FBS*SBIAV
31 SBPI=SBIAV-SBSI
32 SBS0=0.0
33 SBP0=0.0
34 XS0=0.0
35 DO 15 KLM=1,100,1
36 C6=SBSI-KSST-TRN*KMST*XA0
37 SBSS=(C6+(C6*C6+4.0*KSST*SBSI)**0.5)/2.0
38 SBPSS=(SBPI/TRN+(1.0-F)*BHT*P*XA0)/(1.0/RS+KVT*XA0*(FR-XS0/XA0))
39 C1=-(P/RS+KVT*SBPSS)/XA0
40 C2=KVT*SBPSS*(P-KSPT)/P-KMPTEF-KSPT/RS
41 C3=KVT*SBPSS*KSPT*XA0/P
42 XSSS=(-C2-(C2*C2-4.0*C1*C3)**0.5)/(2.0*C1)
43 SB=TRN/RS*(XSSS*P+SBPSS)+SBSS
44 SYNEF=SB/SBIAV
45 XASS=YH*(1.0-SYNEF)*SBIAV*RS/
46 &(TRN*(1.0+BHT*RS*(1.0-P*YH*(1.0-F))))
47 IF(XASS.LT.0.0)GO TO 55
48 IF(XSSS.GE.(FR*XASS)) XSSS=FR*XASS
49 4 DIF=(XS0-XSSS)**2.0
50 DIF2=(SBP0-SBPSS)**2.0
51 IF(DIF.LT.0.01.AND.DIF2.LT.0.01) GO TO 6
52 SBS0=SBSS
53 SBP0=SBPSS
54 XS0=XSSS
55 XA0=XASS
56 IF (KLM.EQ.100) GO TO 55
57 15 CONTINUE
58 6 XESS=F*BHT*XASS*RS
59 XISS=XIIAV*RS/TRN
60 XVSS=XASS+XSSS+XESS+XIIS
61 SUSS=SUIAV
62 NUSS=NUIAV
63 WRITE(5,25) RS,SBSSS,SBPSS,XSSS,SB,XASS,XESS,XISS,XVSS,SYNEF,KLM

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64      25      FORMAT(1H, 5X,10F11.3,I4)
65      NSW=(FN*(XASS+XESS+XISS)+FNS*XSSS)*TRN/RS
66      NONSS=(NONIAV+TRN*((1.0-F)*FN*BHT*XASS-FNS*XSSS))/
67      1(1.0+KRT*XASS*TRN)
68      NH3MAX=NTIAV-NSW-NUSS-NONSS
69      NH3SS=KNT*(BNT+1.0/RS)/(UMT*RSA/RS-BNT-1.0/RS)*(1+VARNTL/TRN)
70      IF(NH3SS.GT.NH3MAX) NH3SS=NH3MAX
71      IF(NH3SS.LE.0.0) NH3SS=NH3MAX
72      XNSS=YN*(NH3MAX-NH3SS)*RS/(TRN*(1.0+BNT*RS))
73      NO3SS=NO3IAV+NH3MAX-NH3SS
74      IF(RS.EQ.RSA) GO TO 36
75      DENCON=((1-P*YH)/2.86)*KMPT*FACT*P*XSSS/(XSSS*P+KSPT*XASS)
76      CR=YH*RS/(1.0+BHT*RS*(1.0-P*YH*(1.0-F)))
77      DENCAP=(FBS*(1.0-P*YH)/2.86+DENCON*(1.0-RSA/RS)*CR)*SBIIV
78      NO3SS=NO3SS-DENCAP
79      IF(NO3SS.LT.0.0) NO3SS=0.0
80      ALKSS=ALKIAV-7.14*(NH3MAX-NH3SS)
81      &+3.57*(NONIAV-NONSS-NSW+DENCAP)
82      GO TO 35
83      55      WRITE(5,65)
84      65      FORMAT(1H ,10X,'NO SOLUTION AFTER 100 ITERATIONS OR XA.LT.0.0')
85      36      ALKSS=ALKIAV-7.14*(NH3MAX-NH3SS)
86      &+3.57*(NONIAV-NONSS-NSW)
87      35      CONTINUE
88      RETURN
89      END

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1 C*****
2 SUBROUTINE AROMOD(D,DT,J,I,FR,FBS,FCS,KVT,KMPT,KSPT,KMST,KSST,
3 1P,NR,YH,BHT,F,KRT,FOE,FN,FOS,FNS,NJ,DDT,UMT,YN,KNT,RT,BNT,
4 2FACT,LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR)
5 C*****
6 C THIS SUBROUTINE CALCULATES THE DYNAMIC RESPONSE OF THE
7 C FIRST REACTOR IN A SERIES UNDER AEROBIC CONDITIONS
8 C*****
9 PARAMETER NE=1
10 PARAMETER NEE=2
11 PARAMETER ND=241
12 PARAMETER NDD=242
13 INTEGER D
14 DOUBLE PRECISION DT,DDT,RTD
15 COMMON/EXTC/SBS(NE,ND),SU(NE,ND),XA(NE,ND),XE(NE,ND),XI(NE,ND),
16 1XV(NE,ND),XS(NE,ND),NU(NE,ND),NON(NE,ND),XN(NE,ND),NH3(NE,ND),
17 2NO3(NE,ND),SBP(NE,ND),ALK(NE,ND)
18 COMMON/BLOCKC/Q(NEE,NDD),V(NE),SBI(ND),SUI(ND),NIT(NE,ND),
19 1NUI(ND),XII(ND),NONI(ND),NH3I(ND),NO3I(ND),SBPR(ND),ALKI(ND),
20 2XAR(ND),XSR(ND),XER(ND),XIR(ND),XNR(ND),SYNTH(NE,ND),QAV(NEE),
21 3ANPOT(NE,ND),AMON(NE,ND),DSFCTR(NE)
22 REAL KVT,KMPT,KSPT,KMST,KSST,KRT,KNT
23 REAL NUI,NONI,NH3I,NO3I,NIT
24 REAL NU,NON,NH3,NO3
25 REAL NH3A,NH3S,NH3T,NH3B,NH3K,NITT,NNIT,NO3A,NO3S,NO3T,NO3B,NO3K
26 C*****
27 CALL DUMVCY(J,I,DUMIF,DUMS,DUMA,DUMB,DUMIR,DUMO,
28 1LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR,
29 1DT,DDT,RTD)
30 C*****
31 K=J-1
32 IF(K.EQ.0) K=1
33 SCER=P*(1.0-F)*BHT*XA(J,I)*DDT
34 SCERN=FN*SCER/P
35 CVNON=KRT*XA(J,I)*NON(J,I)*DDT
36 SCVOD=KMPT*XS(J,I)/(KSPT*XA(J,I)+XS(J,I)*P)*XA(J,I)*DDT
37 SCVO=0.00
38 SCIS=0.00
39 NNIT=0.00
40 SBST=SBS(J,I)
41 DFSBSS=(SBS(NR,I+1)-SBS(NR,I))/NJ
42 DFSBSA=(SBS(LARO,I+1)-SBS(LARO,I))/NJ
43 DFSBSB=(SBS(LBRO,I+1)-SBS(LBRO,I))/NJ
44 DFSBSK=(SBS(K,I+1)-SBS(K,I))/NJ
45 NH3T=NH3(J,I)
46 DFNH3S=(NH3(NR,I+1)-NH3(NR,I))/NJ
47 DFNH3A=(NH3(LARO,I+1)-NH3(LARO,I))/NJ
48 DFNH3B=(NH3(LBRO,I+1)-NH3(LBRO,I))/NJ
49 DFNH3K=(NH3(K,I+1)-NH3(K,I))/NJ
50 NO3T=NO3(J,I)
51 DFNO3S=(NO3(NR,I+1)-NO3(NR,I))/NJ
52 DFNO3A=(NO3(LARO,I+1)-NO3(LARO,I))/NJ
53 DFNO3B=(NO3(LBRO,I+1)-NO3(LBRO,I))/NJ
54 DFNO3K=(NO3(K,I+1)-NO3(K,I))/NJ
55 XNT=XN(J,I)
56 DFXNS=(XNR(I+1)-XNR(I))/NJ
57 DFXNA=(XN(LARO,I+1)-XN(LARO,I))/NJ
58 DFXNB=(XN(LBRO,I+1)-XN(LBRO,I))/NJ
59 DFXNK=(XN(K,I+1)-XN(K,I))/NJ
60 DO 12 IJ=1,NJ,1
61 SCVOT=SCVOD
62 SCIST=KMST*SBST/(KSST+SBST)*XA(J,I)*DDT
63 SBSS=SBS(NR,I)+(IJ-1)*DFSBSS

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64      SBSA=SBS(LARO,I)+(IJ-1)*DFSBSA
65      SBSB=SBS(LBRO,I)+(IJ-1)*DFSBSB
66      SBSK=SBS(K,I)+(IJ-1)*DFSBSK
67      SBST=SBST+DUMIF*FBS*SBI(I)-SCIST+FCS*SCER
68      1+DUMS*SBSS+DUMA*SBSA-DUMO*SBST
69      1+DUMB*SBSB+DUMIR*SBSK
70      IF(SBST.LT.0.00) SBST=0.00
71      SCVO=SCVO+SCVOT
72      SCIS=SCIS+SCIST
73      SYNTHT=SCVOT*P+SCIST
74      NITT=(UMT/YN)*NH3T/(KNT+NH3T)*XNT*DDT
75      NNIT=NNIT+NITT
76      NH3S=NH3(NR,I)+(IJ-1)*DFNH3S
77      NH3A=NH3(LARO,I)+(IJ-1)*DFNH3A
78      NH3B=NH3(LBRO,I)+(IJ-1)*DFNH3B
79      NH3K=NH3(K,I)+(IJ-1)*DFNH3K
80      NO3S=NO3(NR,I)+(IJ-1)*DFNO3S
81      NO3A=NO3(LARO,I)+(IJ-1)*DFNO3A
82      NO3B=NO3(LBRO,I)+(IJ-1)*DFNO3B
83      NO3K=NO3(K,I)+(IJ-1)*DFNO3K
84      TNH3T=NH3T+DUMIF*NH3I(I)+CVNON-NITT
85      1+DUMS*NH3S+DUMA*NH3A-DUMO*NH3T
86      1+DUMB*NH3B+DUMIR*NH3K
87      1+(1.0-FOE)*SCERN-(1.0-FOS)*FN*YH*SYNTHT
88      IF(TNH3T.GT.0.0) GO TO 43
89      NH3T=0.0
90      NO3T=NO3T+DUMIF*NO3I(I)+NITT+TNH3T
91      1+DUMS*NO3S+DUMA*NO3A-DUMO*NO3T
92      1+DUMB*NO3B+DUMIR*NO3K
93      GO TO 44
94      43 NH3T=TNH3T
95      NO3T=NO3T+DUMIF*NO3I(I)+NITT
96      1+DUMS*NO3S+DUMA*NO3A-DUMO*NO3T
97      1+DUMB*NO3B+DUMIR*NO3K
98      44 XNS=XNR(I)+(IJ-1)*DFXNS
99      XNA=XN(LARO,I)+(IJ-1)*DFXNA
100     XNB=XN(LBRO,I)+(IJ-1)*DFXNB
101     XNK=XN(K,I)+(IJ-1)*DFXNK
102     XNT=XNT+YN*NITT-BNT*XNT*DDT
103     1+DUMS*XNS+DUMA*XNA-DUMO*XNT
104     1+DUMB*XNB+DUMIR*XNK
105     12 CONTINUE
106     SYNTH(J,I)=SCVO*P+SCIS
107     NIT(J,I)=NNIT
108     ANPOT(J,I)=0.00
109     SBS(J,I+1)=SBST
110     NH3(J,I+1)=NH3T
111     NO3(J,I+1)=NO3T
112     XN(J,I+1)=XNT
113     DUMIF=DUMIF/RDT
114     DUMIR=DUMIR/RDT
115     DUMA=DUMA/RDT
116     DUMB=DUMB/RDT
117     DUMS=DUMS/RDT
118     DUMO=DUMO/RDT
119     BRAK=FR-(XS(J,I)/(XA(J,I)))
120     IF(BRAK.GT.0.00) GO TO 40
121     BRAK=0.00
122     XS(J,I)=FR*(XA(J,I))
123     40 XV(J,I)=XS(J,I)+XA(J,I)+XE(J,I)+XI(J,I)
124     SCIP=DT*KVT*BRAK*SBP(J,I)*XA(J,I)
125     SBP(J,I+1)=SBP(J,I)+DUMIF*(1.0-FBS)*SBI(I)-SCIP
126     1+DUMS*SBPR(I)+DUMA*SBP(LARO,I)-DUMO*SBP(J,I)
127     1+DUMB*SBP(LBRO,I)+DUMIR*SBP(K,I)

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128      2+(1.0-FCS)*SCER/RDT
129      IF(SBP(J,I+1).LT.0.0) SBP(J,I+1)=0.0
130      XS(J,I+1)=XS(J,I)+SCIP/P-SCVO
131      1+DUMS*XS(I)+DUMA*XS(LARO,I)-DUMO*XS(J,I)
132      1+DUMB*XS(LBRO,I)+DUMIR*XS(K,I)
133      IF(XS(J,I+1).LT.0.0) XS(J,I+1)=0.0
134      SU(J,I+1)=SU(J,I)+DUMIF*SUI(I)
135      1+DUMS*SU(NR,I)+DUMA*SU(LARO,I)-DUMO*SU(J,I)
136      1+DUMB*SU(LBRO,I)+DUMIR*SU(K,I)
137      NU(J,I+1)=NU(J,I)+DUMIF*NUI(I)
138      1+DUMS*NU(NR,I)+DUMA*NU(LARO,I)-DUMO*NU(J,I)
139      1+DUMB*NU(LBRO,I)+DUMIR*NU(K,I)
140      XA(J,I+1)=XA(J,I)+YH*SYNTH(J,I)-BHT*XA(J,I)*DT
141      1+DUMS*XAR(I)+DUMA*XA(LARO,I)-DUMO*XA(J,I)
142      1+DUMB*XA(LBRO,I)+DUMIR*XA(K,I)
143      XE(J,I+1)=XE(J,I)+F*BHT*XA(J,I)*DT
144      1+DUMS*XER(I)+DUMA*XE(LARO,I)-DUMO*XE(J,I)
145      1+DUMB*XE(LBRO,I)+DUMIR*XE(K,I)
146      XI(J,I+1)=XI(J,I)+DUMIF*XII(I)
147      1+DUMS*XIR(I)+DUMA*XI(LARO,I)-DUMO*XI(J,I)
148      1+DUMB*XI(LBRO,I)+DUMIR*XI(K,I)
149      NON(J,I+1)=NON(J,I)+DUMIF*NONI(I)-CVNON/RDT
150      1+DUMS*NON(NR,I)+DUMA*NON(LARO,I)-DUMO*NON(J,I)
151      1+DUMB*NON(LBRO,I)+DUMIR*NON(K,I)
152      2+FOE*SCERN/RDT-FOS*FN*YH*SYNTH(J,I)
153      3-FNS*(SCIP/P-SCVO)
154      AMON(J,I)=(CVNON/RDT+(1.0-FOE)*SCERN/RDT)
155      1-(1.0-FOS)*FN*YH*SYNTH(J,I)
156      ALK(J,I+1)=ALK(J,I)+DUMIF*ALKI(I)
157      1+DUMS*ALK(NR,I)+DUMA*ALK(LARO,I)-DUMO*ALK(J,I)
158      1+DUMB*ALK(LBRO,I)+DUMIR*ALK(K,I)
159      1-7.14*NIT(J,I)+3.57*AMON(J,I)
160      XV(J,I+1)=XS(J,I+1)+XA(J,I+1)+XE(J,I+1)+XI(J,I+1)
161      RETURN
162      END

```

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1 C*****
2 SUBROUTINE ANOMOD(D,DT,J,I,FR,FBS,FCS,KVT,KMPT,KSPT,KMST,KSST,
3 1P,NR,YH,BHT,F,KRT,FOE,FN,FOS,FNS,NJ,DDT,UMT,YN,KNT,RDT,BNT,
4 2FACT,LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR)
5 C*****
6 C THIS SUBROUTINE CALCULATES THE DYNAMIC RESPONSE OF A
7 C REACTOR IN A SERIES UNDER ANOXIC CONDITIONS
8 C*****
9 PARAMETER NE=1
10 PARAMETER NEE=2
11 PARAMETER ND=241
12 PARAMETER NDD=242
13 INTEGER D
14 DOUBLE PRECISION DT,DDT,RDT
15 COMMON/EXTC/SBS(NE,ND),SU(NE,ND),XA(NE,ND),XE(NE,ND),XI(NE,ND),
16 1XV(NE,ND),XS(NE,ND),NU(NE,ND),NON(NE,ND),XN(NE,ND),NH3(NE,ND),
17 2NO3(NE,ND),SBP(NE,ND),ALK(NE,ND)
18 COMMON/BLOCKC/Q(NEE,NDD),V(NE),SBI(ND),SUI(ND),NIT(NE,ND),
19 1NUI(ND),XII(ND),NONI(ND),NH3I(ND),NO3I(ND),SBPR(ND),ALKI(ND),
20 2XAR(ND),XSR(ND),XER(ND),XIR(ND),XNR(ND),SYNTH(NE,ND),QAV(NEE),
21 3ANPOT(NE,ND),AMON(NE,ND),DSFCTR(NE)
22 REAL KVT,KMPT,KSPT,KMST,KSST,KRT,KNT
23 REAL NUI,NONI,NH3I,NO3I,NIT
24 REAL NU,NON,NH3,NO3
25 REAL NH3A,NH3S,NH3T,NH3B,NH3K,NIT,T,NNIT,NO3A,NO3S,NO3T,NO3B,NO3K
26 RNCOD=2.85/(1.0-P*YH)
27 C*****
28 CALL DUMVCY(J,I,DUMIF,DUMS,DUMA,DUMB,DUMIR,DUMO,
29 1LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR,
30 1DT,DDT,RDT)
31 C*****
32 K=J-1
33 IF(K.EQ.0) K=1
34 SCER=P*(1.0-F)*BHT*XA(J,I)*DDT
35 SCERN=FN*SCER/P
36 CVNON=KRT*XA(J,I)*NON(J,I)*DDT
37 SCVOD=KMPT*XS(J,I)/(KSPT*XA(J,I)+XS(J,I)*P)*FACT*XA(J,I)*DDT
38 SCVO=0.00
39 SCIS=0.00
40 NNIT=0.00
41 ANPOTS=0.00
42 SBST=SBS(J,I)
43 DFSBSS=(SBS(NR,I+1)-SBS(NR,I))/NJ
44 DFSBSA=(SBS(LARO,I+1)-SBS(LARO,I))/NJ
45 DFSBSB=(SBS(LBRO,I+1)-SBS(LBRO,I))/NJ
46 DFSBSK=(SBS(K,I+1)-SBS(K,I))/NJ
47 NH3T=NH3(J,I)
48 DFNH3S=(NH3(NR,I+1)-NH3(NR,I))/NJ
49 DFNH3A=(NH3(LARO,I+1)-NH3(LARO,I))/NJ
50 DFNH3B=(NH3(LBRO,I+1)-NH3(LBRO,I))/NJ
51 DFNH3K=(NH3(K,I+1)-NH3(K,I))/NJ
52 NO3T=NO3(J,I)
53 DFNO3S=(NO3(NR,I+1)-NO3(NR,I))/NJ
54 DFNO3A=(NO3(LARO,I+1)-NO3(LARO,I))/NJ
55 DFNO3B=(NO3(LBRO,I+1)-NO3(LBRO,I))/NJ
56 DFNO3K=(NO3(K,I+1)-NO3(K,I))/NJ
57 XNT=XN(J,I)
58 DFXNS=(XNR(I+1)-XNR(I))/NJ
59 DFXNA=(XN(LARO,I+1)-XN(LARO,I))/NJ
60 DFXNB=(XN(LBRO,I+1)-XN(LBRO,I))/NJ
61 DFXNK=(XN(K,I+1)-XN(K,I))/NJ
62 DO 12 IJ=1,NJ,1
63 SCVOT=SCVOD

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64 SCIST=KMST*SBST/(KSST+SBST)*FACT*XA(J,I)*DDT
65 SYNTHT=SCVOT*P+SCIST
66 DENCAP=SYNTHT/RNCOD
67 DENIT=DENCAP
68 RDEN=SCIST/SYNTHT
69 ANPOTT=0.00
70 IF((NO3T-DENIT).GT.0.00) GO TO 13
71 DENIT=NO3T
72 SYNTHT=DENIT*RNCOD
73 SCIST=SYNTHT*RDEN
74 SCVOT=SYNTHT*(1.0-RDEN)/P
75 ANPOTT=DENCAP-DENIT
76 13 CONTINUE
77 SCVO=SCVO+SCVOT
78 SCIS=SCIS+SCIST
79 ANPOTS=ANPOTS+ANPOTT
80 SBSS=SBS(NR,I)+(IJ-1)*DFSBS
81 SBSA=SBS(LARO,I)+(IJ-1)*DFSBSA
82 SBSB=SBS(LBRO,I)+(IJ-1)*DFSBSB
83 SBSK=SBS(K,I)+(IJ-1)*DFSBSK
84 SBST=SBST+DUMIF*FBS*SBI(I)-SCIST+FCS*SCER
85 1+DUMS*SBSS+DUMA*SBSA-DUMO*SBST
86 1+DUMB*SBSB+DUMIR*SBSK
87 IF(SBST.LT.0.00) SBST=0.00
88 NITT=0.00
89 NNIT=NNIT+NITT
90 NH3S=NH3(NR,I)+(IJ-1)*DFNH3S
91 NH3A=NH3(LARO,I)+(IJ-1)*DFNH3A
92 NH3B=NH3(LBRO,I)+(IJ-1)*DFNH3B
93 NH3K=NH3(K,I)+(IJ-1)*DFNH3K
94 NO3S=NO3(NR,I)+(IJ-1)*DFNO3S
95 NO3A=NO3(LARO,I)+(IJ-1)*DFNO3A
96 NO3B=NO3(LBRO,I)+(IJ-1)*DFNO3B
97 NO3K=NO3(K,I)+(IJ-1)*DFNO3K
98 TNH3T=NH3T+DUMIF*NH3I(I)+CVNON-NITT
99 1+DUMS*NH3S+DUMA*NH3A-DUMO*NH3T
100 1+DUMB*NH3B+DUMIR*NH3K
101 1+(1.0-FOE)*SCERN-(1.0-FOS)*FN*YH*SYNTHT
102 IF(TNH3T.GT.0.0) GO TO 43
103 NH3T=0.0
104 NO3T=NO3T+DUMIF*NO3I(I)+NITT-DENIT+TNH3T
105 1+DUMS*NO3S+DUMA*NO3A-DUMO*NO3T
106 1+DUMB*NO3B+DUMIR*NO3K
107 IF(NO3T.LT.0.00) NO3T=0.00
108 GO TO 44
109 43 NH3T=TNH3T
110 NO3T=NO3T+DUMIF*NO3I(I)+NITT-DENIT
111 1+DUMS*NO3S+DUMA*NO3A-DUMO*NO3T
112 1+DUMB*NO3B+DUMIR*NO3K
113 IF(NO3T.LT.0.00) NO3T=0.00
114 44 XNS=XNR(I)+(IJ-1)*DFXNS
115 XNA=XN(LARO,I)+(IJ-1)*DFXNA
116 XNB=XN(LBRO,I)+(IJ-1)*DFXNB
117 XNK=XN(K,I)+(IJ-1)*DFXNK
118 XNT=XNT+YN*NITT-BNT*XNT*DDT
119 1+DUMS*XNS+DUMA*XNA-DUMO*XNT
120 1+DUMB*XNB+DUMIR*XNK
121 12 CONTINUE
122 SYNTH(J,I)=SCVO*P+SCIS
123 ANPOT(J,I)=ANPOTS
124 NIT(J,I)=NNIT
125 SBS(J,I+1)=SBST
126 NH3(J,I+1)=NH3T
127 NO3(J,I+1)=NO3T

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128      XN(J,I+1)=XNT
129      DUMIF=DUMIF/RDT
130      DUMIR=DUMIR/RDT
131      DUMA=DUMA/RDT
132      DUMB=DUMB/RDT
133      DUMS=DUMS/RDT
134      DUMO=DUMO/RDT
135      BRAK=FR-(XS(J,I)/(FACT*XA(J,I)))
136      IF(BRAK.GT.0.00) GO TO 40
137      BRAK=0.00
138      XS(J,I)=FR*(FACT*XA(J,I))
139 40     XV(J,I)=XS(J,I)+XA(J,I)+XE(J,I)+XI(J,I)
140      SCIP=DT*KVT*BRAK*SBP(J,I)*FACT*XA(J,I)
141      SBP(J,I+1)=SBP(J,I)+DUMIF*(1.0-FBS)*SBI(I)-SCIP
142      1+DUMS*SBPR(I)+DUMA*SBP(LARO,I)-DUMO*SBP(J,I)
143      1+DUMB*SBP(LBRO,I)+DUMIR*SBP(K,I)
144      2+(1.0-FCS)*SCER/RDT
145      IF(SBP(J,I+1).LT.0.0) SBP(J,I+1)=0.0
146      XS(J,I+1)=XS(J,I)+SCIP/P-SCVO
147      1+DUMS*XSR(I)+DUMA*XS(LARO,I)-DUMO*XS(J,I)
148      1+DUMB*XS(LBRO,I)+DUMIR*XS(K,I)
149      IF(XS(J,I+1).LT.0.0) XS(J,I+1)=0.0
150      SU(J,I+1)=SU(J,I)+DUMIF*SUI(I)
151      1+DUMS*SU(NR,I)+DUMA*SU(LARO,I)-DUMO*SU(J,I)
152      1+DUMB*SU(LBRO,I)+DUMIR*SU(K,I)
153      NU(J,I+1)=NU(J,I)+DUMIF*NUI(I)
154      1+DUMS*NU(NR,I)+DUMA*NU(LARO,I)-DUMO*NU(J,I)
155      1+DUMB*NU(LBRO,I)+DUMIR*NU(K,I)
156      XA(J,I+1)=XA(J,I)+YH*SYNTH(J,I)-BHT*XA(J,I)*DT
157      1+DUMS*XAR(I)+DUMA*XA(LARO,I)-DUMO*XA(J,I)
158      1+DUMB*XA(LBRO,I)+DUMIR*XA(K,I)
159      XE(J,I+1)=XE(J,I)+F*BHT*XA(J,I)*DT
160      1+DUMS*XER(I)+DUMA*XE(LARO,I)-DUMO*XE(J,I)
161      1+DUMB*XE(LBRO,I)+DUMIR*XE(K,I)
162      XI(J,I+1)=XI(J,I)+DUMIF*XII(I)
163      1+DUMS*XIR(I)+DUMA*XI(LARO,I)-DUMO*XI(J,I)
164      1+DUMB*XI(LBRO,I)+DUMIR*XI(K,I)
165      NON(J,I+1)=NON(J,I)+DUMIF*NONI(I)-CVNON/RDT
166      1+DUMS*NON(NR,I)+DUMA*NON(LARO,I)-DUMO*NON(J,I)
167      1+DUMB*NON(LBRO,I)+DUMIR*NON(K,I)
168      2+FOE*SCERN/RDT-FOS*FN*YH*SYNTH(J,I)
169      3-FNS*(SCIP/P-SCVO)
170      AMON(J,I)=(CVNON/RDT+(1.0-FOE)*SCERN/RDT)
171      1-(1.0-FOS)*FN*YH*SYNTH(J,I)
172      ALK(J,I+1)=ALK(J,I)+DUMIF*ALKI(I)
173      1+DUMS*ALK(NR,I)+DUMA*ALK(LARO,I)-DUMO*ALK(J,I)
174      1+DUMB*ALK(LBRO,I)+DUMIR*ALK(K,I)
175      1+3.57*(SYNTH(J,I)/RNCOD+AMON(J,I))
176      XV(J,I+1)=XS(J,I+1)+XA(J,I+1)+XE(J,I+1)+XI(J,I+1)
177      RETURN
178      END

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1 C*****
2 SUBROUTINE DSTRCY(CRIT,VP,XVSS,NR,LINI,LSRI,LARI,LARO,LBRI,LBRO,
3 1QSR,QAR,QBR,SR,AR,BR,DT,DDT,RDT,NLL)
4 C*****
5 C THIS SUBROUTINE DISTRIBUTES THE SLUDGE MASS IN THE
6 C PROCESS TO EACH REACTOR IN ACCORDANCE WITH THE PROCESS
7 C CONFIGURATION INPUT DATA AND CALCULATES THE SLUDGE
8 C CONCENTRATION IN EACH REACTOR AS A FRACTION OF THE
9 C AVERAGE PROCESS SLUDGE CONCENTRATION
10 C*****
11 PARAMETER NE=1
12 PARAMETER NEE=2
13 PARAMETER ND=241
14 PARAMETER NDD=242
15 DOUBLE PRECISION DT,DDT,RDT
16 COMMON/EXTC/SBS(NE,ND),SU(NE,ND),XA(NE,ND),XE(NE,ND),XI(NE,ND),
17 1XV(NE,ND),XS(NE,ND),NU(NE,ND),NON(NE,ND),XN(NE,ND),NH3(NE,ND),
18 2NO3(NE,ND),SBP(NE,ND),ALK(NE,ND)
19 COMMON/BLOCKC/Q(NEE,NDD),V(NE),SBI(ND),SUI(ND),NIT(NE,ND),
20 1NUI(ND),XII(ND),NONI(ND),NH3I(ND),NO3I(ND),SBPR(ND),ALKI(ND),
21 2XAR(ND),XSR(ND),XER(ND),XIR(ND),XNR(ND),SYNTH(NE,ND),QAV(NEE),
22 3ANPOT(NE,ND),AMON(NE,ND),DSFCTR(NE)
23 DIMENSION XBGN(NE),STXBGN(NE)
24 CRITXB=CRIT*XVSS*0.001
25 I=NLL
26 DO 1 J=1,NR,1
27 XBGN(J)=XVSS
28 Q(J,I)=QAV(1)
29 1 CONTINUE
30 MM=0
31 7 DO 2 J=1,NR,1
32 STXBGN(J)=XBGN(J)
33 2 CONTINUE
34 DO 3 J=1,NR,1
35 K=J-1
36 IF(K.EQ.0) K=1
37 C*****
38 CALL DUMVCY(J,I,DUMIF,DUMS,DUMA,DUMB,DUMIR,DUMO,
39 1LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR,
40 1DT,DDT,RDT)
41 C*****
42 XBGN(J)=XBGN(J)+DUMA*XBGN(LARO)+DUMB*XBGN(LBRO)
43 1+DUMS*(1+SR)/SR*XBGN(NR)+DUMIR*XBGN(K)-DUMO*XBGN(J)
44 3 CONTINUE
45 DO 4 J=1,NR,1
46 IF(ABS(XBGN(J)-STXBGN(J)).GE.CRITXB) GO TO 5
47 4 CONTINUE
48 GO TO 6
49 5 MM=MM+1
50 IF(MM.GE.5000) GO TO 6
51 GO TO 7
52 6 CONTINUE
53 TXBGNO=0.0
54 DO 9 J=1,NR,1
55 TXBGNO=TXBGNO+V(J)*XBGN(J)
56 DSFCTR(J)=XBGN(J)/XVSS
57 9 CONTINUE
58 XBAL=TXBGNO/(XVSS*VP)*100
59 WRITE(5,104) MM,XVSS,XBAL
60 104 FORMAT(1H0,5X,'NO. OF STEPS TO REACH STEADY STATE= ',I5,/,
61 11H,5X,'AVERAGE PROCESS SLUDGE CONCENTRATION= ',F6.1,/,
62 11H,5X,'PERCENTAGE SLUDGE RECOVERY = ',F7.3)
63 WRITE(5,105) (DSFCTR(J),J=1,NR,1)

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64      105 FORMAT((1H ,5X,F10.4))  
65      RETURN  
66      END
```

```

1 C*****
2 SUBROUTINE DUMVCY(J,I,DUMIF,DUMS,DUMA,DUMB,DUMIR,DUMO,
3 1LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR,
4 1DT,DDT,RDT)
5 C*****
6 C THIS SUBROUTINE CALCULATES THE DUMMY VALUES
7 C FROM THE PROCESS CONFIGURATION INPUT DATA
8 C*****
9 PARAMETER NE=1
10 PARAMETER NEE=2
11 PARAMETER ND=241
12 PARAMETER NDD=242
13 DOUBLE PRECISION DT,DDT,RDT
14 COMMON/EXTC/SBS(NE,ND),SU(NE,ND),XA(NE,ND),XE(NE,ND),XI(NE,ND),
15 1XV(NE,ND),XS(NE,ND),NU(NE,ND),NON(NE,ND),XN(NE,ND),NH3(NE,ND),
16 2NO3(NE,ND),SBP(NE,ND),ALK(NE,ND)
17 COMMON/BLOCKC/Q(NEE,NDD),V(NE),SBI(ND),SUI(ND),NIT(NE,ND),
18 1NUI(ND),XII(ND),NONI(ND),NH3I(ND),NO3I(ND),SBPR(ND),ALKI(ND),
19 2XAR(ND),XSR(ND),XER(ND),XIR(ND),XNR(ND),SYNTH(NE,ND),QAV(NEE),
20 3ANPOT(NE,ND),AMON(NE,ND),DSFCTR(NE)
21 DUMIF=0.0
22 IF(J.EQ.LINI) DUMIF=DDT*Q(J,I)/V(J)
23 DUMS=0.0
24 IF(J.EQ.LSRI) DUMS=DDT*QSR/V(J)
25 DUMA=0.0
26 IF(J.EQ.LARI.AND.J.LT.LARO) DUMA=DDT*QAR/V(J)
27 DUMB=0.0
28 IF(J.EQ.LBRI.AND.J.LT.LBRO) DUMB=DDT*QBR/V(J)
29 IF(J.EQ.1) GO TO 11
30 IF(J.LE.LINI.AND.J.GT.LSRI) DUMIR=DDT*QSR/V(J)
31 IF(J.LE.LINI.AND.J.LE.LSRI.AND.J.GT.LBRI.AND.J.LE.LBRO)
32 1DUMIR=DDT*QBR/V(J)
33 IF(J.LE.LINI.AND.J.LE.LSRI.AND.J.GT.LARI.AND.J.LE.LARO)
34 1DUMIR=DDT*QAR/V(J)
35 IF(J.LE.LINI.AND.J.LE.LSRI.AND.J.GT.LARI.AND.J.LE.LARO.
36 1AND.J.GT.LBRI.AND.J.LE.LBRO) DUMIR=DDT*(QAR+QBR)/V(J)
37 IF(J.LE.LINI.AND.J.GT.LSRI.AND.J.GT.LARI.AND.J.LE.LARO)
38 1DUMIR=DDT*(QSR+QAR)/V(J)
39 IF(J.LE.LINI.AND.J.GT.LSRI.AND.J.GT.LBRI.AND.J.LE.LBRO)
40 1DUMIR=DDT*(QSR+QBR)/V(J)
41 IF(J.LE.LINI.AND.J.GT.LSRI.AND.J.GT.LARI.AND.J.LE.LARO.
42 1AND.J.GT.LBRI.AND.J.LE.LBRO) DUMIR=DDT*(QSR+QAR+QBR)/V(J)
43 IF(J.GT.LINI.AND.J.GT.LSRI) DUMIR=DDT*(Q(J,I)+QSR)/V(J)
44 IF(J.GT.LINI.AND.J.LE.LSRI.AND.J.GT.LBRI.AND.J.LE.LBRO)
45 1DUMIR=DDT*(Q(J,I)+QBR)/V(J)
46 IF(J.GT.LINI.AND.J.LE.LSRI.AND.J.GT.LARI.AND.J.LE.LARO)
47 1DUMIR=DDT*(Q(J,I)+QAR)/V(J)
48 IF(J.GT.LINI.AND.J.GT.LSRI.AND.J.GT.LARI.AND.J.LE.LARO)
49 1DUMIR=DDT*(Q(J,I)+QAR+QSR)/V(J)
50 IF(J.GT.LINI.AND.J.GT.LSRI.AND.J.GT.LBRI.AND.J.LE.LBRO)
51 1DUMIR=DDT*(Q(J,I)+QBR+QSR)/V(J)
52 IF(J.GT.LINI.AND.J.GT.LSRI.AND.J.GT.LARI.AND.J.LE.LARO.
53 1AND.J.GT.LBRI.AND.J.LE.LBRO) DUMIR=DDT*(Q(J,I)+QSR+QAR+QBR)/V(J)
54 11 CONTINUE
55 IF(J.EQ.1) DUMIR=0.0
56 DUMO=DUMIF+DUMIR+DUMS+DUMA+DUMB
57 RETURN
58 END

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PROGRAM A.2

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1  C*****
2  C
3  C          UNSTEADY STATE PROGRAM AS AT 5-5-1981
4  C
5  C*****
6  C*****
7  C
8  C          'THIS PROGRAMME IS THE JOINT PROPERTY OF THE
9  C          UNIVERSITY OF CAPE TOWN AND THE WATER RESEARCH
10 C          COMMISSION OF SOUTH AFRICA. BEFORE USER MAY
11 C          LEGALLY UTILIZE IT, HE MUST OBTAIN THE NECESSARY
12 C          AUTHORIZATION, I.E.:
13 C
14 C          (A) IN THE CASE OF STUDENTS AT THE UNIVERSITY OF
15 C          CAPE TOWN WHO WILL USE THE PROGRAMME FOR
16 C          EDUCATIONAL PURPOSES ONLY, THIS AUTHORIZATION
17 C          CAN BE OBTAINED FROM PROF. G. VAN R. MARAIS
18 C          OF THE UNIVERSITY OF CAPE TOWN, AND
19 C          (B) IN ALL OTHER CASES, WRITTEN AUTHORIZATION
20 C          MUST BE OBTAINED FROM BOTH THE UNIVERSITY
21 C          OF CAPE TOWN (PRIVATE BAG, RONDEBOSCH, 7700,
22 C          CAPE, SOUTH AFRICA) AND THE WATER RESEARCH
23 C          COMMISSION (P.O. BOX 824, PRETORIA, 0001,
24 C          SOUTH AFRICA)''
25 C
26 C*****
27 C*****
28 C
29 C
30 C          THE FOLLOWING EXPLANATION REFERS TO :
31 C
32 C          ' VAN HAANDEL,C.A. AND MARAIS,G.V.R. (1981) ''KINETICS OF
33 C          NITRIFICATION AND DENITRIFICATION IN SINGLE SLUDGE ACTIVATED
34 C          SLUDGE PROCESS'', RES. REPT. NO.39, DEPT. OF CIVIL ENG.,
35 C          UNIVERSITY OF CAPE TOWN
36 C
37 C          THIS PROGRAM IS USED TO SIMULATE THE RESPONSE OF SERIES
38 C          REACTOR ACTIVATED SLUDGE SYSTEMS UNDER UNSTEADY STATE
39 C          CONDITIONS.THE REACTORS ARE ASSUMED TO BE COMPLETELY
40 C          MIXED . THE VALUES OF THE VARIABLES ARE CALCULATED AS
41 C          A FUNCTION OF TIME USING THE SUPPLIED STARTING VALUES
42 C          AND THE KINETIC EXPRESSIONS DEVELOPED BY DOLD, EKAMA
43 C          AND MARAIS FOR AN AEROBIC ENVIRONMENT (I.E. THE BI-
44 C          SUBSTRATE /DEATH REGENERATION/ACTIVE SITE THEORY) OR
45 C          THOSE LISTED IN TABLE (4.1 CHAPTER 4) FOR AN ANOXIC
46 C          ENVIRONMENT. THE CALCULATED VALUES OF ALL THE VARIABLES
47 C          IN EACH REACTOR CAN BE PRINTED AS A FUNCTION OF TIME
48 C          AND/OR WRITTEN TO A PLOT FILE FOR SUBSEQUENT PLOTTING
49 C          TOGETHER WITH EXPERIMENTAL DATA.THE PLOTTING PROGRAM IS
50 C          LISTED IN APPENDIC A.6
51 C          THE RESPONSE IS CALCULATED OVER THE PERIOD THAT INPUT
52 C          FLOW AND LOAD DATA IS SUPPLIED
53 C          THE REQUIRED PERIOD OF PLOTTED RESPONSE MUST BE SPECIFIED
54 C          THE MAIN PROGRAM HAS FOUR SUBROUTINES:
55 C          1)UDUMV CALCULATES THE HYDRAULIC EFFCTS IN THE REACTORS
56 C          2)UAERO CALCULATES THE RESPONSE OF A REACTOR IN AN AEROBIC
57 C          ENVIRONMENT
58 C          3)UANOX CALCULATES THE RESPONSE OF A REACTOR IN AN ANOXIC
59 C          ENVIRONMENT
60 C          4)USETL CALCUALTES THE RESPONSE OF THE SETTLER
61 C
62 C          THE PROGRAM AS LISTED BELOW IS SUBJECTED TO THE FOLLOWING
63 C          LIMITATIONS

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64      C      1)THE LENGTHS OF THE ANOXIC AND THE AEROBIC PERIODS IN
65      C      A REACTOR ARE FOR ONE CYCLE CAN EACH ASSUME ONE VALUE ONLY
66      C      2)THE INFLUENT FOLW CAN ONLY BE INTRODUCED INTO ONE
67      C      REACTOR
68      C      3) THE RECYCLE FLOW FROM THE SETTLER CAN ONLY BE CONVEYED
69      C      TO ONE REACTOR (WHICH IS NOT NECESSARILY THE FIRST ONE)
70      C      4) IT IS ONLY POSSIBLE TO HAVE TWO INTERNAL RECYCLES FLOWS
71      C      (A AND B) EACH OF WHICH CONVEYS MIXED LIQUOR FROM ONE
72      C      REACTOR TO ANOTHER THE LATTER BEING NEARER TO THE REACTOR
73      C      RECIEVING THE INFLUENT
74      C      5) ALL RECYCLE FLOWS ARE CONSTANT WITH TIME.
75      C      6) OPTIONS FOR THE INFLUENT FLOW PATTERN ARE:
76      C      A)SINOIDAL
77      C      B)SQUARE WAVE
78      C      C)GENERAL PATTERN WITH LINEAR INTERPOLATION
79      C      7)THE CONCENTRATION PATTERNS OF INFLUENT COD,TKN,NITRATE
80      C      AND ALKALINITY HAVE THE SAME LIMITATIONS AS MENTIONED
81      C      FOR THE INFLUENT FLOW UNDER 6
82      C      8)IT IS ASSUMED THAT DURING A CYCLE THE CONSTANTS CHARA
83      C      CHARACTERIZING THE SEWAGE (FUS,FUP,FBS,FUN,FAN,SN) DO
84      C      NOT CHANGE
85      C      9)IT IS ASSUMED THAT THE D.O. CONCENTRATION IN THE AEROBIC
86      C      REACTORS IS SUFFICIENT SO THAT IT DOES NOT LIMIT THE
87      C      RATES OF UTILIZATION OF ORGANIC SUBSTRATE OR OF NITRI-
88      C      FICATION
89      C      10)IT IS ASSUMED THAT ANY D.O. RECYCLED TO YHE ANOXIC
90      C      REACTORS DOES NOT AFFECT THE RATE OF DENITRIFICATION
91      C      11)THE PH AND TEMPERATURE ARE ASSUMED TO BE CONSTANT
92      C      OVER ONE CYCLE AND THE SAME IN ALL REACTORS
93      C      12)IS IS ASSUMED THAT THE PROCESS IS A SINGLE SLUDGE
94      C      PROCESS I.E. THERE IS ONLY ONE SETTLER
95      C      13) IT IS ASSUMED THAT SLUDGE WASTAGE TAKES PLACE FROM
96      C      ONE OR MORE REACTORS.THE FLOW OF WASTED SLUDGE IS CON-
97      C      STANT DURING THE PERIOD OF WASTAGE AND PROPORTIONAL
98      C      TO THE REACTOR VOLUME.
99      C      14) THE SETTLER IS TAKEN AS AN IDEAL LIQUID SOLID
100     C      SEPARATOR, I.E. THE RETENTION TIME IN THE SETTLER
101     C      IS ZERO AND NO REACTION TAKES PLACE IN THE SETTLER.
102     C
103     C      ALL THE LIMITATIONS ABOVE ,EXCEPT THE LAST ONE, ARE
104     C      IMPOSED TO REDUCE THE REQUIRED CORE STORAGE. IF
105     C      REQUIRED THR PROGRAM CAN BE REWRITTEN IN SUCH
106     C      FASHION THAT NONE OF THE LIMITATIONS 1 TO 13 APPLY.
107     C      THE FOURTEENTH LIMITATION CAN BE OMITTED ONLY IF SUFF
108     C      SUFFICIENT DATA IS AVAILABLE TO DESCRIBE THE
109     C      SETTLING BEHAVOUR UNDER CYCLIC FLOW AND LOAD
110     C      CONDITIONS.
111     C
112     C      NOTE THAT THIS PROGRAM CAN ONLY BE USED TO CHECK A
113     C      DESIGN AND HENCE REQUIRES A DESIGN CONFIGURATION
114     C      WITH SPECIFIED REACTOR VOLUMES.
115     C
116     C      THE PROGRAM HAS BEEN TESTED FOR DIFFERENT VARIANTS
117     C      OF THE ACTIVATED SLUDGE PROCESS, I.E.
118     C      1)SERIES SUSPENSION MIXED AERATED LAGOONS
119     C      2)CONTACT STABILIZATION PROCESS
120     C      3)OXIDATION DITCH
121     C      4)SERIES COMPLETELY NIXED REACTOR SYSTEM(1 TO 6 REACTORS)
122     C      WITH AND WITHOUT ANOXIC REACTORS.
123     C
124     C*****
125     C*****
126     C
127     C      INPUT OF DATA

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128 C -----
129 C
130 C FIRST CARD:
131 C -----
132 C
133 C TYPE = LOGICAL NUMBER DESCRIBING INFLUENT LOAD PATTERN
134 C IF = 1 SINE WAVE LOAD PATTERN
135 C IF = 2 SQUARE WAVE LOAD PATTERN
136 C IF = 3 GENERAL LOADING PATTERN
137 C RESULT = LOGICAL NUMBER TO DIRECT OUTPUT
138 C IF = 1 PRINT RESULTS
139 C IF = 2 PLOT RESULTS
140 C IF = 0 PLOT AND PRINT RESULTS
141 C KOM = TIME AT WHICH TIME BASE COUNTER COMMENCES
142 C NDH = LENGTH OF PERIOD OF PLOTTED SIMULATION
143 C M = LENGTH OF PERIOD OF CALCULATED SIMULATION
144 C DATE,XMONTH AND YEAR REFER TO THE DATE THE EXPERIMENT
145 C TO BE SIMULATED WAS CARRIED OUT.
146 C
147 C
148 C SECOND CARD
149 C -----
150 C (INPUT FLOW AND LOAD)
151 C QO = AVERAGE INFLUENT FLOW FOR SINE WAVE PATTERN
152 C STIO = INFLUENT COD CONCENTRATION DURING PEAK FLOW PERIOD FOR
153 C SQUARE WAVE PATTERN OR AVERAGE FOR SINE WAVE PATTERN
154 C NTIO = INFLUENT TKN CONCENTRATION DURING PEAK FLOW PERIOD FOR
155 C SQUARE WAVE PATTERN OR AVERAGE FOR SINE WAVE PATTERN
156 C NO3IO = INFLUENT NO3 CONCENTRATION DURING PEAK FLOW PERIOD FOR
157 C SQUARE WAVE PATTERN OR AVERAGE FOR SINE WAVE PATTERN
158 C ALKIO = INFLUENT ALK CONCENTRATION DURING PEAK FLOW PERIOD FOR
159 C SQUARE WAVE PATTERN OR AVERAGE FOR SINE WAVE PATTERN
160 C VARSTI = AMPLITUDE OF INPUT COD CONC. WAVE (SINE WAVE)
161 C VARNTI = AMPLITUDE OF INPUT TKN CONC. WAVE (SINE WAVE)
162 C VNO3I = AMPLITUDE OF INPUT NO3 CONC. WAVE (SINE WAVE)
163 C VARQ = AMPLITUDE OF INFLUENT FLOW WAVE (SINE WAVE)
164 C STIOO = INFLUENT COD CONCENTRATION DURING BASE FLOW PERIOD
165 C NTIOO = INFLUENT TKN CONCENTRATION DURING BASE FLOW PERIOD
166 C NO3IOO = INFLUENT NO3 CONCENTRATION DURING BASE FLOW PERIOD
167 C ALKIOO = INFLUENT ALK CONCENTRATION DURING BASE FLOW PERIOD
168 C LFP = LENGTH OF FEED PERIOD (SQUARE WAVE)
169 C PQ = PEAK FLOW RATE IN SQUARE WAVE PATTERN
170 C BQ = BASE FLOW RATE IN SQUARE WAVE PATTERN
171 C VF = VOLUME OF FEED PER DAY
172 C START = BEGIN FEEDING PERIOD (SQ.WAVE)
173 C
174 C THIRD CARD
175 C -----
176 C (KINETIC CONSTANTS AND MASS PARAMETERS)
177 C
178 C AN EXTRA T OR 20 INDICATES VALUES AT T OR 20 DEG. C
179 C KV = MAX. RATE OF COD TRANSFER FROM SEWAGE INTO STORAGE
180 C KR = RATE OF CONVERSION OF ORGANIC N TO SALINE NH3
181 C KMP = MAX. RATE OF ORGANISM SYNTHESIS FROM STORED COD
182 C KMS = MAX. RATE OF ORGANISM SYNTHESIS FROM SOLUBLE COD
183 C KSP = HALF SATURATION COEF. FOR STORED COD UTILIZATION
184 C KSS = HALF SATURATION COEF. FOR SOLUBLE COD UTILIZATION
185 C UM = MAXIMUM GROWTH RATE OF NITRIFYING ORGANISMS
186 C KN = HALF SATURATION COEF. FOR NH3 UTILIZATION
187 C BH = DEATH RATE
188 C BN = DEATH RATE FOR NITRIFICATION
189 C YH = YIELD OF ORGANISMS FROM COD UTILIZED
190 C YN = ACTIVE MASS YIELD OF NITRIFYING ORGANISMS FROM AMMONIA
191 C F = UNBIODEG. FRACTION OF ORGANISM

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256 C      SR = UNDERFLOW RECYCLE  RATIO
257 C      AR = A-RECYCLE RATIO
258 C      BR = B-RECYCLE RATIO
259 C
260 C      EIGHTH CARD
261 C      -----
262 C      (ANOXIC SLUDGE MASS FRACTION)
263 C      INIT = INITIAL AEROBIC PERIOD
264 C      ANOX = DURATION OF AN ANOXIC PERIOD]
265 C      AROB = DUARTION OF AN AEROBIC PERIOD
266 C      NOTE
267 C      -----
268 C      NITROGEN CONCENTRATIONS MEASURED IN MG-N/L WITH TKN TEST
269 C      COD CONCENTRATIONS MEASURED IN MG/L OF COD
270 C      VSS CONCENTRATIONS MEASURED IN MG/L OF VSS
271 C      OXYGEN CONSUMPTIONS IN MG/L/HOUR
272 C      UNIT OF MASS IS THE MG
273 C      UNIT OF VOLUME IS THE L
274 C      UNIT OF TIME IS THE DAY
275 C      SYMBOL S REFERS TO COD
276 C      SYMBOL X REFERS TO VSS
277 C      SYMBOL N REFERS TO NITROGEN
278 C      SYMBOL C REFERS TO OXYGEN
279 C
280 C
281 C
282 C      DIMENSION STATEMENTS
283 C      -----
284 C
285 C      PARAMETER NE=6
286 C      PARAMETER NEE=7
287 C      PARAMETER ND=241
288 C      PARAMETER NDD=242
289 C      COMMON/EXTC/SBS(NE,ND),SU(NE,ND),XA(NE,ND),XE(NE,ND),XI(NE,ND),
290 C      1XV(NE,ND),XS(NE,ND),NU(NE,ND),NON(NE,ND),XN(NE,ND),NH3(NE,ND),
291 C      2NO3(NE,ND),SBP(NE,ND),ALK(NE,ND)
292 C      COMMON/3BLOCKC/Q(NEE,NDD),V(NE),SBI(ND),SUI(ND),NIT(NE,ND),
293 C      1NUI(ND),XII(ND),NONI(ND),NH3I(ND),NO3I(ND),SBPR(ND),ALKI(ND),
294 C      2XAR(ND),XSR(ND),XER(ND),XIR(ND),XNR(ND),SYNTH(NE,ND),QAV(NEE),
295 C      3ANPOT(NE,ND),AMON(NE,ND)
296 C      CHARACTER IMFT*19/'((20X,I4,(E13.5)))'/,CH*1
297 C      REAL KVT,KMPT,KSPT,KMST,KSST,KRT,KNT,NTITI,NTITO
298 C      REAL NUI,NONI,NH3I,NO3I,NIT
299 C      REAL NU,NON,NH3,NO3
300 C      INTEGER D,DD,HK,TYPE,RESULT,TN
301 C      DOUBLE PRECISION DT,DDT,RDT
302 C      DIMENSION STI(ND),STQ(ND)
303 C      REAL NTI(ND),NTQ(ND),NO3Q(ND),NXII(ND)
304 C      REAL NTIAV,NUIAV,NONIAV,NO3IAV,NH3IAV,NXIIAV
305 C      REAL NTQAV,NO3QAV
306 C      REAL NTIMAX,NO3IAX,NTQMAX,NO3QAX
307 C      REAL NTI0,NTI00,NO3I0,NO3I00,LFP
308 C      DIMENSION RA(NE),RN(NE),LRSD(NE),NTDN(NE),INTDN(NE),ALKQ(ND)
309 C      REAL KV20,KMP20,KSP20,KMS20,KSS20,KNMAX,KN20,KR20
310 C      DIMENSION KO(ND),WQ(NE,ND),INIT(NE),ANOX(NE),AROB(NE),FANOX(NE),
311 C      1OC(NE,ND),ON(NE,ND),OT(NE,ND),ST(NE,ND),RXS(NE,ND)
312 C      REAL NT(NE,ND)
313 C      DIMENSION SBSAV(NE),ODAV(NE),OCAV(NE),RNO3(NE),ALKAV(NE),AMAV(NE),
314 C      1ONAV(NE),OTAV(NE),STAV(NE),SUAV(NE),SBPAV(NE),
315 C      2XAAV(NE),XEAV(NE),XSAV(NE),XIAV(NE),XVAV(NE),
316 C      3XNAV(NE),RXSAV(NE),NAME(NE),LNAME(3),APTAV(NE),APTNR(NE)
317 C      REAL NONAV(NE),NTAV(NE),NO3AV(NE),NH3AV(NE),NUAV(NE),NSWAV(NE)
318 C      REAL NONSS,NUSS,NO3SS,NH3SS
319 C

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320 C
321 C      INPUT OF DATA
322 C      -----
323 100 FORMAT ( )
324 101 FORMAT(2I1,I4,I3,I4,A2,A3,A4)
325     READ 101,TYPE,RESULT,KOM,NDH,M,DATE,XMONTH,YEAR
326     IF(TYPE.EQ.1) GO TO 111
327     IF(TYPE.EQ.2) GO TO 112
328     IF(TYPE.EQ.3) GO TO 113
329 111 READ 100,PI
330     WRITE (5,100)PI
331     READ 100,Q0,STI0,NTI0,NO3I0,ALKI0,VARQ,VARSTI,VARNTI,VNO3I,VALKI
332     GO TO 113
333 112 READ 100,VF,STI0,NTI0,NO3I0,ALKI0,BQ,STI00,NTI00,NO3I00,ALKI00,LFP
334     &,START
335 113 READ(8,100) KV20,KMP20,KSP20,KMS20,KSS20,UMAX,KNMAX,BH20,BN20,
336     1KR20,FOS,FBS,FR,YH,YN,F,P,FN,FOE,FACT,FCS
337     READ 100,RS,NR,D,NJ,FUS,FUP,UN,SN
338     READ 100,THEA,THEM,THES,THEE,PHIN,THEN,THEZ,TEMP,PH
339     READ(8,100) (V(J),LRSD(J),J=1,NR,1)
340     READ(8,100) ICSD,ISSD,NRSD,LINI,LSRI,LARI,LARO,LBRI,LBRO,SR,AR,BR
341     READ(8,100) (INIT(J),ANOX(J),AROB(J),J=1,NR)
342     KH=D/24
343     NL=D+1
344     NLL=D+2
345     DD=NJ*D
346     NRR=NR+1
347 C
348 C
349 C      CALCULATION CHARACTERISTICS
350 C      -----
351 C      THIS PROGRAMME REQUIRES THE FOLLOWING CORE STORAGE CAPACITY:-
352 C      FOR ND=241 AND NF=4*(ND-1)+1 AND NEE=NE+1
353 C      STORAGE=10*NE+23 KILOWORDS
354 C      KO = TIME BASE VARIABLE
355 C      D = NUMBER OF INTERGRATION STEPS PER DAY
356 C      DT = INTERGRATION STEP LENGTH
357 C      DD = NUMBER OF INTERGRATION STEPS PER DAY FOR NITRIFICATION
358 C      DDT = INTERGRATION STEP LENGTH FOR NITRIFICATION
359 C      RDT = RATIO OF INTERGRATION STEP LENGTHS
360 C      KH = NUMBER OF TIME STEPS PER HOUR
361 C      HK = NO. OF INTERVALS PER DAY OF SLUDGE WASTAGE
362 C
363 C      SEWAGE CHARACTERISTICS
364 C      -----
365 C      Q = INFLUENT FLOW
366 C      STI = TOTAL INFLUENT COD
367 C      NTI = TOTAL INFLUENT TKN
368 C      STQ = TOTAL INFLUENT COD LOAD
369 C      NTQ = TOTAL INFLUENT TKN LOAD
370 C      NO3Q = TOTAL INFLUENT NO3 LOAD
371 C      NONI = INFLUENT ORGANIC NITROGEN CONCENTRATION
372 C      NH3I = INFLUENT AMMONIA CONCENTRATION
373 C      NO3I = INFLUENT NITRATE CONCENTRATION
374 C      NUI = UNBIODEGRADABLE FRACTION OF SEWAGE N
375 C      NXII = UNSABLE N IN INERT MATERIAL IN INFLUENT
376 C      XII = UNBIODEG. SOLID FRACTION OF SEWAGE COD AS VSS
377 C      SBI = BIODEG. FRACTION OF SEWAGE COD
378 C      SUI = UNBIODEG. SOLUBLE FRACTION OF SEWAGE COD
379 C      NM,NN = LIMITS OF PEAK FLOW PERIOD
380 C      QMAX = MAXIMUM FLOW VALUE OF THE DAY
381 C      STIMAX = MAXIMUM COD CONCENTRATION VALUE OF THE DAY
382 C      NTIMAX = MAXIMUM TKN CONCENTRATION VALUE OF THE DAY
383 C      NO3IAX = MAXIMUM NO3 CONCENTRATION VALUE OF THE DAY

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384 C      STQMAX = MAXIMUM COD LOAD VALUE OF THE DAY
385 C      NTQMAX = MAXIMUM TKN LOAD VALUE OF THE DAY
386 C      NO3QAX = MAXIMUM NO3 LOAD VALUE OF THE DAY
387 C      VARSTL = AMPLITUDE OF INFLUENT COD LOAD WAVE (SINE WAVE)
388 C      VARNTL = AMPLITUDE OF INFLUENT TKN LOAD WAVE (SINE WAVE)
389 C      VRNO3L = AMPLITUDE OF INFLUENT NO3 LOAD WAVE (SINE WAVE)
390 C
391 C      PROCESS CHARACTERISTICS
392 C      -----
393 C
394 C      NTDN = LOGICAL NUMBER DESCRIBING PROCESS OPERATION
395 C          IF = 1 PROCESS INCLUDES NITRIFICATION ONLY
396 C          IF = 1 REACTOR IS IN AN AEROBIC ENVIRONMENT
397 C          IF = 0 REACTOR IS IN AN ANOXIC ENVIRONMENT
398 C      TEMP = OPERATING TEMPERATURE OF MIXED LIQUOR
399 C      PH = OPERATING PH OF MIXED LIQUOR
400 C      RSM = MINIMUM SLUDGE AGE FOR NITRIFICATION
401 C      RN = NOMINAL HYDRAULIC RETENTION TIME IN REACTORS
402 C      RA = ACTUAL HYDRAULIC RETENTION TIME IN REACTORS
403 C      TRN = TOTAL HYDRAULIC RETENTION TIME OF PROCESS
404 C      QSR = SLUDGE UNDERFLOW RECYCLE RATE
405 C      QAR = MIXED LIQUOR RECYCLE RATE (A-RECYCLE)
406 C      QBR = MIXED LIQUOR RECYCLE RATE (B-RECYCLE)
407 C      NR = NUMBER OF REACTORS IN SERIES
408 C      NRR = NUMBER OF REACTORS IN SERIES INCLUDING CLARIFIER
409 C      VP = TOTAL VOLUME OF PROCESS
410 C      END = END FEEDING PERIOD (SQ.WAVE)
411 C      WQ = SLUDGE WASTE FLOW
412 C      N.B. THE SLUDGE WASTE FLOW MAY NEVER BE GREATER THAN
413 C           THE INFLUENT FLOW AT ANY INTERVAL OF THE DAY
414 C      I.E. THE EFFLUENT FLOW FROM A REACTOR MAY NEVER BE LESS
415 C           THAN ZERO AT ANY INTERVAL OF THE DAY
416 C
417 C      GENERAL NOTATION FOR PROCESS VARIABLES IN REACTOR
418 C      -----
419 C
420 C      SBP = BIODEGRADABLE PARTICULATE COD
421 C      SBS = BIODEGRADABLE SOLUBLE COD
422 C      SU = UNBIODEGRADABLE COD
423 C      ST = TOTAL COD
424 C      XS = STORED COD
425 C      XA = ACTIVE ORGANISMS
426 C      XE = INERT RESIDUE FROM DECAYING ORGANISMS
427 C      XI = INERT MATERIAL FROM SEWAGE
428 C      XN = ACTIVE NITROSOMONAS
429 C      XV = TOTAL VSS
430 C      OA = OXYGEN CONSUMPTION RATE FOR ADSORPTION OF COD
431 C      OS = OXYGEN CONSUMPTION RATE FOR SYNTHESIS
432 C      OE = OXYGEN CONSUMPTION RATE FOR ENDOGENOUS RESPIRATION
433 C      OC = TOTAL CARBONACEOUS OXYGEN CONSUMPTION RATE
434 C      ON = OXYGEN CONSUMPTION FOR NITRIFICATION
435 C      OT = TOTAL OXYGEN CONSUMPTION RATE
436 C      AMON = AMMONIFICATION RATE (MG N/L/D)
437 C      NSW = NITROGEN REMOVED IN SLUDGE WASTAGE PER DAY
438 C      NT = TOTAL TKN IN REACTOR
439 C      NU = UNBIODEGRADABLE TKN IN REACTOR
440 C      NON = ORGANIC NITROGEN CONCENTRATION IN REACTOR
441 C      NH3 = AMMONIA CONCENTRATION IN REACTOR
442 C      NO3 = NITRATES CONCENTRATION IN REACTOR
443 C      ALK = ALKALINITY CONCENTRATION IN REACTOR
444 C      SCI = ENERGY ENTERING STORAGE FROM LIQUID PHASE (MG-COD/L)
445 C      SCVO = ENERGY REMOVED FROM STORAGE DUE TO CELL SYNTHESIS (MG-V
446 C
447 C      THE SUFFIX 'AV' AFTER A SYMBOL INDICATES DAILY AVERAGE

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448      C          VALUES FOR VARIABLES IN INFLUENT OR REACTORS
449      C          THE SUFFIX 'R' AFTER A SYMBOL INDICATES
450      C          VALUES FOR VARIABLES IN RECYCLE
451      C          THE SUFFIX 'P' AFTER A SYMBOL INDICATES VALUES
452      C          FOR VARIABLES CARRIED OVER TO PLOTTER ROUTINES
453      C          THE SUFFIX 'MAV' AFTER A SYMBOL INDICATES DAILY AVERAGE
454      C          VALUES OF VARIABLES FOR PROCESS
455      C
456      C          BALC = PERCENTAGE COD RECOVERY
457      C          TCODI = TOTAL COD MASS INPUT
458      C          TCODO = TOTAL COD MASS OUTPUT
459      C          TNTI = TOTAL TKN MASS INPUT
460      C          TNTO = TOTAL TKN MASS OUTPUT
461      C          TALKI = TOTAL ALKALINITY MASS INPUT
462      C          TALKO = TOTAL ALKALINITY MASS OUTPUT
463      C          TWCOD = TOTAL COD MASS IN WASTE FLOWS FROM REACTORS
464      C          TWN = TOTAL TKN MASS IN WASTE FLOWS FROM REACTORS
465      C          TWALK = TOTAL ALK MASS IN WASTE FLOWS FROM REACTORS
466      C          WCOD = COD MASS IN WASTE FLOW FROM REACTOR
467      C          ECOD = TOTAL COD MASS IN EFFLUENT
468      C          BALN = PERCENTAGE NITROGEN RECOVERY
469      C          TNTI = TOTAL NITROGEN MASS INPUT
470      C          TNTO = TOTAL NITROGEN MASS OUTPUT
471      C          TWN = TOTAL NITROGEN MASS IN WASTE FLOWS FROM REACTORS
472      C          WN = NITROGEN MASS IN WASTE FLOW FROM REACTOR
473      C          ECOD = TOTAL COD IN EFFLUENT
474      C          EN = TOTAL NITROGEN MASS IN EFFLUENT
475      C          EALK = TOTAL ALKALINTY IN EFFLUENT
476      C
477      IF(TYPE.EQ.1) GO TO 115
478      IF(TYPE.EQ.2) GO TO 116
479      READ(8,100) (Q(1,I),I=1,NL,KH)
480      READ(8,100) (STI(I),I=1,NL,KH)
481      READ(8,100) (NTI(I),I=1,NL,KH)
482      READ(8,100) (NO3I(I),I=1,NL,KH)
483      READ(8,100) (ALKI(I),I=1,NL,KH)
484      IF(TYPE.EQ.3) GO TO 117
485      115 DO 8 I=1,NL,KH
486          KL=(I-1)/KH
487          DUM=SIN(2*PI*(KL-8)/24)
488          Q(1,I)=Q0*(1+VARQ*DUM)
489          STI(I)=STI0*(1+VARSTI*DUM)
490          NTI(I)=NTI0*(1+VARNTI*DUM)
491          NO3I(I)=NO3I0*(1+VNO3I*DUM)
492          ALKI(I)=ALKI0*(1+VALKI*DUM)
493      8 CONTINUE
494      C
495      C          INTERPOLATION OF STI, NO3I, Q AND NTI AT REQUIRED INTERVALS
496      C          -----
497      C
498      117 DO 25 I=1,24
499          KI=KH*I+1
500          KJ=KH*(I-1)+1
501          DIFSTI=(STI(KI)-STI(KJ))/KH
502          DIFFQ=(Q(1,KI)-Q(1,KJ))/KH
503          DIFNTI=(NTI(KI)-NTI(KJ))/KH
504          DNO3I=(NO3I(KI)-NO3I(KJ))/KH
505          DALKI=(ALKI(KI)-ALKI(KJ))/KH
506          DO 25 J=2,KH,1
507              KK=KJ+J-1
508              STI(KK)=STI(KJ)+(J-1)*DIFSTI
509              Q(1,KK)=Q(1,KJ)+(J-1)*DIFFQ
510              NTI(KK)=NTI(KJ)+(J-1)*DIFNTI
511              NO3I(KK)=NO3I(KJ)+(J-1)*DNO3I

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512     ALKI(KK)=ALKI(KJ)+(J-1)*DALKI
513     25 CONTINUE
514     GO TO 118
515     116 PQ=BQ+(VF-BQ)*24.0/LFP
516     DO 24 I=1,NL,1
517     NTI(I)=NTI00
518     STI(I)=STI00
519     Q(1,I)=BQ
520     ALKI(I)=ALKI00
521     24 NO3I(I)=NO3I00
522     NM=START*KH+1
523     END=START+LFP
524     NN=LFP*KH+NM
525     DO 23 I=NM,NN,1
526     STI(I)=STI0
527     Q(1,I)=PQ
528     NTI(I)=NTI0
529     NO3I(I)=NO3I0
530     ALKI(I)=ALKI0
531     23 CONTINUE
532     118 DO 1 I=1,NL
533     STQ(I)=STI(I)*Q(1,I)
534     NTQ(I)=NTI(I)*Q(1,I)
535     NO3Q(I)=NO3I(I)*Q(1,I)
536     ALKQ(I)=ALKI(I)*Q(1,I)
537     SUI(I)=FUS*STI(I)
538     NUI(I)=UN*NTI(I)
539     XII(I)=FUP*STI(I)
540     NXII(I)=XII(I)*FN
541     NH3I(I)=SN*NTI(I)
542     NONI(I)=NTI(I)-NUI(I)-NXII(I)-NH3I(I)
543     1 SBI(I)=STI(I)*(1.0-FUS-FUP*P)
544     C
545     C     READ IN STARTING VALUES OF THE VARIABLES
546     C
547     C
548     READ(8,100)(SBP(J,1),J=1,NR,1)
549     READ(8,100)(SBS(J,1),J=1,NR,1)
550     READ(8,100)(SU(J,1),J=1,NR,1)
551     READ(8,100)(XS(J,1),J=1,NR,1)
552     READ(8,100)(XA(J,1),J=1,NR,1)
553     READ(8,100)(XE(J,1),J=1,NR,1)
554     READ(8,100)(XI(J,1),J=1,NR,1)
555     READ(8,100)(XN(J,1),J=1,NR,1)
556     READ(8,100)(NU(J,1),J=1,NR,1)
557     READ(8,100)(NON(J,1),J=1,NR,1)
558     READ(8,100)(NH3(J,1),J=1,NR,1)
559     READ(8,100)(NO3(J,1),J=1,NR,1)
560     READ(8,100)(ALK(J,1),J=1,NR,1)
561     DO 46 J=1,NR,1
562     XV(J,1)=XS(J,1)+XA(J,1)+XE(J,1)+XI(J,1)
563     46 CONTINUE
564     C
565     C     CALCULATION OF AVERAGE SEWAGE CHARACTERISTICS
566     C     -----
567     C
568     SUMQ=0
569     SUMSTI=0
570     SUMNTI=0
571     SNO3I=0.0
572     SALKI=0.0
573     QMAX=0.0
574     STIMAX=0.0
575     SUMSTQ=0.0

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576      STQMAX=0.0
577      SUMNTQ=0.0
578      NTQMAX=0.0
579      NTIMAX=0.0
580      SNO3Q=0.0
581      NO3QAX=0.0
582      NO3IAX=0.0
583      SALKQ=0.0
584      ALKQAX=0.0
585      ALKIAX=0.0
586      DO 2 I=1,D,1
587      SUMNTI=SUMNTI+NTI(I)
588      SNO3I=SNO3I+NO3I(I)
589      SALKI=SALKI+ALKI(I)
590      SUMSTI=SUMSTI+STI(I)
591      SUMSTQ=SUMSTQ+STQ(I)
592      SUMNTQ=SUMNTQ+NTQ(I)
593      SNO3Q=SNO3Q+NO3Q(I)
594      SALKQ=SALKQ+ALKQ(I)
595      IF(STQ(I).GT.STQMAX) STQMAX=STQ(I)
596      IF(NTQ(I).GT.NTQMAX) NTQMAX=NTQ(I)
597      IF(NO3Q(I).GT.NO3QAX) NO3QAX=NO3Q(I)
598      IF(ALKQ(I).GT.ALKQAX) ALKQAX=ALKQ(I)
599      IF(Q(1,I).GT.QMAX) QMAX=Q(1,I)
600      IF(STI(I).GT.STIMAX) STIMAX=STI(I)
601      IF(NTI(I).GT.NTIMAX) NTIMAX=NTI(I)
602      IF(NO3I(I).GT.NO3IAX) NO3IAX=NO3I(I)
603      IF(ALKI(I).GT.ALKIAI) ALKIAI=ALKI(I)
604      2 SUMQ=SUMQ+Q(1,I)
605      QAV(1)=SUMQ/D
606      STQAV=SUMSTQ/D
607      NTQAV=SUMNTQ/D
608      NO3QAV=SNO3Q/D
609      ALKQAV=SALKQ/D
610      IF(TYPE.EQ.2) GO TO 119
611      NTIAV=SUMNTI/D
612      STIAV=SUMSTI/D
613      NO3IAV=SNO3I/D
614      ALKIAV=SALKI/D
615      GO TO 120
616      119 STIAV=STQAV/VF
617      NTIAV=NTQAV/VF
618      NO3IAV=NO3QAV/VF
619      ALKIAV=ALKQAV/VF
620      120 SBIAV=STIAV*(1.0-FUS-FUP*P)
621      SUIAV=FUS*STIAV
622      NUIAV=NTIAV*UN
623      XIIAV=FUP*STIAV
624      NXIIAV=FN*XIIAV
625      NH3IAV=SN*NTIAV
626      NONIAV=NTIAV-NXIIAV-NUIAV-NH3IAV
627      QSR=SR*QAV(1)
628      QAR=AR*QAV(1)
629      QBR=BR*QAV(1)
630      IF(TYPE.EQ.2) GO TO 121
631      VARSTL=STQMAX/STQAV-1.00
632      VARNTL=NTQMAX/NTQAV-1.00
633      VRNO3L=NO3QAX/NO3QAV-1.00
634      IF(VRALKL.LT.0.0) VRALKL=0.0
635      IF(VRNO3L.LT.0.0) VRNO3L=0.0
636      IF(VRALKL.LT.0.0) VRALKL=0.0
637      IF(TYPE.EQ.1) GO TO 121
638      VARQ=QMAX/QAV(1)-1.00
639      VARSTI=STIMAX/STIAV-1.00

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640      VARNTI=NTIMAX/NTIAV-1.00
641      VNO3I=NO3IAX/NO3IAV-1.00
642      VALKI=ALKIAX/ALKIAV-1.00
643      IF(VNO3I.LT.0.0) VNO3I=0.0
644      IF(VALKI.LT.0.0) VALKI=0.0
645 121 CONTINUE
646 C
647 C      CALCULATE TOTAL AND AEROBIC VOLUMES
648 C      -----
649 C
650      DO 37 J=1,NR,1
651      IFREQ=24/(ANOX(J)+AROB(J))
652      DO 37 I=1,NL,1
653      NTDN(J,I)=1
654 37 CONTINUE
655      VP=0.0
656      DO 133 J=1,NR,1
657      VP=VP+V(J)
658      DO 133 I=1,NL,1
659      DO 133 II=1,IFREQ,1
660      IB=(INIT(J)+(II-1)*(ANOX(J)+AROB(J)))*KH+1
661      IE=(INIT(J)+(II-1)*(ANOX(J)+AROB(J))+ANOX(J))*KH+1
662      IF(I.GE.IB.AND.I.LT.IE) NTDN(J,I)=0
663      NTDN(J,NL)=NTDN(J,1)
664 133 CONTINUE
665      JJ=3
666      DO 77 J=1,NR,1
667      INTDN(J)=0
668      DO 77 I=1,NL,1
669      INTDN(J)=INTDN(J)+NTDN(J,I)
670 77 CONTINUE
671      SANOX=0.0
672      DO 78 J=1,NR,1
673      IF(INTDN(J).EQ.0) JJ=1
674      IF(INTDN(J).EQ.NL) JJ=2
675      NAME(J)=LNAME(JJ)
676      FANOX(J)=1.0-INTDN(J)/NL
677      SANOX=SANOX+FANOX(J)
678 78 CONTINUE
679      LNAME(1)='AROB'
680      LNAME(2)='ANOX'
681      LNAME(3)='ANAE'
682 C
683 C      ADJUST TEMPERATURE AND PH DEPENDENT CONSTANTS
684 C      -----
685 C
686      IF(PH.LT.8.0) UM20=UMAX
687      IF(PH.LT.7.2) UM20=UMAX*(PHIN)**(PH-7.20)
688      IF(PH.LT.8.0) KN20=KNMAX
689      IF(PH.LT.7.2) KN20=KNMAX*(PHIN)**(7.20-PH)
690      UMT=UM20*(THEN)**(TEMP-20.0)
691      KNT=KN20*(THEN)**(TEMP-20.0)
692      KSPT=KSP20*(THES)**(20.0-TEMP)
693      KMPT=KMP20*(THEM)**(TEMP-20.0)
694      KSST=KSS20*(THEZ)**(TEMP-20.0)
695      KMST=KMS20*(THEA)**(TEMP-20.0)
696      KRT=KR20*(THEE)**(TEMP-20.0)
697      KVT=KV20*(THEE)**(TEMP-20.0)
698      BHT=BH20*(THEE)**(TEMP-20.0)
699      BNT=BN20*(THEE)**(TEMP-20.0)
700      RSM=1.0/((UMT/(1.0+(KNT/(SN*NTIAV)))))-BNT)
701      RSA=(1.0-SANOX)*RS
702      TRN=VP/QAV(1)
703      FNS=NONIAV/SBIAV

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704      FACTC=STQAV/(STIAV*QAV(1))
705      FACTN=NTQAV/(NTIAV*QAV(1))
706      CC=(1.0-F)*BHT*YH*RS/(1.0+BHT*(1.0-YH*P*(1.0-F))*RS)
707      FNS=(FNS*(1.0-FBS)+FN*CC)/((1.0-FBS)+P*CC)
708  C
709  C      CALCULATE INTEGRATION STEP LENGTHS
710  C      -----
711      DT=1./D
712      DDT=1./DD
713      RDT=DDT/DT
714  C
715  C      RELATE STEP INTERVALS TO THE TIME BASE
716  C      -----
717  C
718      II=0
719      KN=M/D
720      KHL=KH+1
721      DO 27 I=1,NL,1
722      JE=I-1
723      IF(JE.EQ.0) GO TO 3
724      KO(I)=KO(I-1)+KN
725      GO TO 4
726  3 KO(I)=KOM
727  4 II=II+1
728      IF (II.EQ.KHL) KO(I)=KO(I)+40
729      IF (II.EQ.KHL) II=1
730      IF (KO(I).GE.2400) KO(I)=KO(I)-2400
731      IF(KO(I).EQ.ICSD) IC=I-1
732      IF(KOM.EQ.ICSD) IC=1
733      IF(KO(I).EQ.ISSD) IS=I
734  27 CONTINUE
735      HK=IS-IC
736      DO 9 J=1,NR,1
737      DO 29 I=1,NL,1
738      WQ(J,I)=0.0
739  29 CONTINUE
740      IF(LRSD(J).EQ.0) GO TO 32
741      DO 28 I=IC,IS,1
742      WQ(J,I)=VP/RS*D/HK*1.0/NRSD
743  28 CONTINUE
744  32 DO 48 I=1,NL,1
745      Q(J+1,I)=Q(J,I)-WQ(J,I)
746      IF(Q(J+1,I).GT.0.00) GO TO 48
747      Q(J+1,I)=0.00
748      WQ(J,I)=Q(J,I)
749  48 CONTINUE
750  9 CONTINUE
751  C
752  C      SET INITIAL CONDITIONS
753  C      -----
754  C
755      DUM=(QSR+Q(NRR,1))/QSR
756      SBPR(1)=SBP(NR,1)*DUM
757      XSR(1)=XS(NR,1)*DUM
758      XAR(1)=XA(NR,1)*DUM
759      XER(1)=XE(NR,1)*DUM
760      XIR(1)=XI(NR,1)*DUM
761      XNR(1)=XN(NR,1)*DUM
762      DO 5 J=1,NR,1
763      WRITE(5,200) SBS(J,1),SBP(J,1),XS(J,1),XA(J,1),XE(J,1),XI(J,1),
764      1XV(J,1),XN(J,1),NO3(J,1),ALK(J,1),SU(J,1),NON(J,1),NH3(J,1)
765  5 CONTINUE
766  200 FORMAT(1H ,5X,13F7.2)
767      DO 15 I=1,D,1

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768      17 J=0
769      19 J=J+1
770      C*****
771      IF(NTDN(J,I).EQ.1) CALL UAERO(D,DT,J,FR,FBS,FCS,KVT,KMPT,KSPT,
772      1KMST,KSST,P,NR,YH,BHT,F,KRT,FOE,FN,FOS,FNS,NJ,DDT,UMT,YN,KNT,
773      2RDT,BNT,FACT,LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR,I)
774      C*****
775      IF(NTDN(J,I).EQ.0) CALL UANOX(D,DT,J,FR,FBS,FCS,KVT,KMPT,KSPT,
776      1KMST,KSST,P,NR,YH,BHT,F,KRT,FOE,FN,FOS,FNS,NJ,DDT,UMT,YN,KNT,
777      2RDT,BNT,FACT,LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR,I)
778      C*****
779      IF(J.EQ.NR) GO TO 20
780      GO TO 19
781      20 CONTINUE
782      C*****
783      CALL USETL(NL,J,QSR,I)
784      C*****
785      15 CONTINUE
786      DO 26 J=1,NR,1
787      DO 26 I=1,NL
788      JA=I-1
789      IF (JA.EQ.0) JA=D
790      JB=I+1
791      IF (JB.EQ.NLL) JB=2
792      JD=I
793      IF(JD.EQ.NL) JD=1
794      RXS(J,I)=(XS(J,JB)-XS(J,JA))/(DT*2.0)
795      ST(J,I)=SU(J,I)+SBS(J,I)
796      NT(J,I)=NON(J,I)+NU(J,I)+NH3(J,I)
797      OC(J,I)=(1.0-P*YH)*SYNTH(J,JD)/(DT*24.0)
798      ANPOT(J,I)=ANPOT(J,JD)/(DT*24.0)
799      ON(J,I)=4.57*NIT(J,JD)/(DT*24.0)
800      OT(J,I)=OC(J,I)+ON(J,I)
801      AMON(J,I)=AMON(J,JD)/(DT*24.0)
802      26 CONTINUE
803      C
804      C      CALCULATION OF AVERAGE RESULTS FROM DYNAMIC MODEL
805      C      -----
806      C      EXAMPLE:  XVAV = AVERAGE XV
807      C
808      TRN=0.0
809      SUMXVT=0.0
810      SUMWST=0.0
811      SUMXAM=0.0
812      SUMXEM=0.0
813      SUMXIM=0.0
814      SUMXSM=0.0
815      SUMXNM=0.0
816      SUMODM=0.0
817      SMAPT=0.0
818      APTMNR=0.0
819      RNO3AV=0.0
820      SUMOCM=0.0
821      SUMONM=0.0
822      SUMOTM=0.0
823      SUMAMM=0.0
824      SUNSWM=0.0
825      TWCOD=0.0
826      TWN=0.0
827      TWALK=0.0
828      EN=0.0
829      ENH3=0.0
830      ENON=0.0
831      ENO3=0.0

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832      EALK=0.0
833      ENT=0.0
834      ECOD=0.0
835      DO 31 J=2,NRR,1
836      SUMQ=0.0
837      DO 30 I=1,D,1
838 30    SUMQ=SUMQ+Q(J,I)
839 31    QAV(J)=SUMQ/D
840      DO 22 J=1,NR,1
841      SUMWS=0.0
842      SUMNSW=0.0
843      SUMNU=0.0
844      SUMXA=0.0
845      SUMXE=0.0
846      SUMXI=0.0
847      SUMSBP=0.0
848      SUMSBS=0.0
849      SUMXV=0.0
850      SUMXS=0.0
851      SUMXN=0.0
852      SUMOC=0.0
853      SUMOD=0.0
854      SUMAPT=0.0
855      SUMON=0.0
856      SUMSU=0.0
857      SUMRXS=0.0
858      SUMNON=0.0
859      SUMNH3=0.0
860      SUMNT=0.0
861      SUMNO3=0.0
862      SUMALK=0.0
863      SUMOT=0.0
864      SUMAM=0.0
865      WCOD=0.0
866      WN=0.0
867      WALK=0.0
868      WNON=0.0
869      DO 21 I=1,D
870      SUMWS=SUMWS+XV(J,I)*WQ(J,I)*DT
871      SUMNSW=SUMNSW+(FN*(XA(J,I)+XE(J,I)+XI(J,I))+FNS*XS(J,I))
872 1*WQ(J,I)*DT
873      IF(J.NE.NR) GO TO 11
874      ECOD=ECOD+Q(J+1,I)*ST(J,I)*DT/QAV(J+1)
875      ESBS=ESBS+Q(J+1,I)*(SBS(J,I))*DT/QAV(J+1)
876      ESU=ESU+Q(J+1,I)*(SU(J,I))*DT/QAV(J+1)
877      EN=EN+Q(J+1,I)*(NT(J,I)+NO3(J,I))*DT/QAV(J+1)
878      ENH3=ENH3+Q(J+1,I)*(NH3(J,I))*DT/QAV(J+1)
879      ENON=ENON+Q(J+1,I)*(NON(J,I))*DT/QAV(J+1)
880      ENO3=ENO3+Q(J+1,I)*(NO3(J,I))*DT/QAV(J+1)
881      EALK=EALK+Q(J+1,I)*(ALK(J,I))*DT/QAV(J+1)
882      ENT=ENT+Q(J+1,I)*(NT(J,I))*DT/QAV(J+1)
883 11    SUMNU=SUMNU+NU(J,I)
884      SUMXA=SUMXA+XA(J,I)
885      SUMXE=SUMXE+XE(J,I)
886      SUMXI=SUMXI+XI(J,I)
887      SUMSBP=SUMSBP+SBP(J,I)
888      SUMSBS=SUMSBS+SBS(J,I)
889      SUMXV=SUMXV+XV(J,I)
890      SUMXN=SUMXN+XN(J,I)
891      SUMXS=SUMXS+XS(J,I)
892      SUMOC=SUMOC+OC(J,I)
893      SUMAPT=SUMAPT+ANPOT(J,I)
894      IF(NTDN(J,I).EQ.1) GO TO 12
895      SUMOD=SUMOD+OC(J,I)

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896      12 SUMON=SUMON+ON(J,I)
897      SUMSU=SUMSU+SU(J,I)
898      SUMRXS=SUMRXS+RXS(J,I)
899      SUMNO3=SUMNO3+NO3(J,I)
900      SUMALK=SUMALK+ALK(J,I)
901      SUMNON=SUMNON+NON(J,I)
902      SUMNH3=SUMNH3+NH3(J,I)
903      SUMNT=SUMNT+NT(J,I)
904      WCOD=WCOD+WQ(J,I)*(ST(J,I)+SBP(J,I))*DT
905      WN=WN+WQ(J,I)*(NT(J,I)+NO3(J,I))*DT
906      WALK=WALK+WQ(J,I)*ALK(J,I)*DT
907      WNON=WNON+WQ(J,I)*NON(J,I)*DT
908      SUMOT=SUMOT+OT(J,I)
909      SUMAM=SUMAM+AMON(J,I)
910      21 CONTINUE
911      NUAV(J)=SUMNU/D
912      XAAV(J)=SUMXA/D
913      XEAV(J)=SUMXE/D
914      XIAV(J)=SUMXI/D
915      SBPAV(J)=SUMSBP/D
916      SBSAV(J)=SUMSBS/D
917      XVAV(J)=SUMXV/D
918      XSAV(J)=SUMXS/D
919      XNAV(J)=SUMXN/D
920      OCAV(J)=SUMOC/D
921      ODAV(J)=SUMOD/D
922      APTAV(J)=SUMAPT/D
923      APTNR(J)=APTAV(J)*24.0*V(J)/QAV(1)
924      RNO3(J)=ODAV(J)*24.0*V(J)/(QAV(1)*2.85)
925      ONAV(J)=SUMON/D
926      SUAV(J)=SUMSU/D
927      RXSAV(J)=SUMRXS/D
928      STAV(J)=SUAV(J)+SBSAV(J)
929      NSWAV(J)=SUMNSW/QAV(1)
930      SUNSWM=SUNSWM+NSWAV(J)
931      OTAV(J)=SUMOT/D
932      AMAV(J)=SUMAM/D
933      NO3AV(J)=SUMNO3/D
934      ALKAV(J)=SUMALK/D
935      NONAV(J)=SUMNON/D
936      NH3AV(J)=SUMNH3/D
937      NTAV(J)=SUMNT/D
938      SUMWST=SUMWST+SUMWS
939      SUMXVT=SUMXVT+XVAV(J)*V(J)
940      SUMXAM=SUMXAM+XAAV(J)*V(J)
941      SUMXEM=SUMXEM+XEAV(J)*V(J)
942      SUMXIM=SUMXIM+XIAV(J)*V(J)
943      SUMXSM=SUMXSM+XSAV(J)*V(J)
944      SUMXNM=SUMXNM+XNAV(J)*V(J)
945      SUMODM=SUMODM+ODAV(J)*V(J)
946      SMAPTM=SMAPTM+APTAV(J)*V(J)
947      APTMNR=APTMNR+APTNR(J)
948      RNO3AV=RNO3AV+RNO3(J)
949      SUMOCM=SUMOCM+OCAV(J)*V(J)
950      SUMONM=SUMONM+ONAV(J)*V(J)
951      SUMOTM=SUMOTM+OTAV(J)*V(J)
952      SUMAMM=SUMAMM+AMAV(J)*V(J)
953      I=NL
954      Q(J,I)=QAV(J)
955      C*****
956      CALL UDUMV(J,I,DUMIF,DUMS,DUMA,DUMB,DUMIR,DUMO,
957      1LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR,
958      1DT,DDT,RDT)
959      C*****

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960      RA(J)=24.0*DDT/DUMO
961      RN(J)=V(J)/QAV(J)*24.0
962      TWCOD=TWCOD+WCOD
963      TWN=TWN+WN
964      TWALK=TWALK+WALK
965      TWNON=TWNON+WNON
966      TRN=TRN+RN(J)
967      IF(NTDN(J,I).EQ.1) GO TO 22
968      SUMXVA=SUMXVA+XVAV(J)*V(J)
969 22    CONTINUE
970      RSAV=SUMXVT/SUMWST
971      XAMAV=SUMXAM/VP
972      XEMAV=SUMXEM/VP
973      XIMAV=SUMXIM/VP
974      XSMAY=SUMXSM/VP
975      XVMAY=SUMXVT/VP
976      XNMAV=SUMXNM/VP
977      ODMAY=SUMODM/VP
978      APTMAV=SMAPTM/VP
979      OCMAY=SUMOCM/VP
980      ONMAV=SUMONM/VP
981      OTMAV=SUMOTM/VP
982      AMMAV=SUMAMM/VP
983      CODSI=0.0
984      NTITI=0.0
985      CODSO=0.0
986      NTITO=0.0
987      ALKSI=0.0
988      ALKSO=0.0
989      DO 5000 J=1,NR,1
990      CODSI=CODSI+(XV(J,1)*P+SBP(J,1)+ST(J,1))*V(J)
991      NTITI=NTITI+((XA(J,1)+XE(J,1)+XI(J,1))*FN+XS(J,1)*FNS
992      &+NON(J,1)+NU(J,1)+NH3(J,1)+NO3(J,1))*V(J)
993      CODSO=CODSO+(XV(J,D)*P+SBP(J,D)+ST(J,D))*V(J)
994      NTITO=NTITO+((XA(J,D)+XE(J,D)+XI(J,D))*FN+XS(J,D)*FNS
995      &+NCN(J,D)+NU(J,D)+NH3(J,D)+NO3(J,D))*V(J)
996      ALKSI=ALKSI+ALK(J,1)*V(J)
997      ALKSO=ALKSO+ALK(J,D)*V(J)
998 5000 CONTINUE
999  C
1000  C   CHECK MASS BALANCES ON COD,NITROGEN AND ALKALINITY
1001  C   1.   COD
1002  C   -----
1003      TCODI=STQAV+CODSI
1004      TCODO=P*SUMWST+SUMOCM*24.0+TWCOD+ECOD*QAV(NRR)+CODSO
1005      BALC=TCODO*100.0/TCODI
1006  C   2.   NITROGEN
1007  C   -----
1008      TNTI=NTQAV+NO3QAV+NTITI
1009      TNTO=SUNSWM*QAV(1)+TWN+EN*QAV(NRR)+SUMODM*24/2.85+NTITO
1010      BALN=TNTO*100.0/TNTI
1011  C   3.   ALKALINITY
1012  C   -----
1013      TALKI=ALKQAV+ALKSI
1014      TALKO=TWALK+EALK*QAV(NRR)+ALKSO+7.14*SUMONM*24.0/4.57
1015      &-3.57*(SUMODM*24.0/2.86+SUMAMM*24.0)
1016      BALALK=TALKO/TALKI*100.0
1017  C
1018  C
1019  C
1020  C   OUTPUT AND FORMAT STATEMENTS
1021  C   -----
1022  C
1023      IF(RESULT.EQ.2)GO TO 126

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1024      WRITE(5,98)DATE,XMONTH,YEAR
1025 98      FORMAT(1H0,5X,'DAY OF EXPERIMENT:',1X,A2,1X,'OF',1X,A3,1X,A4)
1026      WRITE(5,201)TEMP,PH,YH,BHT,KVT,YN,THEN,F,FBS,FN,KNT,PHIN,
1027      1THEE,P,FOE,UMT,BNT,FOS,KRT,KSPT,THEZ,KMPT,THEA,THES,
1028      2KMST,KSST,FR,FNS,RS
1029 201     FORMAT(1H1,20X,22('*'),/,1H,20X,22('*'),/,
1030      11H0,10X,'KINETIC CONSTANTS',/,1H,10X,17('-'),/,
1031      21H,5X,'PROCESS OPERATING TEMPERATURE=',F5.1,1X,'DEG. C',/,
1032      31H,28X,'AND PH=',F5.2,/,1H0,5X,'HETEROTROPHS',28X,'AUTOTROPHS',/,
1033      41H,5X,12('-'),28X,10('-'),/,1H,6X,'YH=',F5.3,4X,'BH=',F5.3,4X,
1034      5'KV=',F5.3,8X,'YN=',F5.3,6X,'THEN=',F5.3,/,1H,7X,'F=',F5.3,3X,
1035      6'FBS=',F5.3,4X,'FN=',F5.3,8X,'KN=',F5.2,6X,'PHIN=',F5.3,/,
1036      75X,'THEE=',F5.3,5X,'P=',F5.3,3X,'FOE=',F5.3,8X,'UM=',F5.3,
1037      28X,'BN=',F5.3,/,
1038      86X,'FOS=',F5.3,4X,'KR=',F5.3,3X,'KSP=',F5.3,6X,'THEZ=',F5.3,/,
1039      16X,'KMP=',F5.3,2X,'THEA=',F5.3,2X,'THES=',F5.3,/,
1040      26X,'KMS=',F5.2,3X,'KSS=',F5.1,/,
1041      27X,'FR=',F5.3,3X,'FNS=',F5.3,/,
1042      11H,5X,'OPERATING SLUDGE AGE OF PROCESS=',F5.2,1X,'DAYS')
1043      WRITE(5,202)FUP,FUS,SN,UN,QAV(1),STQAV,NTQAV,STIAV,SBIIV,SUIIV,
1044      1XIIIV,NTIIV,NH3IIV,NONIIV,NUIIV,NO3IIV,ALKIIV
1045 202     FORMAT(1H0,10X,'AVERAGE COMPOSITION OF SEWAGE',/,
1046      11H,10X,29('-'),/,1H0,5X,'SOLID INERT FRACTION OF INFLUENT COD=',
1047      2F5.3,/,1H,5X,'SOLUBLE INERT FRACTION OF INFLUENT COD=',F5.3,/,
1048      31H,5X,'AMMONIA TO TKN FRACTION=',F5.3,/,
1049      41H,5X,'INERT TKN FRACTION=',F5.3,/,1H0,4X,'QAV=',F9.5,
1050      15X,'STQAV=',E13.5,5X,'NTQAV=',E13.5,/,
1051      51H,5X,'STI=',F7.2,5X,'SBI=',F7.2,4X,'SUI=',F6.2,
1052      64X,'XII=',F6.2,/,1H,5X,'NTI=',F6.2,4X,'NH3I=',F6.2,
1053      74X,'NONI=',F5.2,5X,'NUI=',F5.2,5X,'NO3I=',F5.2,/,
1054      84X,'ALKI=',F7.2)
1055      IF(TYPE.EQ.1) GO TO 122
1056      IF(TYPE.EQ.2) GO TO 123
1057      IF(TYPE.EQ.3) GO TO 122
1058 122     WRITE(5,205)VARQ,VARSTI,VARNTI,VNO3I,VARSTL,VARNTL,VNO3L
1059 205     FORMAT(1H0,10X,'SINE WAVE INFLUENT FLOW AND FEED PATTERN',/,
1060      11H,10X,40('-'),/,
1061      41H0,10X,'AMPLITUDE OF FLOW WAVE=',F5.2,4X,'(FRAC OF AVE)',/,
1062      51H,10X,'AMPLITUDE OF COD WAVE=',F5.2,4X,'(FRAC OF AVE)',/,
1063      61H,10X,'AMPLITUDE OF TKN WAVE=',F5.2,4X,'(FRAC OF AVE)',/,
1064      71H,10X,'AMPLITUDE OF NO3 WAVE=',F5.2,4X,'(FRAC OF AVE)',/,
1065      81H,10X,'AMPLITUDE OF COD LOAD WAVE=',F5.2,4X,'(FRAC OF AVE)',/,
1066      91H,10X,'AMPLITUDE OF TKN LOAD WAVE=',F5.2,4X,'(FRAC OF AVE)',/,
1067      01H,10X,'AMPLITUDE OF NO3 LOAD WAVE=',F5.2,4X,'(FRAC OF AVE)')
1068 204     FORMAT(1H0,10X,'PROCESS CONFIGURATION DATA',/,1H,5X,26('-'),/,
1069      11H0,5X,'NUMBER OF TANKS IN SERIES',I2,/,
1070      11H,5X,'INFLUENT FLOW INTO TANK',I2,/,
1071      11H,5X,'S-RECYCLE FROM SETTLER TO TANK',I2,/,
1072      11H,5X,'A-RECYCLE FROM TANK',I2,1X,'INTO TANK',I2,/,
1073      11H,5X,'B-RECYCLE FROM TANK',I2,1X,'INTO TANK',I2,/,
1074      11H,5X,'S RECYCLE RATIO=',F4.1,/,
1075      11H,5X,'A RECYCLE RATIO=',F4.1,/,
1076      11H,5X,'B RECYCLE RATIO=',F4.1)
1077      GO TO 124
1078 123     WRITE(5,206)LFP,PQ,BQ,VF
1079 206     FORMAT(1H0,10X,'SQUARE WAVE INFLUENT FLOW AND FEED PATTERN',/,
1080      11H,10X,42('-'),/,
1081      11H,10X,'LENGTH OF FEED PERIOD (HRS)=',F5.2,/,
1082      21H,10X,'PEAK FLOW (L/D)=',F9.5,/,
1083      31H,10X,'BASE FLOW (L/D)=',F9.5,/,
1084      41H,10X,'VOLUME OF FEED PER DAY (LITRES)=' F9.5)
1085 124     WRITE(5,203) TRN,VP,ANOXAV,RSIV
1086      WRITE(5,204) NR,LINI,LSRI,LARO,LARI,LBRO,LBRI,SR,AR,BR
1087 203     FORMAT(1H1,10X,21('-'),/,1H,10X,'DAILY AVERAGE RESULTS',/,

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1088      11H ,10X,21('-'),/,
1089      11H0,5X,'TOTAL HYDRAULIC RETENTION TIME=',F8.3,1X,'HOURS',/,
1090      11H ,5X,'TOTAL VOLUME=',E13.5,1X,'LITRES',/,
1091      11H ,5X,'ANOXIC FRACTION=',F5.3,/,
1092      21H ,5X,'SLUDGE AGE =',F5.2,1X,'DAYS')
1093      WRITE(5,244)TCODI,TCODO,BALC,TNTI,TNTO,BALN,TALKI,TALKO,BALALK
1094 244 FORMAT(1H0,10X,'MASS BALANCES ON COD AND NITROGEN',/,
1095      11H ,10X,33('-'),/,
1096      21H0,5X,'TOTAL COD MASS INPUT=',1X,E13.6,1X,'(MG-COD/D)',/,
1097      21H ,5X,'TOTAL COD MASS OUTPUT=',1X,E13.6,1X,'(MG-COD/D)',/,
1098      31H ,5X,'PERCENTAGE COD RECOVERY=',1X,F7.3,1X,'(%)',/,
1099      21H ,5X,'TOTAL NITROGEN MASS INPUT=',1X,E13.6,1X,'(MG-N/D)',/,
1100      21H ,5X,'TOTAL NITROGEN MASS OUTPUT=',1X,E13.6,1X,'(MG-N/D)',/,
1101      31H ,5X,'PERCENTAGE NITROGEN RECOVERY=',1X,F7.3,1X,'(%)',/,
1102      21H ,5X,'TOTAL ALKALINITY MASS INPUT=',1X,E13.6,1X,'(CACO3/D)',/,
1103      21H ,5X,'TOTAL ALKALINITY MASS OUTPUT=',1X,E13.6,1X,'(CACO3/D)',/,
1104      31H ,5X,'PERCENTAGE ALKALINITY RECOVERY=',1X,F7.3,1X,'(%)')
1105      WRITE(5,300)
1106 300 FORMAT(1H0,30X,'TANK 1',6X,'TANK 2',
1107      26X,'TANK 3',6X,'TANK 4',6X,'TANK 5',6X,'TANK 6',
1108      36X,'MEAN',/,1H ,30X,7('-'),6X,7('-'),6X,7('-'),6X,7('-'),
1109      46X,7('-'),6X,7('-'),6X,4('-'))
1110      WRITE(5,327) (NAME(J),J=1,NR,1)
1111 327 FORMAT(1H ,23X,6A13)
1112      WRITE(5,328)
1113 328 FORMAT(1H ,30X,6('-'),7X,6('-'),7X,6('-'),
1114      17X,6('-'),7X,6('-'),7X,6('-'))
1115      WRITE(5,301) (XAAV(J),J=1,NR,1),XAMAV
1116      WRITE(5,302) (XSAV(J),J=1,NR,1),XSMAV
1117      WRITE(5,303) (XEAV(J),J=1,NR,1),XEMAV
1118      WRITE(5,304) (XIAV(J),J=1,NR,1),XIMAV
1119      WRITE(5,305) (XVAV(J),J=1,NR,1),XVMAV
1120      WRITE(5,319) (XNAV(J),J=1,NR,1),XNMAV
1121      WRITE(5,313) (ODAV(J),J=1,NR,1),ODMAV
1122      WRITE(5,314) (OCAV(J),J=1,NR,1),OCMAV
1123      WRITE(5,315) (ONAV(J),J=1,NR,1),ONMAV
1124      WRITE(5,316) (OTAV(J),J=1,NR,1),OTMAV
1125      WRITE(5,3160) (AMAV(J),J=1,NR,1),AMMAV
1126      WRITE(5,309) (NSWAV(J),J=1,NR,1),SUNSWM
1127      WRITE(5,321) (SBPAV(J),J=1,NR,1)
1128      WRITE(5,320) (SBSAV(J),J=1,NR,1)
1129      WRITE(5,324) (SUAV(J),J=1,NR,1)
1130      WRITE(5,311) (STAV(J),J=1,NR,1)
1131      WRITE(5,306) (NONAV(J),J=1,NR,1)
1132      WRITE(5,310) (NH3AV(J),J=1,NR,1)
1133      WRITE(5,323) (NUAV(J),J=1,NR,1)
1134      WRITE(5,312) (NTAV(J),J=1,NR,1)
1135      WRITE(5,307) (NO3AV(J),J=1,NR,1)
1136      WRITE(5,703) (ALKAV(J),J=1,NR,1)
1137      WRITE(5,308) (RNO3(J),J=1,NR,1),RNO3AV
1138      WRITE(5,326) (APTAV(J),J=1,NR,1),APTMMAV
1139      WRITE(5,329) (APTNR(J),J=1,NR,1),APTMNR
1140      WRITE(5,322) (V(J),J=1,NR,1)
1141      WRITE(5,325) (QAV(J),J=1,NRR,1)
1142      WRITE(5,317) (RA(J),J=1,NR,1)
1143      WRITE(5,318) (RN(J),J=1,NR,1)
1144      WRITE(5,3003) (FANOX(J),J=1,NR,1),ANOXAV
1145      WRITE(5,3000)ECOD,ESBS,ESU,EN,ENT,ENH3,ENON,ENO3,EALK
1146 3000 FORMAT(1H0,8X,'AVERAGE EFFLUENT CONCENTRATIONS',/,
1147      &8X,'TOT-COD=',E13.5,/,
1148      &8X,'SBS',E13.5,/,
1149      &8X,'SU',E13.5,/,
1150      &8X,'TOTAL-N=',E13.5,/,
1151      &8X,'TKN',E13.5,/,

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1152      &8X,'NH3',E13.5,/,
1153      &8X,'NON',E13.5,/,
1154      &8X,'NO3',E13.5,/,
1155      &8X,'ALK',E13.5)
1156 301 FORMAT(1H ,8X,'XA (MG-VSS/L)',2X,7(E13.5))
1157 302 FORMAT(1H ,8X,'XS (MG-VSS/L)',2X,7(E13.5))
1158 303 FORMAT(1H ,8X,'XE (MG-VSS/L)',2X,7(E13.5))
1159 304 FORMAT(1H ,8X,'XI (MG-VSS/L)',2X,7(E13.5))
1160 305 FORMAT(1H ,8X,'XV (MG-VSS/L)',2X,7(E13.5))
1161 319 FORMAT(1H ,8X,'XN (MG-VSS/L)',2X,7(E13.5))
1162 313 FORMAT(1H ,8X,'OD (MG-O/L/HR)',2X,7(E13.5))
1163 314 FORMAT(1H ,8X,'OC (MG-O/L/HR)',2X,7(E13.5))
1164 315 FORMAT(1H ,8X,'ON (MG-O/L/HR)',2X,7(E13.5))
1165 316 FORMAT(1H ,8X,'OT (MG-O/L/HR)',2X,7(E13.5))
1166 3160 FORMAT(1H ,8X,'AM (MG-N/L/HR)',2X,7(E13.5))
1167 309 FORMAT(1H ,8X,'NSW (MG-N/L/D)',2X,7(E13.5))
1168 321 FORMAT(1H ,8X,'SBP (MG-COD/L)',2X,6(E13.5))
1169 320 FORMAT(1H ,8X,'SBS (MG-COD/L)',2X,6(E13.5))
1170 324 FORMAT(1H ,8X,'SU (MG-COD/L)',2X,6(E13.5))
1171 311 FORMAT(1H ,8X,'ST (MG-COD/L)',2X,6(E13.5))
1172 306 FORMAT(1H ,8X,'NON (MG-N/L)',2X,6(E13.5))
1173 310 FORMAT(1H ,8X,'NH3 (MG-N/L)',2X,6(E13.5))
1174 323 FORMAT(1H ,8X,'NU (MG-N/L)',2X,6(E13.5))
1175 312 FORMAT(1H ,8X,'NT (MG-N/L)',2X,6(E13.5))
1176 307 FORMAT(1H ,8X,'NO3 (MG-N/L)',2X,6(E13.5))
1177 703 FORMAT(1H ,8X,'ALK (MG-CACO3/L)',2X,6(E13.5))
1178 326 FORMAT(1H ,8X,'APT (MG-N/L/HR)',2X,7(E13.5))
1179 329 FORMAT(1H ,8X,'APTNR (MG-N/L)',2X,7(E13.5))
1180 308 FORMAT(1H ,8X,'RNO3 (MG-N/L)',2X,7(E13.5))
1181 322 FORMAT(1H ,8X,'VOL (LITRES)',2X,6(E13.5))
1182 325 FORMAT(1H ,8X,'FLOW (L/D)',2X,7(E13.5))
1183 317 FORMAT(1H ,8X,'AHRT (HOURS)',2X,6(E13.5))
1184 318 FORMAT(1H ,8X,'NHRT (HOURS)',2X,6(E13.5))
1185 3003 FORMAT(1H ,8X,'ANOX FR (%)',2X,7(E13.5))
1186 WRITE(CH,'(I1)')NR
1187 IMFT(10:10)=CH
1188 WRITE(5,399) (KO(I),STQ(I),NTQ(I),Q(1,I),SBI(I),STI(I),
1189 INTI(I),SUI(I),NUI(I),XII(I),NH3I(I),NONI(I),I=1,NL,KH)
1190 399 FORMAT(1H1,10X,'DYNAMIC INPUTS',/,1H ,10X,14('-'),/,
1191 11H ,2X,'TIME',2X,'COD LOAD',2X,'TKN LOAD',4X,'FLOW',7X,'SBI',7X,
1192 2'STI',7X,'NTI',7X,'SUI',7X,'NUI',7X,'XII',6X,'NH3I',6X,'NONI',/,
1193 31H ,2X,4('-'),2X,8('-'),3X,8('-'),4X,4('-'),7X,3('-'),7X,3('-'),
1194 47X,3('-'),7X,3('-'),7X,3('-'),7X,3('-'),6X,4('-'),6X,4('-'),/,
1195 5(1H ,2X,I4,11(E10.4)))
1196 WRITE(5,400)
1197 400 FORMAT(1H1,10X,21('-'),/,1H ,10X,'DAILY DYNAMIC RESULTS',/,
1198 11H ,10X,21('-'),/,
1199 11H0,20X,'TIME',5X,'TANK 1',6X,'TANK 2',
1200 26X,'TANK 3',6X,'TANK 4',6X,'TANK 5',6X,'TANK 6',/,
1201 31H ,20X,4('-'),5X,7('-'),6X,7('-'),6X,7('-'),6X,7('-'),
1202 46X,7('-'),6X,7('-'))
1203 WRITE(5,401)
1204 401 FORMAT(1H0,3X,'XA (MG-VSS/L)')
1205 WRITE(5,IMFT) (KO(I),(XA(J,I),J=1,NR,1),I=1,NL,KH)
1206 WRITE(5,402)
1207 402 FORMAT(1H0,3X,'XS (MG-VSS/L)')
1208 WRITE(5,IMFT) (KO(I),(XS(J,I),J=1,NR,1),I=1,NL,KH)
1209 WRITE(5,403)
1210 403 FORMAT(1H0,3X,'XE (MG-VSS/L)')
1211 WRITE(5,IMFT) (KO(I),(XE(J,I),J=1,NR,1),I=1,NL,KH)
1212 WRITE(5,404)
1213 404 FORMAT(1H0,3X,'XI (MG-VSS/L)')
1214 WRITE(5,IMFT) (KO(I),(XI(J,I),J=1,NR,1),I=1,NL,KH)
1215 WRITE(5,405)

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1216 405 FORMAT(1H0,3X,'XV (MG-VSS/L)')
1217 WRITE(5,IMFT) (KO(I),(XV(J,I),J=1,NR,1),I=1,NL,KH)
1218 WRITE(5,419)
1219 419 FORMAT(1H0,3X,'XN (MG-VSS/L)')
1220 WRITE(5,IMFT) (KO(I),(XN(J,I),J=1,NR,1),I=1,NL,KH)
1221 WRITE(5,407)
1222 407 FORMAT(1H0,3X,'OC (MG-O/L/HR)')
1223 WRITE(5,IMFT) (KO(I),(OC(J,I),J=1,NR,1),I=1,NL,KH)
1224 WRITE(5,408)
1225 408 FORMAT(1H0,3X,'ON (MG-O/L/HR)')
1226 WRITE(5,IMFT) (KO(I),(ON(J,I),J=1,NR,1),I=1,NL,KH)
1227 WRITE(5,409)
1228 409 FORMAT(1H0,3X,'OT (MG-O/L/HR)')
1229 WRITE(5,IMFT) (KO(I),(OT(J,I),J=1,NR,1),I=1,NL,KH)
1230 WRITE(5,4090)
1231 4090 FORMAT(1H0,3X,'AM (MG-N/L/HR)')
1232 WRITE(5,IMFT) (KO(I),(AMON(J,I),J=1,NR,1),I=1,NL,KH)
1233 WRITE(5,417)
1234 417 FORMAT(1H0,3X,'WQ (L/DAY)')
1235 WRITE(5,IMFT) (KO(I),(WQ(J,I),J=1,NR,1),I=1,NL,KH)
1236 WRITE(5,411)
1237 411 FORMAT(1H0,3X,'SBP (MG-COD/L)')
1238 WRITE(5,IMFT) (KO(I),(SBP(J,I),J=1,NR,1),I=1,NL,KH)
1239 WRITE(5,418)
1240 418 FORMAT(1H0,3X,'SBS (MG-COD/L)')
1241 WRITE(5,IMFT) (KO(I),(SBS(J,I),J=1,NR,1),I=1,NL,KH)
1242 WRITE(5,420)
1243 420 FORMAT(1H0,3X,'SU (MG-COD/L)')
1244 WRITE(5,IMFT) (KO(I),(SU(J,I),J=1,NR,1),I=1,NL,KH)
1245 WRITE(5,412)
1246 412 FORMAT(1H0,3X,'ST (MG-COD/L)')
1247 WRITE(5,IMFT) (KO(I),(ST(J,I),J=1,NR,1),I=1,NL,KH)
1248 WRITE(5,413)
1249 413 FORMAT(1H0,3X,'NON (MG-N/L)')
1250 WRITE(5,IMFT) (KO(I),(NON(J,I),J=1,NR,1),I=1,NL,KH)
1251 WRITE(5,421)
1252 421 FORMAT(1H0,3X,'NU (MG-N/L)')
1253 WRITE(5,IMFT) (KO(I),(NU(J,I),J=1,NR,1),I=1,NL,KH)
1254 WRITE(5,415)
1255 415 FORMAT(1H0,3X,'NH3 (MG-N/L)')
1256 WRITE(5,IMFT) (KO(I),(NH3(J,I),J=1,NR,1),I=1,NL,KH)
1257 WRITE(5,414)
1258 414 FORMAT(1H0,3X,'NT (MG-N/L)')
1259 WRITE(5,IMFT) (KO(I),(NT(J,I),J=1,NR,1),I=1,NL,KH)
1260 WRITE(5,422)
1261 422 FORMAT(1H0,3X,'APT (MG-N/L/HR)')
1262 WRITE(5,IMFT) (KO(I),(ANPOT(J,I),J=1,NR,1),I=1,NL,KH)
1263 WRITE(5,416)
1264 416 FORMAT(1H0,3X,'NO3 (MG-N/L)')
1265 WRITE(5,IMFT) (KO(I),(NO3(J,I),J=1,NR,1),I=1,NL,KH)
1266 WRITE(5,461)
1267 461 FORMAT(1H0,3X,'ALK (MG-CACO3/L)')
1268 WRITE(5,IMFT) (KO(I),(ALK(J,I),J=1,NR,1),I=1,NL,KH)
1269 IF(RESULT.NE.0) GO TO 127
1270 126 WRITE(18,522)ECOD,ENT,ENH3,ENO3,EALK
1271 WRITE(18,522)KOM,D,INIT(1),ANOX(1),AROB(1),IFREQ,NDH
1272 522 FORMAT(
1273 WRITE(18,45)(OT(1,I),XV(1,I),ST(1,I),NT(1,I),NH3(1,I),NO3(1,I),
1274 &ALK(1,I),I=1,NDH)
1275 WRITE(18,522) START,END
1276 45 FORMAT(7E13.5)
1277 ENDFILE 18
1278 127 STOP
1279 END

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1  C*****
2      SUBROUTINE UAERO(D,DT,J,FR,FBS,FCS,KVT,KMPT,KSPT,KMST,KSST,
3      1P,NR,YH,BHT,F,KRT,FOE,FN,FOS,FNS,NJ,DDT,UNT,YN,KNT,RTD,BNT,
4      2FACT,LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR,I)
5  C*****
6  C          THIS SUBROUTINE CALCULATES THE DYNAMIC RESPONSE OF A
7  C          REACTOR IN A SERIES UNDER AEROBIC CONDITIONS
8  C*****
9      PARAMETER NE=3
10     PARAMETER NEE=4
11     PARAMETER ND=241
12     PARAMETER NDD=242
13     INTEGER D
14     DOUBLE PRECISION DT,DDT,RTD
15     COMMON/EXTC/SBS(NE,ND),SU(NE,ND),XA(NE,ND),XE(NE,ND),XI(NE,ND),
16     1XV(NE,ND),XS(NE,ND),NU(NE,ND),NON(NE,ND),XN(NE,ND),NH3(NE,ND),
17     2NO3(NE,ND),SBP(NE,ND),ALK(NE,ND)
18     COMMON/BLOCKC/Q(NEE,NDD),V(NE),SBI(ND),SUI(ND),NIT(NE,ND),
19     1NUI(ND),XII(ND),NONI(ND),NH3I(ND),NO3I(ND),SBPR(ND),ALKI(ND),
20     2XAR(ND),XSR(ND),XER(ND),XIR(ND),XNR(ND),SYNTH(NE,ND),QAV(NEE),
21     3ANPOT(NE,ND),AMON(NE,ND)
22     REAL KVT,KMPT,KSPT,KMST,KSST,KRT,KNT
23     REAL NUI,NONI,NH3I,NO3I,NIT
24     REAL NU,NON,NH3,NO3
25     REAL NH3T,NITT,NNIT,NO3T
26  C*****
27     CALL UDUMV(J,I,DUMIF,DUMS,DUMA,DUMB,DUMIR,DUMO,
28     1LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR,
29     1DT,DDT,RTD)
30  C*****
31     K=J-1
32     IF(K.EQ.0) K=1
33     SCER=P*(1.0-F)*BHT*XA(J,I)*DDT
34     SCERN=FN*SCER/P
35     CVNON=KRT*XA(J,I)*NON(J,I)*DDT
36     SBST=SBS(J,I)
37     SBST=SBS(J,I)
38     NH3T=NH3(J,I)
39     NO3T=NO3(J,I)
40     XNT=XN(J,I)
41     SCVO=0.00
42     SCIS=0.00
43     NNIT=0.0
44     ANPOTS=0.0
45     DO 212 IJ=1,NJ,1
46     SCIST=KMST*SBST/(KSPT*XA(J,I)*DDT
47     SCVOT=KMPT*XS(J,I)/(KSPT*XA(J,I)+XS(J,I)*P)*XA(J,I)*DDT
48     SYNTHT=SCVOT*P+SCIST
49     NITT=(UNT/YN)*NH3T/(KNT+NH3T)*XNT*DDT
50     SBST=SBST+DUMIF*FBS*SBI(I)-SCIST
51     1+DUMS*SBS(NR,I)+DUMA*SBS(LARO,I)-DUMO*SBST
52     1+DUMB*SBS(LBRO,I)+DUMIR*SBS(K,I)
53     IF(SBST.LE.0.0) SBST=0.0
54     TNH3T=NH3T+DUMIF*NH3I(I)+CVNON-NITT
55     1+DUMS*NH3(NR,I)+DUMA*NH3(LARO,I)-DUMO*NH3T
56     1+DUMB*NH3(LBRO,I)+DUMIR*NH3(K,I)
57     1+(1.0-FOE)*SCERN-(1.0-FOS)*FN*YH*SYNTHT
58     IF(TNH3T.GT.0.0) GO TO 433
59     NH3T=0.0
60     NO3T=NO3T+DUMIF*NO3I(I)+TNH3T
61     1+DUMS*NO3(NR,I)+DUMA*NO3(LARO,I)-DUMO*NO3T
62     1+DUMB*NO3(LBRO,I)+DUMIR*NO3(K,I)
63     GO TO 444

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64      433 NH3T=TNH3T
65      NO3T=NO3T+DUMIF*NO3I(I)+NITT
66      1+DUMS*NO3(NR,I)+DUMA*NO3(LARO,I)-DUMO*NO3T
67      1+DUMB*NO3(LBRO,I)+DUMIR*NO3(K,I)
68      444 XNT=XNT+YN*NITT-BNT*XNT*DDT
69      1+DUMS*XNR(I)+DUMA*XN(LARO,I)-DUMO*XNT
70      1+DUMB*XN(LBRO,I)+DUMIR*XN(K,I)
71      SCVO=SCVO+SCVOT
72      SCIS=SCIS+SCIST
73      NNIT=NNIT+NITT
74      212 CONTINUE
75      SYNTH(J,I)=SCVO*P+SCIS
76      NIT(J,I)=NNIT
77      ANPOT(J,I)=ANPOTS
78      SBS(J,I+1)=SBST
79      NH3(J,I+1)=NH3T
80      NO3(J,I+1)=NO3T
81      XN(J,I+1)=XNT
82      DUMIF=DUMIF/RDT
83      DUMIR=DUMIR/RDT
84      DUMA=DUMA/RDT
85      DUMB=DUMB/RDT
86      DUMS=DUMS/RDT
87      DUMO=DUMO/RDT
88      BRAK=FR-(XS(J,I)/(XA(J,I)))
89      IF(BRAK.GT.0.00) GO TO 40
90      BRAK=0.00
91      XS(J,I)=FR*(XA(J,I))
92      40  XV(J,I)=XS(J,I)+XA(J,I)+XE(J,I)+XI(J,I)
93      SCIP=DT*KVT*BRAK*SBP(J,I)*XA(J,I)
94      SBP(J,I+1)=SBP(J,I)+DUMIF*(1.0-FBS)*SBI(I)-SCIP
95      1+DUMS*SBP(I)+DUMA*SBP(LARO,I)-DUMO*SBP(J,I)
96      1+DUMB*SBP(LBRO,I)+DUMIR*SBP(K,I)
97      2+(1.0-FCS)*SCER/RDT
98      IF(SBP(J,I+1).LT.0.0) SBP(J,I+1)=0.0
99      XS(J,I+1)=XS(J,I)+SCIP/P-SCVO
100     1+DUMS*XS(I)+DUMA*XS(LARO,I)-DUMO*XS(J,I)
101     1+DUMB*XS(LBRO,I)+DUMIR*XS(K,I)
102     IF(XS(J,I+1).LT.0.0) XS(J,I+1)=0.0
103     SU(J,I+1)=SU(J,I)+DUMIF*SUI(I)
104     1+DUMS*SUI(NR,I)+DUMA*SUI(LARO,I)-DUMO*SUI(J,I)
105     1+DUMB*SUI(LBRO,I)+DUMIR*SUI(K,I)
106     NU(J,I+1)=NU(J,I)+DUMIF*NUI(I)
107     1+DUMS*NU(NR,I)+DUMA*NU(LARO,I)-DUMO*NU(J,I)
108     1+DUMB*NU(LBRO,I)+DUMIR*NU(K,I)
109     XA(J,I+1)=XA(J,I)+YH*SYNTH(J,I)-BHT*XA(J,I)*DT
110     1+DUMS*XAR(I)+DUMA*XA(LARO,I)-DUMO*XA(J,I)
111     1+DUMB*XA(LBRO,I)+DUMIR*XA(K,I)
112     XE(J,I+1)=XE(J,I)+F*BHT*XA(J,I)*DT
113     1+DUMS*XER(I)+DUMA*XE(LARO,I)-DUMO*XE(J,I)
114     1+DUMB*XE(LBRO,I)+DUMIR*XE(K,I)
115     XI(J,I+1)=XI(J,I)+DUMIF*XII(I)
116     1+DUMS*XIR(I)+DUMA*XI(LARO,I)-DUMO*XI(J,I)
117     1+DUMB*XI(LBRO,I)+DUMIR*XI(K,I)
118     NON(J,I+1)=NON(J,I)+DUMIF*NONI(I)-CVNON/RDT
119     1+DUMS*NON(NR,I)+DUMA*NON(LARO,I)-DUMO*NON(J,I)
120     1+DUMB*NON(LBRO,I)+DUMIR*NON(K,I)
121     2+FOE*SCERN/RDT-FOS*FN*YH*SYNTH(J,I)
122     3-FNS*(SCIP/P-SCVO)
123     AMON(J,I)=(CVNON/RDT+(1.0-FOE)*SCERN/RDT)
124     1-(1.0-FOS)*FN*YH*SYNTH(J,I)
125     ALK(J,I+1)=ALK(J,I)+DUMIF*ALKI(I)
126     1+DUMS*ALK(NR,I)+DUMA*ALK(LARO,I)-DUMO*ALK(J,I)
127     1+DUMB*ALK(LBRO,I)+DUMIR*ALK(K,I)

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128      1-7.14*NIT(J,I)+3.57*AMON(J,I)
129      XV(J,I+1)=XS(J,I+1)+XA(J,I+1)+XE(J,I+1)+XI(J,I+1)
130      RETURN
131      END
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1 C*****
2 SUBROUTINE UANOX(D,DT,J,FR,FBS,FCS,KVT,KMPT,KSPT,KMST,KSST,
3 1P,NR,YH,BHT,F,KRT,FOE,FN,FOS,FNS,NJ,DDT,UMT,YN,KNT,RTD,BNT,
4 2FACT,LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QER,I)
5 C*****
6 C THIS SUBROUTINE CALCULATES THE DYNAMIC RESPONSE OF A
7 C REACTOR IN A SERIES UNDER ANOXIC CONDITIONS
8 C*****
9 PARAMETER NE=3
10 PARAMETER NEE=4
11 PARAMETER ND=241
12 PARAMETER NDD=242
13 INTEGER D
14 DOUBLE PRECISION DT,DDT,RTD
15 COMMON/EXTC/SBS(NE,ND),SU(NE,ND),XA(NE,ND),XE(NE,ND),XI(NE,ND),
16 1XV(NE,ND),XS(NE,ND),NU(NE,ND),NON(NE,ND),XN(NE,ND),NH3(NE,ND),
17 2NO3(NE,ND),SBP(NE,ND),ALK(NE,ND)
18 COMMON/BLOCKC/Q(NEE,NDD),V(NE),SBI(ND),SUI(ND),NIT(NE,ND),
19 1NUI(ND),XII(ND),NONI(ND),NH3I(ND),NO3I(ND),SBPR(ND),ALKI(ND),
20 2XAR(ND),XSR(ND),XER(ND),XIR(ND),XNR(ND),SYNTH(NE,ND),QAV(NEE),
21 3ANPOT(NE,ND),AMON(NE,ND)
22 REAL KVT,KMPT,KSPT,KMST,KSST,KRT,KNT
23 REAL NUI,NONI,NH3I,NO3I,NIT
24 REAL NU,NON,NH3,NO3
25 REAL NH3T,NITT,NNIT,NO3T
26 RNCOD=2.85/(1.0-P*YH)
27 C*****
28 CALL UDUMV(J,I,DUMIF,DUMS,DUMA,DUMB,DUMIR,DUMO,
29 1LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR,
30 1DT,DDT,RTD)
31 C*****
32 K=J-1
33 IF(K.EQ.0) K=1
34 SCER=P*(1.0-F)*BHT*XA(J,I)*DDT
35 SCERN=FN*SCER/P
36 CVNON=KRT*FACT*XA(J,I)*NON(J,I)*DDT
37 SCVOD=KMPT*XS(J,I)/(KSPT*XA(J,I)+XS(J,I)*P)*FACT*XA(J,I)*DDT
38 SCVO=0.00
39 SCIS=0.00
40 NNIT=0.00
41 ANPOTS=0.00
42 SBST=SBS(J,I)
43 SBST=SBS(J,I)
44 NH3T=NH3(J,I)
45 NO3T=NO3(J,I)
46 XNT=XN(J,I)
47 DO 213 IJ=1,NJ,1
48 SCIST=KMST*SBST/(KSST+SBST)*FACT*XA(J,I)*DDT
49 SCVOT=KMPT*XS(J,I)/(KSPT*XA(J,I)+XS(J,I)*P)*FACT*XA(J,I)*DDT
50 SYNTHT=SCVOT*P+SCIST
51 DENCAP=SYNTHT/RNCOD
52 DENIT=DENCAP
53 RDEN=SCIST/SYNTHT
54 NITT=0.0
55 ANPOTT=0.00
56 IF((NO3T-DENIT).GT.0.00) GO TO 13
57 DENIT=NO3T
58 SYNTHT=DENIT*RNCOD
59 SCIST=SYNTHT*RDEN
60 SCVOT=SYNTHT*(1.0-RDEN)/P
61 ANPOTT=DENCAP-DENIT
62 13 CONTINUE
63 SCVO=SCVO+SCVOT

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64      SCIS=SCIS+SCIST
65      ANPOTS=ANPOTS+ANPOTT
66      SBST=SBST+DUMIF*FBS*SBI(I)-SCIST
67      1+DUMS*SBS(NR,I)+DUMA*SBS(LARO,I)-DUMO*SBST
68      1+DUMB*SBS(LBRO,I)+DUMIR*SBS(K,I)
69      IF(SBST.LE.0.0) SBST=0.0
70      NITT=0.00
71      TNH3T=NH3T+DUMIF*NH3I(I)+CVNON
72      1+DUMS*NH3(NR,I)+DUMA*NH3(LARO,I)-DUMO*NH3T
73      1+DUMB*NH3(LBRO,I)+DUMIR*NH3(K,I)
74      1+(1.0-FOE)*SCERN-(1.0-FOS)*FN*YH*SYNTHT
75      IF(TNH3T.GT.0.0) GO TO 43
76      NH3T=0.0
77      NO3T=NO3T+DUMIF*NO3I(I)+NITT-DENIT+TNH3T
78      1+DUMS*NO3(NR,I)+DUMA*NO3(LARO,I)-DUMO*NO3T
79      1+DUMB*NO3(LBRO,I)+DUMIR*NO3(K,I)
80      IF(NO3T.LT.0.00) NO3T=0.00
81      GO TO 44
82      43 NH3T=TNH3T
83      NO3T=NO3T+DUMIF*NO3I(I)+NITT-DENIT
84      1+DUMS*NO3(NR,I)+DUMA*NO3(LARO,I)-DUMO*NO3T
85      1+DUMB*NO3(LBRO,I)+DUMIR*NO3(K,I)
86      44 XNT=XNT+YN*NITT-BHT*XNT*DDT
87      1+DUMS*XNR(I)+DUMA*XN(LARO,I)-DUMO*XNT
88      1+DUMB*XN(LBRO,I)+DUMIR*XN(K,I)
89      213 CONTINUE
90      SYNTH(J,I)=SCVO*P+SCIS
91      NIT(J,I)=NNIT
92      ANPOT(J,I)=ANPOTS
93      SBS(J,I+1)=SBST
94      NH3(J,I+1)=NH3T
95      NO3(J,I+1)=NO3T
96      XN(J,I+1)=XNT
97      DUMIF=DUMIF/RDT
98      DUMIR=DUMIR/RDT
99      DUMA=DUMA/RDT
100     DUMB=DUMB/RDT
101     DUMS=DUMS/RDT
102     DUMO=DUMO/RDT
103     BRAK=FR-(XS(J,I)/(XA(J,I)))
104     IF(BRAK.GT.0.00) GO TO 40
105     BRAK=0.00
106     XS(J,I)=FR*(XA(J,I))
107     40 XV(J,I)=XS(J,I)+XA(J,I)+XE(J,I)+XI(J,I)
108     SCIP=DT*KVT*BRAK*SBP(J,I)*XA(J,I)
109     SBP(J,I+1)=SBP(J,I)+DUMIF*(1.0-FBS)*SBI(I)-SCIP
110     1+DUMS*SBPR(I)+DUMA*SBP(LARO,I)-DUMO*SBP(J,I)
111     1+DUMB*SBP(LBRO,I)+DUMIR*SBP(K,I)
112     2+(1.0-FCS)*SCER/RDT
113     IF(SBP(J,I+1).LT.0.0) SBP(J,I+1)=0.0
114     XS(J,I+1)=XS(J,I)+SCIP/P-SCVO
115     1+DUMS*XS(I)+DUMA*XS(LARO,I)-DUMO*XS(J,I)
116     1+DUMB*XS(LBRO,I)+DUMIR*XS(K,I)
117     IF(XS(J,I+1).LT.0.0) XS(J,I+1)=0.0
118     SU(J,I+1)=SU(J,I)+DUMIF*SUI(I)
119     1+DUMS*SU(NR,I)+DUMA*SU(LARO,I)-DUMO*SU(J,I)
120     1+DUMB*SU(LBRO,I)+DUMIR*SU(K,I)
121     NU(J,I+1)=NU(J,I)+DUMIF*NUI(I)
122     1+DUMS*NU(NR,I)+DUMA*NU(LARO,I)-DUMO*NU(J,I)
123     1+DUMB*NU(LBRO,I)+DUMIR*NU(K,I)
124     XA(J,I+1)=XA(J,I)+YH*SYNTH(J,I)-BHT*XA(J,I)*DT
125     1+DUMS*XAR(I)+DUMA*XA(LARO,I)-DUMO*XA(J,I)
126     1+DUMB*XA(LBRO,I)+DUMIR*XA(K,I)
127     XE(J,I+1)=XE(J,I)+F*BHT*XA(J,I)*DT

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128      1+DUMS*XER(I)+DUMA*XE(LARO,I)-DUMO*XE(J,I)
129      1+DUMB*XE(LBRO,I)+DUMIR*XE(K,I)
130      XI(J,I+1)=XI(J,I)+DUMIF*XII(I)
131      1+DUMS*XIR(I)+DUMA*XI(LARO,I)-DUMO*XI(J,I)
132      1+DUMB*XI(LBRO,I)+DUMIR*XI(K,I)
133      NON(J,I+1)=NON(J,I)+DUMIF*NONI(I)-CVNON/RDT
134      1+DUMS*NON(NR,I)+DUMA*NON(LARO,I)-DUMO*NON(J,I)
135      1+DUMB*NON(LBRO,I)+DUMIR*NON(K,I)
136      2+FOE*SCERN/RDT-FOS*FN*YH*SYNTH(J,I)
137      3-FNS*(SCIP/P-SCVO)
138      AMON(J,I)=(CVNON/RDT+(1.0-FOE)*SCERN/RDT)
139      1-(1.0-FOS)*FN*YH*SYNTH(J,I)
140      ALK(J,I+1)=ALK(J,I)+DUMIF*ALKI(I)
141      1+DUMS*ALK(NR,I)+DUMA*ALK(LARO,I)-DUMO*ALK(J,I)
142      1+DUMB*ALK(LBRO,I)+DUMIR*ALK(K,I)
143      1+3.57*(SYNTH(J,I)/RNCOD+AMON(J,I))
144      XV(J,I+1)=XS(J,I+1)+XA(J,I+1)+XE(J,I+1)+XI(J,I+1)
145      RETURN
146      END

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1  C*****
2  SUBROUTINE USETL(NL,J,QSR,I)
3  C*****
4  C          THIS SUBROUTINE CALCULATES THE DYNAMIC RESPONSE OF THE
5  C          SETTLING TANK
6  C*****
7  PARAMETER NE=3
8  PARAMETER NEE=4
9  PARAMETER ND=241
10 PARAMETER NDD=242
11 COMMON/EXTC/SBS(NE,ND),SU(NE,ND),XA(NE,ND),XE(NE,ND),XI(NE,ND),
12 1XV(NE,ND),XS(NE,ND),NU(NE,ND),NON(NE,ND),XN(NE,ND),NH3(NE,ND),
13 2NO3(NE,ND),SBP(NE,ND),ALK(NE,ND)
14 COMMON/BLOCKC/Q(NEE,NDD),V(NE),SBI(ND),SUI(ND),NIT(NE,ND),
15 1NUI(ND),XII(ND),NONI(ND),NH3I(ND),NO3I(ND),SBPR(ND),ALKI(ND),
16 2XAR(ND),XSR(ND),XER(ND),XIR(ND),XNR(ND),SYNTH(NE,ND),QAV(NEE),
17 3ANPOT(NE,ND),AMON(NE,ND)
18 REAL NUI,NONI,NH3I,NO3I,NIT
19 REAL NU,NON,NH3,NO3
20 DUM=(QSR+Q(J+1,I))/QSR
21 XAR(I+1)=XA(J,I+1)*DUM
22 XSR(I+1)=XS(J,I+1)*DUM
23 XER(I+1)=XE(J,I+1)*DUM
24 XIR(I+1)=XI(J,I+1)*DUM
25 SBPR(I+1)=SBP(J,I+1)*DUM
26 XNR(I+1)=XN(J,I+1)*DUM
27 RETURN
28 END

```

```

1  C*****
2      SUBROUTINE UDUMV(J,I,DUMIF,DUMS,DUMA,DUMB,DUMIR,DUMO,
3      1LINI,LSRI,LARI,LARO,LBRI,LBRO,QSR,QAR,QBR,
4      1DT,DDT,DDT)
5  C*****
6  C          THIS SUBROUTINE CALCULATES THE DUMMY VALUES
7  C          FROM THE PROCESS CONFIGURATION INPUT DATA
8  C*****
9      PARAMETER NE=3
10     PARAMETER NEE=4
11     PARAMETER ND=241
12     PARAMETER NDD=242
13     INTEGER D
14     DOUBLE PRECISION DT,DDT,DDT
15     COMMON/EXTC/SBS(NE,ND),SU(NE,ND),XA(NE,ND),XE(NE,ND),XI(NE,ND),
16     1XV(NE,ND),XS(NE,ND),NU(NE,ND),NON(NE,ND),XN(NE,ND),NH3(NE,ND),
17     2NO3(NE,ND),SBP(NE,ND),ALK(NE,ND)
18     COMMON/BLOCKC/Q(NEE,NDD),V(NE),SBI(ND),SUI(ND),NIT(NE,ND),
19     1NUI(ND),XII(ND),NONI(ND),NH3I(ND),NO3I(ND),SEPR(ND),ALKI(ND),
20     2XAR(ND),XSR(ND),XER(ND),XIR(ND),XNR(ND),SYNTH(NE,ND),QAV(NEE),
21     3ANPOT(NE,ND),AMON(NE,ND)
22     DUMIF=0.0
23     IF(J.EQ.LINI) DUMIF=DDT*Q(J,I)/V(J)
24     DUMS=0.0
25     IF(J.EQ.LSRI) DUMS=DDT*QSR/V(J)
26     DUMA=0.0
27     IF(J.EQ.LARI.AND.J.LT.LARO) DUMA=DDT*QAR/V(J)
28     DUMB=0.0
29     IF(J.EQ.LBRI.AND.J.LT.LBRO) DUMB=DDT*QBR/V(J)
30     IF(J.EQ.1) GO TO 11
31     IF(J.LE.LINI.AND.J.GT.LSRI) DUMIR=DDT*QSR/V(J)
32     IF(J.LE.LINI.AND.J.LE.LSRI.AND.J.GT.LBRI.AND.J.LE.LBRO)
33     1DUMIR=DDT*QBR/V(J)
34     IF(J.LE.LINI.AND.J.LE.LSRI.AND.J.GT.LARI.AND.J.LE.LARO)
35     1DUMIR=DDT*QAR/V(J)
36     IF(J.LE.LINI.AND.J.LE.LSRI.AND.J.GT.LARI.AND.J.LE.LARO.
37     1AND.J.GT.LBRI.AND.J.LE.LBRO) DUMIR=DDT*(QAR+QBR)/V(J)
38     IF(J.LE.LINI.AND.J.GT.LSRI.AND.J.GT.LARI.AND.J.LE.LARO)
39     1DUMIR=DDT*(QSR+QAR)/V(J)
40     IF(J.LE.LINI.AND.J.GT.LSRI.AND.J.GT.LBRI.AND.J.LE.LBRO)
41     1DUMIR=DDT*(QSR+QBR)/V(J)
42     IF(J.LE.LINI.AND.J.GT.LSRI.AND.J.GT.LARI.AND.J.LE.LARO.
43     1AND.J.GT.LBRI.AND.J.LE.LBRO) DUMIR=DDT*(QSR+QAR+QBR)/V(J)
44     IF(J.GT.LINI.AND.J.GT.LSRI) DUMIR=DDT*(Q(J,I)+QSR)/V(J)
45     IF(J.GT.LINI.AND.J.LE.LSRI.AND.J.GT.LBRI.AND.J.LE.LBRO)
46     1DUMIR=DDT*(Q(J,I)+QBR)/V(J)
47     IF(J.GT.LINI.AND.J.LE.LSRI.AND.J.GT.LARI.AND.J.LE.LARO)
48     1DUMIR=DDT*(Q(J,I)+QAR)/V(J)
49     IF(J.GT.LINI.AND.J.GT.LSRI.AND.J.GT.LARI.AND.J.LE.LARO)
50     1DUMIR=DDT*(Q(J,I)+QAR+QSR)/V(J)
51     IF(J.GT.LINI.AND.J.GT.LSRI.AND.J.GT.LBRI.AND.J.LE.LBRO)
52     1DUMIR=DDT*(Q(J,I)+QBR+QSR)/V(J)
53     IF(J.GT.LINI.AND.J.GT.LSRI.AND.J.GT.LARI.AND.J.LE.LARO.
54     1AND.J.GT.LBRI.AND.J.LE.LBRO) DUMIR=DDT*(Q(J,I)+QSR+QAR+QBR)/V(J)
55 11 CONTINUE
56     IF(J.EQ.1) DUMIR=0.0
57     DUMO=DUMIF+DUMIR+DUMS+DUMA+DUMB
58     RETURN
59     END

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PROGRAM A.3

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1  C*****
2  C  THIS PROGRAMME CAN BE USED FOR PLOTTING OF BOTH
3  C  SIMULATED AND EXPERIMENTAL DATA OF ALL THE MEASURABLE
4  C  VARIABLES IN AN ACTIVATED SLUDGE PROCESS VIZ.:
5  C  1) OXYGEN UPTAKE RATE (OT IN MG/L/H)
6  C  2) FILTERED COD CONCENTRATION (CCD IN MG COD/L)
7  C  3) VSS CONCENTRATION ( VSS IN MG VSS/L)
8  C  4) FILTERED TKN CONCENTRATION (TKN IN MG TKN/L)
9  C  5) AMMONIA CONCENTRATION (NH3-N IN MG NH3-N/L)
10 C  6) NITRATE CONCENTRATION (NO3-N IN MG NO3-N/L)
11 C  7) ALKALINITY (ALK IN MG CaCO3/L)
12 C  THESE PLOTS ALLOW A GRAPHICAL COMPARISON BETWEEN THE
13 C  EXPERIMENTAL AND SIMULATED PROCESS RESPONSE FOR ALL
14 C  THE MEASURABLE VARIABLES.
15 C
16 C
17 C  THE SIMULATED VALUES OF THE VARIABLES ARE READ FROM A
18 C  DATA FILE GENERATED BY THE DYNAMIC STEADY STATE OR THE
19 C  UNSTEADY STATE PROGRAMS. THE EXPERIMENTAL VALUES ARE
20 C  ENTERED AS DATA.
21 C
22 C
23 C  THE SUBROUTINE NAXIS IS WRITTEN BY PROF H.O.BUHR
24 C  AND IS UTILIZED FOR SPECIFYING THE DIVISION OF THE
25 C  Y- AND X-AXIS. THE OTHER SUBROUTINES CALLED IN THIS
26 C  PROGRAM ARE PART OF THE GENERAL DISPLAY PACKAGE UNIT
27 C  (G.D.P.) AVAILABLE AT THE UNIVERSITY OF CAPE TOWN.
28 C*****
29 C  THE NOMENCLATURE OF THE SIMULATED VARIABLES IS EXPLAINED
30 C  IN THE PROGRAM FOR UNSTEADY STATE(H.UNST).
31 C  FOR EXPERIMENTAL VALUES:
32 C  PREFIX T REFERS TO THE TIME OF EXPERIMENTAL VALUE
33 C  PREFIX P REFERS TO THE EXPERIMENTAL VALUE OF THE VARIABLE
34 C  PREFIX N REFERS TO THE NUMBER OF EXPERIMENTAL DETERMI-
35 C  NATIONS OF THE VARIABLE IN THE SET OF DATA
36 C*****
37 C  NAMES BEGINNING WITH Y REFER TO THE ORDINATES OF THE DIAGRAMS
38 C  YAX =LENGTH OF THE Y-AXIS
39 C  YDOT=LENGTH OF INTERRUPTED LINE INDICATING ANOXIC PERIODS
40 C  Y1,Y2,Y3,Y4,Y5 AND YPLUS INDICATE ORDINATES FOR TEXT
41 C  YSTUK=NUMBER OF SUBDIVISIONS OF THE Y-AXIS
42 C  XSTUK=NUMBER OF SUBDIVISIONS OF THE X-AXIS
43 C  SIZE=SIZE OF EXPERIMENTAL POINTS (IN MM)
44 C  NDEL REFERS TO THE NUMBER OF UNITS PER CM OF ORDINATE
45 C  NFIRST REFERS TO THE STARTING VALUE OF THE VARIABLES
46 C  OEQ REFERS TO EQUIVALENT OXYGEN UPTAKE RATE
47 C  (I.E. DENITRIFICATION RATE*2,86)
48 C  PARAMETER ND=30
49 C  PARAMETER NE=244
50 C  INTEGER D
51 C  REAL NT(NE),NH3(NE),NO3(NE)
52 C  DIMENSION T(NE),OT(NE),XV(NE),ST(NE),ALK(NE),TIME(ND),T1(NE)
53 C  DIMENSION STXV(NE),STST(NE),STNT(NE),STNH3(NE),TOEQ(ND),POEQ(ND)
54 C  DIMENSION TOT(ND),TXV(ND),TCOD(ND),TTKN(ND),TNH3(ND),TNO3(ND),
55 C  &TALK(ND)
56 C  DIMENSION POT(ND),PXV(ND),PCOD(ND),PTKN(ND),PNH3(ND),PNO3(ND),
57 C  &PALK(ND)
58 C  READ IN SIMULATED DATA
59 C  READ(18,35)ECOD,ETKN,ENH3,ENO3,EALK
60 C  WRITE(5,35)ECOD,ETKN,ENH3,ENO3,EALK
61 C  READ(18,35)KOM,D,INIT,ANOX,AROB,IPREQ,NDH
62 C  WRITE(5,35)KOM,D,INIT,ANOX,AROB,IFREQ,NDH
63 C  NFIRST=NDH+1

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128      TOT(I)=(TOT(I)-T(NFIRST))/T(NDEL)
129      POT(I)=(POT(I)-OT(NFIRST))/OT(NDEL)
130      65      CONTINUE
131      DO 56 I=1,K,1
132      TOEQ(I)=(TOEQ(I)-T(NFIRST))/T(NDEL)
133      POEQ(I)=(POEQ(I)-OT(NFIRST))/OT(NDEL)
134      56      CONTINUE
135      DO 75 I=1,NXV,1
136      TXV(I)=(TXV(I)-T(NFIRST))/T(NDEL)
137      PXV(I)=(PXV(I)-XV(NFIRST))/XV(NDEL)
138      75      CONTINUE
139      DO 85 I=1,NCOD,1
140      TCOD(I)=(TCOD(I)-T(NFIRST))/T(NDEL)
141      PCOD(I)=(PCOD(I)-ST(NFIRST))/ST(NDEL)
142      85      CONTINUE
143      DO 95 I=1,NTKN,1
144      TTKN(I)=(TTKN(I)-T(NFIRST))/T(NDEL)
145      PTKN(I)=(PTKN(I)-NT(NFIRST))/NT(NDEL)
146      95      CONTINUE
147      DO 105 I=1,NNH3,1
148      TNH3(I)=(TNH3(I)-T(NFIRST))/T(NDEL)
149      PNH3(I)=(PNH3(I)-NH3(NFIRST))/NH3(NDEL)
150      105     CONTINUE
151      DO 115 I=1,NNO3,1
152      TNO3(I)=(TNO3(I)-T(NFIRST))/T(NDEL)
153      PNO3(I)=(PNO3(I)-NO3(NFIRST))/NO3(NDEL)
154      115     CONTINUE
155      DO 125 I=1,NALK,1
156      TALK(I)=(TALK(I)-T(NFIRST))/T(NDEL)
157      PALK(I)=(PALK(I)-ALK(NFIRST))/ALK(NDEL)
158      125     CONTINUE
159      DO 38 I=2,NDH,1
160      T(I)=(T(I-1)+24.0/D)
161      38      CONTINUE
162      DO 335 I=1,NDH,1
163      TI(I)=(T(I)-T(1))/T(NDEL)
164      STXV(I)=(XV(I)-XV(NFIRST))/XV(NDEL)
165      STST(I)=(ST(I)-ST(NFIRST))/ST(NDEL)
166      STNT(I)=(NT(I)-NT(NFIRST))/NT(NDEL)
167      STNH3(I)=(NH3(I)-NH3(NFIRST))/NH3(NDEL)
168      335     CONTINUE
169      TIME(1)=INIT/T(NDEL)
170      DO 205 I=1,IFREQ,1
171      J=2*I-1
172      TIME(J+1)=TIME(J)+ANOX/T(NDEL)
173      TIME(J+2)=TIME(J+1)+AROB/T(NDEL)
174      205     CONTINUE
175      START=START/T(NDEL)+SIZE
176      END=END/T(NDEL)
177      NFR=IFREQ+1
178      45      FORMAT(7E13.5)
179      35      FORMAT( )
180      X2=XAX+10.0
181      XPAGE=3.0*X2
182      X3=-2.0*X2
183      CALL PLOTS(0,0,0)
184      CALL FACTOR(0.33)
185      CALL NEWPEN(1)
186      CALL OPMES(24,'PLEASE LOAD P1-BK/I4  ')
187      CALL PAGDEF(-5.0,-6.0,XPAGE,52.0)
188      CALL PLOT(0.0,26.0,-3)
189      CALL NAXIS(0.0,0.0,XAX,0,XSTUK,0.0,1,0.25,2)
190      CALL NAXIS(0.0,YAX,XAX,0,XSTUK,0.0,-1,0.25,2)
191      CALL NAXIS(0.0,0.0,YAX,0,YSTUK,90.0,-1,0.25,2)

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192      CALL NAXIS(XAX,0.0,YAX,0,YSTUK,90.0,1,0.25,2)
193      DO 106 I=1,ITYD,1
194      TYD=T(1)+(I-1)*XAX*T(NDEL)/(ITYD-1)
195      IF(TYD.GT.24.0)TYD=TYD-24.0
196      XVAL=(1-1)*XAX/(ITYD-1.0)-0.5
197      IF(XVAL.LT.0.0) XVAL=0.0
198      CALL NUMBER(XVAL,-1.0,SIZE,TYD,0.0,-1)
199      106  CONTINUE
200      DO 101 I=1,IJ,1
201      YOT=(I-1)*YAX*OT(NDEL)/(IJ-1)
202      YVAL=YOT/OT(NDEL)
203      IF(YOT.LT.9.9) GO TO 301
204      IF(YOT.GT.9.9.AND.YOT.LT.99.9)GO TO 302
205      IF(YOT.GT.99.9)GO TO 303
206      301  CALL NUMBER(-0.5,YVAL,SIZE,YOT,0.0,-1)
207      GO TO 101
208      302  CALL NUMBER(-1.0,YVAL,SIZE,YOT,0.0,-1)
209      GO TO 101
210      303  CALL NUMBER(-1.5,YVAL,SIZE,YOT,0.0,-1)
211      101  CONTINUE
212      CALL SYMBOL(3.5,-2.0,SIZE,'TIME(HOURS)',0.0,11)
213      CALL SYMBOL(-2.0,4.0,SIZE,'OX. UPT. RATE(MGO/L/HR)',90.0,23)
214      DO 215 I=1,NFR,1
215      DO 215 J=1,KJ,1
216      Y=J-1.0
217      IF(TIME(I).GT.XAX) GO TO 215
218      CALL SYMBOL(TIME(I),Y,0.5,13,0.0,-1)
219      215  CONTINUE
220      IF(NAMBA.EQ.1) GO TO 256
221      TEXT=START +2.5
222      CALL SYMBOL(START,YFEED,SIZE,22,90.0,-1)
223      CALL SYMBOL(END,YFEED,SIZE,23,90.0,-2)
224      CALL SYMBOL(TEXT,YWRITE,SIZE,'FEEDING PERIOD',0.0,14)
225      256  CONTINUE
226      DO 255 I=1,IFREQ,2
227      BEGIN=TIME(I)+SIZE
228      CALL SYMBOL(BEGIN,YDOT,SIZE,22,90.0,-1)
229      CALL SYMBOL(TIME(I+1),YDOT,SIZE,23,90.0,-2)
230      X=(TIME(I)+TIME(I+1))/2-0.75
231      CALL SYMBOL(X,Y5,SIZE,'ANOX',0.0,4)
232      255  CONTINUE
233      CALL SYMBOL(0.5,YPLUS,1.0,'OX UPT RATE',0.0,11)
234      CALL SYMBOL(1.0,Y2,SIZE,0,0.0,-1)
235      CALL SYMBOL(1.0,Y4,SIZE,1,0.0,-1)
236      CALL SYMBOL(1.5,Y2,SIZE,'= O2 UPT. RATE',0.0,14)
237      CALL SYMBOL(1.5,Y4,SIZE,'= EQ. O2 UPT. RATE',0.0,18)
238      DO 55 I=1,NOT,1
239      CALL SYMBOL(TOT(I),POT(I),SIZE,0,0.0,-1)
240      55  CONTINUE
241      DO 44 I=1,K,1
242      CALL SYMBOL(TOEQ(I),POEQ(I),SIZE,1,0.0,-1)
243      44  CONTINUE
244      CALL LINE(T,OT,NDH,1,0,0)
245      16  FORMAT(1H ,5X,'T=',F13.5,5X,'OT=',F13.5)
246      CALL PLOT (X2,0.0,-3)
247      CALL NAXIS(0.0,0.0,XAX,0,XSTUK,0.0,1,0.25,2)
248      CALL NAXIS(0.0,YAX,XAX,0,XSTUK,0.0,-1,0.25,2)
249      CALL NAXIS(0.0,0.0,YAX,0,YSTUK,90.0,-1,0.25,2)
250      CALL NAXIS(XAX,0.0,YAX,0,YSTUK,90.0,1,0.25,2)
251      DO 107 I=1,ITYD,1
252      TYD=T(1)+(I-1)*XAX*T(NDEL)/(ITYD-1)
253      IF(TYD.GT.24.0)TYD=TYD-24.0
254      XVAL=(I-1)*XAX/(ITYD-1.0)-0.5
255      IF(XVAL.LT.0.0) XVAL=0.0

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256      CALL NUMBER(XVAL,-1.0,SIZE,TYD,0.0,-1)
257      107 CONTINUE
258      IF(NAMBA.EQ.1) GO TO 257
259      CALL SYMBOL(START,YFEED,SIZE,22,90.0,-1)
260      CALL SYMBOL(END,YFEED,SIZE,23,90.0,-2)
261      CALL SYMBOL(TEXT,YWRITE,SIZE,'FEEDING PERIOD',0.0,14)
262      257 CONTINUE
263      DO 102 I=1,IJ,1
264      YST=(I-1)*YAX*ST(NDEL)/(IJ-1)
265      YVAL=YST/ST(NDEL)
266      IF(YST.LT.9.9) GO TO 401
267      IF(YST.GT.9.9.AND.YST.LT.99.9)GO TO 402
268      IF(YST.GT.99.9)GO TO 403
269      401 CALL NUMEER(-0.5,YVAL,SIZE,YST,0.0,-1)
270      GO TO 102
271      402 CALL NUMBER(-1.0,YVAL,SIZE,YST,0.0,-1)
272      GO TO 102
273      403 CALL NUMBER(-1.5,YVAL,SIZE,YST,0.0,-1)
274      102 CONTINUE
275      XAX1=XAX+SIZE
276      DO 111 I=1,3,1
277      YXV=(I-1)*YAX*XV(NDEL)/(IJ-1)
278      YVAL=YXV/XV(NDEL)
279      IF(YXV.LT.9.9) GO TO 501
280      IF(YXV.GT.9.9.AND.YXV.LT.99.9)GO TO 502
281      IF(YXV.GT.99.9)GO TO 503
282      501 CALL NUMBER(XAX1,YVAL,SIZE,YXV,0.0,-1)
283      GO TO 111
284      502 CALL NUMBER(XAX1,YVAL,SIZE,YXV,0.0,-1)
285      GO TO 111
286      503 CALL NUMBER(XAX1,YVAL,SIZE,YXV,0.0,-1)
287      111 CONTINUE
288      CALL SYMBOL(3.5,-2.0,SIZE,'TIME(HOURS)',0.0,11)
289      CALL SYMBOL (-2.0,6.0,SIZE,'COD CONC.(MG/L)',90.0,15)
290      CALL SYMBOL (XVSS,2.0,SIZE,'VSS CONC.(MG/L)',90.0,15)
291      DO 315 I=1,NFR,1
292      DO 315 J=1,KJ,1
293      Y=J-1.0
294      IF(TIME(I).GT.XAX) GO TO 315
295      CALL SYMBOL(TIME(I),Y,0.5,13,0.0,-1)
296      315 CONTINUE
297      DO 355 I=1,IFREQ,2
298      BEGIN=TIME(I)+SIZE
299      CALL SYMBOL(BEGIN,YDOT,SIZE,22,90.0,-1)
300      CALL SYMBOL(TIME(I+1),YDOT,SIZE,23,90.0,-2)
301      X=(TIME(I)+TIME(I+1))/2-0.75
302      CALL SYMBOL(X,Y5,SIZE,'ANOX',0.0,4)
303      355 CONTINUE
304      CALL SYMBOL(0.5,YPLUS,1.0,'COD AND VSS',0.0,11)
305      CALL SYMBOL(1.0,Y4,SIZE,1,0.0,-1)
306      CALL SYMBOL(1.0,Y2,SIZE,2,0.0,-1)
307      CALL SYMBOL(1.5,Y4,SIZE,'= VSS',0.0,5)
308      CALL SYMBOL(1.5,Y2,SIZE,'= COD',0.0,5)
309      CALL SYMBOL(5.5,Y0,SIZE,'EFFLUENT',0.0,8)
310      CALL SYMBOL(5.0,Y1,SIZE,'THEO. EXP.',0.0,10)
311      CALL NUMBER(5.5,Y2,SIZE,ECOD,0.0,-1)
312      CALL NUMBER(8.5,Y2,SIZE,AVCOD,0.0,-1)
313      DO 135 I=1,NXV,1
314      CALL SYMBOL(TXV(I),PXV(I),SIZE,1,0.0,-1)
315      135 CONTINUE
316      DO 145 I=1,NCOD,1
317      CALL SYMBOL(TCOD(I),PCOD(I),SIZE,2,0.0,-1)
318      145 CONTINUE
319      CALL LINE(T,XV,NDH,1,0,0)

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320      CALL LINE(T,ST,NDH,1,0,0)
321      CALL PLOT (X2,0.0,-3)
322      CALL NAXIS(0.0,0.0,XAX,0,XSTUK,0.0,1,0.25,2)
323      CALL NAXIS(0.0,YAX,XAX,0,XSTUK,0.0,-1,0.25,2)
324      CALL NAXIS(0.0,0.0,YAX,0,YSTUK,90.0,-1,0.25,2)
325      CALL NAXIS(XAX,0.0,YAX,0,YSTUK,90.0,1,0.25,2)
326      DO 109 I=1,ITYD,1
327      TYD=T(1)+(I-1)*XAX*T(NDEL)/(ITYD-1)
328      IF(TYD.GT.24.0)TYD=TYD-24.0
329      XVAL=(I-1)*XAX/(ITYD-1.0)-0.5
330      IF(XVAL.LT.0.0) XVAL=0.0
331      CALL NUMBER(XVAL,-1.0,SIZE,TYD,0.0,-1)
332      109  CONTINUE
333      IF(NAMBA.EQ.1) GO TO 258
334      CALL SYMBOL(START,YFEED,SIZE,22,90.0,-1)
335      CALL SYMBOL(END,YFEED,SIZE,23,90.0,-2)
336      CALL SYMBOL(TEXT,YWRITE,SIZE,'FEEDING PERIOD',0.0,14)
337      258  CONTINUE
338      DO 104 I=1,IJ,1
339      YNH3=(I-1)*YAX*NH3(NDEL)/(IJ-1)
340      YVAL=YNH3/NH3(NDEL)
341      IF(YNH3.LT.9.9) GO TO 601
342      IF(YNH3.GT.9.9.AND.YNH3.LT.99.9)GO TO 602
343      IF(YNH3.GT.99.9)GO TO 603
344      601  CALL NUMBER(-0.5,YVAL,SIZE,YNH3,0.0,-1)
345      GO TO 104
346      602  CALL NUMBER(-1.0,YVAL,SIZE,YNH3,0.0,-1)
347      GO TO 104
348      603  CALL NUMBER(-1.5,YVAL,SIZE,YNH3,0.0,-1)
349      104  CONTINUE
350      CALL SYMBOL(3.5,-2.0,SIZE,'TIME(HOURS)',0.0,11)
351      CALL SYMBOL(-2.0,6.0,SIZE,'TKN OR NH3(MGN/L)',90.0,17)
352      DO 225 I=1,NFR,1
353      DO 225 J=1,KJ,1
354      Y=J-1.0
355      IF(TIME(I).GT.XAX) GO TO 225
356      CALL SYMBOL(TIME(I),Y,0.5,13,0.0,-1)
357      225  CONTINUE
358      DO 265 I=1,IFREQ,2
359      BEGIN=TIME(I)+SIZE
360      CALL SYMBOL(BEGIN,YDOT,SIZE,22,90.0,-1)
361      CALL SYMBOL(TIME(I+1),YDOT,SIZE,23,90.0,-2)
362      X=(TIME(I)+TIME(I+1))/2-0.75
363      CALL SYMBOL(X,Y5,SIZE,'ANOX',0.0,4)
364      265  CONTINUE
365      CALL SYMBOL(4.0,YPLUS,1.0,'TKN',0.0,3)
366      CALL SYMBOL(1.0,Y2,SIZE,0,0.0,-1)
367      CALL SYMBOL(1.5,Y2,SIZE,'= TKN',0.0,5)
368      CALL SYMBOL(5.5,Y0,SIZE,'EFFLUENT',0.0,8)
369      CALL SYMBOL(5.0,Y1,SIZE,'THEO. EXP.',0.0,10)
370      CALL NUMBER(5.5,Y2,SIZE,ETKN,0.0,1)
371      CALL NUMBER(8.5,Y2,SIZE,AVTKN,0.0,1)
372      DO 155 I=1,NTKN,1
373      CALL SYMBOL(TTKN(I),PTKN(I),SIZE,0,0.0,-1)
374      155  CONTINUE
375      CALL LINE(T,NT,NDH,1,0,0)
376      CALL PLOT (X3,-26.0,-3)
377      CALL NAXIS(0.0,0.0,XAX,0,XSTUK,0.0,1,0.25,2)
378      CALL NAXIS(0.0,YAX,XAX,0,XSTUK,0.0,-1,0.25,2)
379      CALL NAXIS(0.0,0.0,YAX,0,YSTUK,90.0,-1,0.25,2)
380      CALL NAXIS(XAX,0.0,YAX,0,YSTUK,90.0,1,0.25,2)
381      DO 114 I=1,ITYD,1
382      TYD=T(1)+(I-1)*XAX*T(NDEL)/(ITYD-1)
383      IF(TYD.GT.24.0)TYD=TYD-24.0

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```

384      XVAL=(I-1)*XAX/(ITYD-1.0)-0.5
385      IF(XVAL.LT.0.0) XVAL=0.0
386      CALL NUMBER(XVAL,-1.0,SIZE,TYD,0.0,-1)
387 114    CONTINUE
388      DO 116 I=1,IJ,1
389      YNH3=(I-1)*YAX*NH3(NDEL)/(IJ-1)
390      YVAL=YNH3/NH3(NDEL)
391      IF(YNH3.LT.9.9) GO TO 901
392      IF(YNH3.GT.9.9.AND.YNH3.LT.99.9)GO TO 902
393      IF(YNH3.GT.99.9)GO TO 903
394 901    CALL NUMBER(-0.5,YVAL,SIZE,YNH3,0.0,-1)
395      GO TO 116
396 902    CALL NUMBER(-1.0,YVAL,SIZE,YNH3,0.0,-1)
397      GO TO 116
398 903    CALL NUMBER(-1.5,YVAL,SIZE,YNH3,0.0,-1)
399 116    CONTINUE
400      CALL SYMBOL(3.5,-2.0,SIZE,'TIME(HOURS)',0.0,11)
401      CALL SYMBOL(-2.0,6.0,SIZE,'TKN OR NH3(MGN/L)',90.0,17)
402      DO 226 I=1,NFR,1
403      DO 226 J=1,KJ,1
404      Y=J-1.0
405      IF(TIME(I).GT.XAX) GO TO 226
406      CALL SYMBOL(TIME(I),Y,0.5,13,0.0,-1)
407 226    CONTINUE
408      DO 266 I=1,IFREQ,2
409      BEGIN=TIME(I)+SIZE
410      CALL SYMBOL(BEGIN,YDOT,SIZE,22,90.0,-1)
411      CALL SYMBOL(TIME(I+1),YDOT,SIZE,23,90.0,-2)
412      X=(TIME(I)+TIME(I+1))/2-0.75
413      CALL SYMBOL(X,Y5,SIZE,'ANOX',0.0,4)
414 266    CONTINUE
415      CALL SYMBOL(4.0,YPLUS,1.0,'NH3',0.0,3)
416      CALL SYMBOL(5.5,Y0,SIZE,'EFFLUENT',0.0,8)
417      CALL SYMBOL(5.0,Y1,SIZE,'THEO. EXP.',0.0,10)
418      CALL SYMBOL(1.0,Y2,SIZE,1,0.0,-1)
419      CALL SYMBOL(1.5,Y2,SIZE,'= NH3',0.0,5)
420      CALL NUMBER(5.5,Y2,SIZE,ENH3,0.0,1)
421      CALL NUMBER(8.5,Y2,SIZE,AVNH3,0.0,1)
422      DO 165 I=1,NNH3,1
423      CALL SYMBOL(TNH3(I),PNH3(I),SIZE,1,0.0,-1)
424 165    CONTINUE
425      IF(NAMBA.EQ.1) GO TO 259
426      CALL SYMBOL(START,YFEED,SIZE,22,90.0,-1)
427      CALL SYMBOL(END,YFEED,SIZE,23,90.0,-2)
428      CALL SYMBOL(TEXT,YWRITE,SIZE,'FEEDING PERIOD',0.0,14)
429 259    CONTINUE
430      CALL LINE(T,NH3,NDH,1,0,0)
431      CALL PLOT (X2,0.0,-3)
432      CALL NAXIS(0.0,0.0,XAX,0,XSTUK,0.0,1,0.25,2)
433      CALL NAXIS(0.0,YAX,XAX,0,XSTUK,0.0,-1,0.25,2)
434      CALL NAXIS(0.0,0.0,YAX,0,YSTUK,90.0,-1,0.25,2)
435      CALL NAXIS(XAX,0.0,YAX,0,YSTUK,90.0,1,0.25,2)
436      DO 112 I=1,ITYD,1
437      TYD=T(1)+(I-1)*XAX*T(NDEL)/(ITYD-1)
438      IF(TYD.GT.24.0)TYD=TYD-24.0
439      XVAL=(I-1)*XAX/(ITYD-1.0)-0.5
440      IF(XVAL.LT.0.0) XVAL=0.0
441      CALL NUMBER(XVAL,-1.0,SIZE,TYD,0.0,-1)
442 112    CONTINUE
443      DO 113 I=1,IJ,1
444      YNO3=(I-1)*YAX*NO3(NDEL)/(IJ-1)
445      YVAL=YNO3/NO3(NDEL)
446      IF(YNO3.LT.9.9) GO TO 801
447      IF(YNO3.GT.9.9.AND.YNO3.LT.99.9)GO TO 802

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448      IF(YNO3.GT.99.9)GO TO 803
449      801  CALL NUMBER(-0.5,YVAL,SIZE,YNO3,0.0,-1)
450      GO TO 113
451      802  CALL NUMBER(-1.0,YVAL,SIZE,YNO3,0.0,-1)
452      GO TO 113
453      803  CALL NUMBER(-1.5,YVAL,SIZE,YNO3,0.0,-1)
454      113  CONTINUE
455      CALL SYMBOL(3.5,-2.0,SIZE,'TIME(HOURS)',0.0,11)
456      CALL SYMBOL(-1.5,6.5,SIZE,'NO3(MGN/L)',90.0,10)
457      DO 425 I=1,NFR,1
458      DO 425 J=1,KJ,1
459      Y=J-1.0
460      IF(TIME(I).GT.XAX) GO TO 425
461      CALL SYMBOL(TIME(I),Y,0.5,13,0.0,-1)
462      425  CONTINUE
463      DO 465 I=1,IFREQ,2
464      BEGIN=TIME(I)+SIZE
465      CALL SYMBOL(BEGIN,YDOT,SIZE,22,90.0,-1)
466      CALL SYMBOL(TIME(I+1),YDOT,SIZE,23,90.0,-2)
467      X=(TIME(I)+TIME(I+1))/2-0.75
468      CALL SYMBOL(X,Y5,SIZE,'ANOX',0.0,4)
469      465  CONTINUE
470      CALL SYMBOL(1.5,YPLUS,1.0,'NITRATE',0.0,7)
471      CALL SYMBOL(1.0,Y2,SIZE,0,0.0,-1)
472      CALL SYMBOL(1.5,Y2,SIZE,'= NO3',0.0,5)
473      CALL SYMBOL(5.5,Y0,SIZE,'EFFLUENT',0.0,8)
474      CALL SYMBOL(5.0,Y1,SIZE,'THEO. EXP.',0.0,10)
475      CALL NUMBER(5.5,Y2,SIZE,ENO3,0.0,1)
476      CALL NUMBER(8.5,Y2,SIZE,AVNO3,0.0,1)
477      DO 565 I=1,NNO3,1
478      CALL SYMBOL(TNO3(I),PNO3(I),SIZE,0,0.0,-1)
479      565  CONTINUE
480      IF(NAMBA.EQ.1) GO TO 261
481      CALL SYMBOL(START,YFEED,SIZE,22,90.0,-1)
482      CALL SYMBOL(END,YFEED,SIZE,23,90.0,-2)
483      CALL SYMBOL(TEXT,YWRITE,SIZE,'FEEDING PERIOD',0.0,14)
484      261  CONTINUE
485      CALL LINE(T,NO3,NDH,1,0,0)
486      CALL PLOT (X2,0.0,-3)
487      CALL NAXIS(0.0,0.0,XAX,0,XSTUK,0.0,1,0.25,2)
488      CALL NAXIS(0.0,YAX,XAX,0,XSTUK,0.0,-1,0.25,2)
489      CALL NAXIS(0.0,0.0,YAX,0,YSTUK,90.0,-1,0.25,2)
490      CALL NAXIS(XAX,0.0,YAX,0,YSTUK,90.0,1,0.25,2)
491      DO 108 I=1,ITYD,1
492      TYD=T(1)+(I-1)*XAX*T(NDEL)/(ITYD-1)
493      IF(TYD.GT.24.0)TYD=TYD-24.0
494      XVAL=(I-1)*XAX/(ITYD-1.0)-0.5
495      IF(XVAL.LT.0.0) XVAL=0.0
496      CALL NUMBER(XVAL,-1.0,SIZE,TYD,0.0,-1)
497      108  CONTINUE
498      DO 103 I=1,IJ,1
499      YALK=(I-1)*YAX*ALK(NDEL)/(IJ-1)
500      YVAL=YALK/ALK(NDEL)
501      IF(YALK.LT.9.9) GO TO 701
502      IF(YALK.GT.9.9.AND.YALK.LT.99.9)GO TO 702
503      IF(YALK.GT.99.9)GO TO 703
504      701  CALL NUMBER(-0.5,YVAL,SIZE,YALK,0.0,-1)
505      GO TO 103
506      702  CALL NUMBER(-1.0,YVAL,SIZE,YALK,0.0,-1)
507      GO TO 103
508      703  CALL NUMBER(-1.5,YVAL,SIZE,YALK,0.0,-1)
509      103  CONTINUE
510      CALL SYMBOL(3.5,-2.0,SIZE,'TIME(HOURS)',0.0,11)
511      CALL SYMBOL(-2.0,6.0,SIZE,'ALK(PPM CACO3)',90.0,14)

```

```

512      DO 235 I=1,NFR,1
513      DO 235 J=1,KJ,1
514      Y=J-1.0
515      IF(TIME(I).GT.XAX) GO TO 235
516      CALL SYMBOL(TIME(I),Y,0.5,13,0.0,-1)
517 235   CONTINUE
518      DO 245 I=1,IFREQ,2
519      BEGIN=TIME(I)+SIZE
520      CALL SYMBOL(BEGIN,YDOT,SIZE,22,90.0,-1)
521      CALL SYMBOL(TIME(I+1),YDOT,SIZE,23,90.0,-2)
522      X=(TIME(I)+TIME(I+1))/2-0.75
523      CALL SYMBOL(X,Y5,SIZE,'ANOX',0.0,4)
524 245   CONTINUE
525      CALL SYMBOL(1.5,YPLUS,1.0,'ALKALINITY',0.0,10)
526      CALL SYMBOL(1.5,Y2,SIZE,'= ALK',0.0,5)
527      CALL SYMBOL(1.0,Y2,SIZE,0,0.0,-1)
528      CALL SYMBOL(5.5,Y0,SIZE,'EFFLUENT',0.0,8)
529      CALL SYMBOL(5.0,Y1,SIZE,'THEO. EXP.',0.0,10)
530      CALL NUMBER(5.5,Y2,SIZE,EALK,0.0,-1)
531      CALL NUMBER(8.5,Y2,SIZE,AVALK,0.0,-1)
532      DO 185 I=1,NALK,1
533      CALL SYMBOL(TALK(I),PALK(I),SIZE,0,0.0,-1)
534 185   CONTINUE
535      IF(NAMBA.EQ.1) GO TO 262
536      CALL SYMBOL(START,YFEED,SIZE,22,90.0,-1)
537      CALL SYMBOL(END,YFEED,SIZE,23,90.0,-2)
538      CALL SYMBOL(TEXT,YWRITE,SIZE,'FEEDING PERIOD',0.0,14)
539 262   CONTINUE
540      CALL LINE(T,ALK,NDH,1,0,0)
541      CALL PLOT(0.0,0.0,999)
542      STOP
543      END

```



```

1      SUBROUTINE NAXIS(XST,YST,AXLEN,LOG,ANUMBR,ANG,LR,TICK,LINE)
2      C.... ROUTINE FOR DRAWING AXIS WITHOUT ANNOTATION
3      C.... (EITHER LOG OR LINEAR)
4      C....
5      C.... COMPLIMENTS OF YOUR FRIENDLY CHEM. ENG. DEPARTMENT.
6      C.... XST,YST = STARTING POSITION
7      C.... AXLEN = LENGTH OF AXIS TO BE PLOTTED, CM
8      C.... (AXLEN MAY BE NEGATIVE)
9      C.... LOG = STARTING POSN FOR LOG SCALE, USUALLY 1,
10     C.... OR = 0 FOR LINEAR SCALE
11     C.... ANUMBR = NO. OF LOG CYCLES OR NO. OF TICK INTERVALS
12     C.... ALONG AXLEN.
13     C.... ANG = 0. OR 90. FOR HORIZ. OR VERTICAL AXIS (ONLY)
14     C.... LR = LOCATION OF TICK MARKS RELATIVE TO AXIS LINE:
15     C.... 1 FOR CCW, -1 FOR CLOCKWISE.
16     C.... TICK = SIZE OF TICK MARK, CM
17     C.... LINE = 2 FOR SOLID, 3 FOR BLANK AXIS LINE:
18     C.... I.E., 3 WILL GIVE TICK MARKS ONLY.
19     AXIS=0.
20     DX1=0.
21     DDX1=0.
22     DX2=0.
23     DY1=0.
24     DDY1=0.
25     DY2=0.
26     LPOSN=LOG
27     IF(AXLEN.NE.0.) GO TO 11
28     WRITE(NPR,15)
29     15 FORMAT(' WARNING - AXIS LENGTH IN NAXIS ROUTINE IS ZERO.')
30     RETURN
31     11 DELTA=AXLEN/ANUMBR
32     IF(ANG.LT.10.) GO TO 1
33     DX1=TICK
34     IF(LR.GT.0) DX1=-DX1
35     DDX1=DX1
36     GO TO 2
37     1 DY1=TICK
38     IF(LR.LT.0) DY1=-DY1
39     DDY1=DY1
40     C.... START AXIS
41     2 CALL PLOT(XST,YST,3)
42     X=XST
43     Y=YST
44     3 IF(LOG.EQ.0) GO TO 4
45     C.... SET UP LOG SCALE
46     DELTA=(ALOG10(LPOSN+1)-ALOG10(LPOSN))*AXLEN/ANUMBR
47     LPOSN=LPOSN+1
48     IF(LPOSN.EQ.10) LPOSN=1
49     DX1=DDX1
50     DY1=DDY1
51     IF(LPOSN.NE.2) GO TO 4
52     IF(TICK.GT.(.1*AXLEN)) GO TO 4
53     DX1=2*DX1
54     DY1=2*DY1
55     4 IF(ANG.LT.10.)DX2=DELTA
56     IF(ANG.GT.10.) DY2=DELTA
57     AXIS=AXIS+ABS(DELTA)
58     CALL PLOT(X+DX1,Y+DY1,2)
59     CALL PLOT(X,Y,3)
60     IF(AXIS.GT.ABS(AXLEN)) GO TO 5
61     X=X+DX2
62     Y=Y+DY2
63     CALL PLOT(X,Y,LINE)

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```
64      GO TO 3
65      5  IF(ANG.LT.10.) X=XST+AXLEN
66      IF(ANG.GT.10.) Y=YST+AXLEN
67      CALL PLOT(X,Y,LINE)
68      RETURN
69      END
```

APPENDIX B

MASS BALANCE OF A PREDENITRIFICATION SYSTEM

- B.1 General considerations of nitrogen and COD mass balances
- B.2 The COD equivalent content of the sludge
- B.3 Nitrogen mass balance formulation
- B.4 COD mass balance formulation
- B.5 Calculation of nitrogen and COD mass balances on the pre-denitrification system given in Fig. 3.1

B.1 General considerations of nitrogen and COD mass balances

It is possible to perform nitrogen and COD mass balances on the series system in Fig. 3.1 operated under constant flow and load conditions.

The nitrogen mass balance can be used to check on the mixed liquor and underflow recycle pump ratings respectively, subject to inaccuracies related to (1) nitrate, TKN and VSS measurement and (2) possible denitrification in the settler that is unaccounted for. The formulation of nitrogen balance is given in Section B.3.

A prerequisite to a satisfactory COD mass balance is an accurate determination of both the system nitrification and the system denitrification. In addition, errors related to the oxygen uptake rate test will ultimately manifest themselves in the COD mass balance. The formulation of COD balance is given in Section B.4.

In order to dampen any possible irregularities, the average values of the measured parameters for each batch of sewage disregarding outliers were used. The following parameters were measured:

CODIN = Influent total COD concentration $[\text{mg COD} \cdot \ell^{-1}]$
CODOUT = Effluent total COD concentration $[\text{mg COD} \cdot \ell^{-1}]$
TKNIN = Influent total TKN concentration $[\text{mg N} \cdot \ell^{-1}]$
TKNOUT = Effluent total TKN concentration $[\text{mg N} \cdot \ell^{-1}]$

VSS = Volatile suspended solids concentration [$\text{mg VSS} \cdot \ell^{-1}$]
 P = COD equivalent content of the sludge [$\text{mg COD} \cdot \text{mg VSS}^{-1}$]
 a = Mixed liquor recycle ratio (see Fig. 3.1)
 s = Underflow recycle ratio (see Fig. 3.1)
 Q = Volumetric influent flow rate [$\ell \cdot \text{d}^{-1}$]
 q = Volumetric wastage flow rate [$\ell \cdot \text{d}^{-1}$]
 OUR₁ = Oxygen uptake rate in A1 (see Fig. 3.1) [$\text{mg O}_2 \cdot \ell^{-1} \cdot \text{h}^{-1}$]
 OUR₂ = Oxygen uptake rate in A2 (see Fig. 3.1) [$\text{mg O}_2 \cdot \ell^{-1} \cdot \text{h}^{-1}$]
 NO₃AN = Anoxic reactor nitrate concentration [$\text{mg NO}_3\text{-N} \cdot \ell^{-1}$]
 NO₃A1 = Nitrate concentration in A1 (see Fig. 3.1) [$\text{mg NO}_3\text{-N} \cdot \ell^{-1}$]
 NO₃A2 = Nitrate concentration in A2 (see Fig. 3.1) [$\text{mg NO}_3\text{-N} \cdot \ell^{-1}$]
 NO₃EFF = Effluent nitrate concentration [$\text{mg NO}_3\text{-N} \cdot \ell^{-1}$]
 OXFEED = Dissolved oxygen concentration in the feed [$\text{mg O}_2 \cdot \ell^{-1}$]

B.2 The COD equivalent content of the sludge

B.2a Mean value for the COD equivalent content of the sludge

The COD equivalent content of the sludge, P, has been measured regularly by performing the COD test on a known amount of sludge and proved to be $1,54 \pm 0,01$ [$\text{mg COD} \cdot \text{mg VSS}^{-1}$] at 14°C , pH = 7,2 and $R_s = 15[\text{d}]$ (see Fig. 2.10). Hence the mean value of the COD content of the sludge, $P = 1,54$ [$\text{mg COD} \cdot \text{mg VSS}^{-1}$], has been assumed in all the calculations and simulations involving a nitrification-denitrification system at 14°C , pH = 7,2 and $R_s = 15[\text{d}]$.

B.2b Variations in P for different batches of the same sewage source

Experimentally measured values of the COD/VSS ratios at 14°C , pH = 7,2 and $R_s = 15[\text{d}]$ are plotted in Fig. 2.11 for each batch of sewage, and the mean COD/VSS ratio for each batch, disregarding any outliers is indicated. From Fig. 2.11 it seems that the VSS/COD ratio varies for different batches of the same sewage source (Zeekoevlei outfall). This can be explained if it is hypothesized that the COD/VSS ratio *inter alia* depends on the sewage characteristics; variation in characteristics of different batches of the same sewage source may be explained if it is accepted that the industrial fraction in some batches varied.

B.3 Nitrogen mass balance formulation

B.3a Nitrate mass balance on the anoxic reactor

Nitrate mass balance can be performed on the anoxic reactor in Fig. 3.1, provided nitrate is always present in the anoxic reactor. Assuming there is no nitrate in the influent waste flow, the system nitrogen removal is given by the following nitrate mass balance:

$$\text{DENIT} = Q \cdot [a \cdot \text{NO}_3\text{A2} + s \cdot (\text{NO}_3\text{EFF} + \text{NO}_3\text{A2}) / 2 - (a + s + 1) \cdot \text{NO}_3\text{AN}] [\text{mg NO}_3\text{-N} \cdot \text{d}^{-1}]$$

where

$$(\text{NO}_3\text{EFF} + \text{NO}_3\text{A2}) / 2 = \text{estimated nitrate concentration in the underflow recycle } [\text{mg NO}_3 \cdot \text{l}^{-1}]$$

B.3b Heterotrophic nitrogenous nutrient requirement

Heterotrophic nitrogenous nutrient requirement is based on the measured VSS value and is given by

$$\text{NSL} = f_n \cdot X_v \cdot q [\text{mg N} \cdot \text{d}^{-1}]$$

where

$$\text{NSL} = \text{Nitrogen incorporated in the sludge } [\text{mg N} \cdot \text{d}^{-1}]$$

$$f_n = \text{mass fraction of nitrogen in the organic sludge} = 0,1 [\text{mg N} \cdot \text{mg VSS}^{-1}]$$

$$X_v = \text{Volatile suspended solids concentration } [\text{mg VSS} \cdot \text{l}^{-1}]$$

B.3c System nitrogen balance

The system nitrogen balance is given by:

$$Q \cdot \text{TKNIN} = Q \cdot \text{TKNOUT} + \text{NSL} + Q \cdot \text{NO}_3\text{EFF} + \text{DENIT}.$$

Hence the nitrogen mass balance (recovery) is given by

$$\text{NBAL} = \text{NRECOV} = (Q \cdot \text{TKNOUT} + \text{NSL} + Q \cdot \text{NO}_3\text{EFF} + \text{DENIT}) / (Q \cdot \text{TKNIN})$$

B.4 COD mass balance formulation

B.4a COD utilization due to denitrification

Accepting that denitrification of 1 mg $\text{NO}_3\text{-N}$ is equivalent to the utilization of 2,86 mg COD [Section 2.3.1], the COD utilization due to denitrification, CODDEN, is given by

$$\text{CODDEN} = 2,86 [Q \cdot \text{TKNIN} - (Q \cdot \text{TKNOUT} + Q \cdot \text{NO}_3\text{EFF} + \text{NSL})] [\text{mg COD} \cdot \text{d}^{-1}]$$

B.4b Heterotrophic carbonaceous oxygen requirements

Accepting that nitrification of 1 mg N requires 4,57 mg O [Section 2.1.3], the heterotrophic carbonaceous oxygen requirements

may be calculated by subtracting from the total oxygen requirements the amount of oxygen that is needed for nitrification i.e.

$$\text{COR} = [\text{OUR}_1 \cdot 24 \cdot V_{A1} + \text{OUR}_2 \cdot 24 \cdot V_{A2}] - [Q \cdot \text{TKNIN} - (Q \cdot \text{TKNOUT} + \text{NSL})] \cdot 4,57 [\text{mg O.d}^{-1}]$$

where

V_{A1} = Volume of A1 [ℓ]

V_{A2} = Volume of A2 [ℓ]

COR = Carbonaceous oxygen requirements [mg O.d⁻¹]

B.4c COD removal by sludge wastage

The COD removed with the wasted sludge is given by

$$\text{CODSL} = P \cdot q \cdot X_v [\text{mg COD.d}^{-1}]$$

B.4d Dissolved oxygen effects of recycle streams

The mixed liquor and the underflow recycle stream, respectively, introduce dissolved oxygen into the anoxic reactor. The dissolved oxygen concentration in the aerobic reactor was maintained at about 2 [mg O.ℓ⁻¹] in order to ensure efficient nitrification. This means that the mixed liquor 'a' recycle stream will have a dissolved oxygen concentration of about 2 [mg O.ℓ⁻¹]. Because the dissolved oxygen in the settler is lower than that in the aerobic reactor, an approximate dissolved oxygen concentration of 1,5 [mg O.ℓ⁻¹] has been assumed for each of the recycle streams entering the anoxic reactor. Hence an estimation of the total dissolved oxygen returning with the recycle streams to the anoxic reactor is given by:

$$\text{RETOX} = Q \cdot (a+s) \cdot 1,5 [\text{mg COD.d}^{-1}]$$

B.4e Dissolved oxygen effects of feed

The feed is an additional source of dissolved oxygen, considering that it is very well aerated prior to its introduction into the anoxic reactor and is thereafter kept in good contact with oxygen in the air due to the continuous stirring. (The influent is stirred in order to affect an homogenous feed). The amount of oxygen introduced by the feed is given by $Q \cdot \text{OXFEED}$ where OXFEED is the dissolved oxygen concentration in the feed as [mg O.ℓ⁻¹].

B.4f Errors inherent in the oxygen uptake rate test

The oxygen uptake rate test was done by raising the dissolved oxygen concentration to a high value ($6 \text{ [mg O}_2\text{ l}^{-1}\text{]}$), stopping aeration and thereafter monitoring the rate of oxygen uptake by the organisms. Inherent in the oxygen uptake rate test are two effects that tend to act in a compensatory fashion with each other:

- (1) An error made in oxygen uptake rate measurement is proportional to the dissolved oxygen concentration gradient (i.e. the difference between the reactor dissolved oxygen concentration and the influent dissolved oxygen concentration to the reactor).
- (2) An error with the opposite effect is the continuous penetration and dissolution of atmospheric oxygen into the mixed liquor during the oxygen uptake rate test.

In Appendix C.1 a relationship allowing for these two effects is developed and in Appendix C.2 it becomes apparent that for the system depicted in Fig. 3.1 these two effects are of the same order of magnitude and opposed to each other, hence they are ultimately ignored in the COD mass balance.

B.4g System COD balance

In view of Section B.4 (a to f) the COD system mass balance is given by

$$Q.CODIN = CODDEN + COR + CODSL + Q.CODOUT + RETOX + Q.OXFEED \text{ [mg COD.d}^{-1}\text{]}$$

where

$$CODIN = \text{Influent total COD concentration [mg COD.l}^{-1}\text{]}$$

$$CODOUT = \text{Effluent total COD concentration [mg COD.l}^{-1}\text{]}$$

Hence the COD mass balance (recovery) is given by

$$CODREC = (CODDEN + COR + CODSL + Q.CODOUT + RETOX + Q.OXFEED) / (Q.CODIN).$$

B.5 Calculation of mass balance on the predenitrification system given by Fig. 3.1

In view of Appendix B (1 to 4), a Fortran program (program B.1) has been written to calculate nitrogen and COD mass balances as follows:

PROGRAM B.1

```

1      C      THIS PROGRAM CALCULATES NITROGEN AND COD MASS BALANCES ON
2      C      THE PREDENITRIFICATION SYSTEM GIVEN IN FIGURE 3.1
3      C
4      C      *****
5      C      THE MEANING OF THE INPUT PARAMETERS IS AS FOLLOWS:
6      C
7      C      CODIN=INFLUENT COD CONCENTRATION
8      C      CODOUT=EFFLUENT COD CONCENTRATION
9      C      TKNIN=INFLUENT TKN CONCENTRATION
10     C      TKNOUT=EFFLUENT TKN CONCENTRATION
11     C      VSS=VOLATILE SUSPENDED SOLIDS CONCENTRATION
12     C      P=COD/VSS RATIO (THE COD CONTENT OF THE SLUDGE)
13     C      A=MIXED LIQUOR RECYCLE RATIO
14     C      S=UNDERFLOW RECYCLE RATIO
15     C      Q=INFLUENT FLOW RATE
16     C      QQ=WASTAGE FLOW RATE
17     C      OUR1=OXYGEN UPTAKE RATE IN THE FIRST AEROBIC REACTOR - A1
18     C      OUR2=OXYGEN UPTAKE RATE IN THE SECOND AEROBIC REACTOR - A2
19     C      NO3AN=NITRATE CONCENTRATION IN THE ANOXIC REACTOR - AN
20     C      NO3A2=NITRATE CONCENTRATION IN A2
21     C      NO3EFF=EFFLUENT NITRATE CONCENTRATION
22     C      OXFEED=DISSOLVED OXYGEN CONCENTRATION IN THE INFLUENT FEED
23     C
24     C      *****
25     C
26     C      THE PROGRAM OUTPUT VARIABLES ARE:
27     C
28     C      DATE=DAY AT WHICH MASS BALANCE WAS PERFORMED
29     C      MONTH=MONTH AT WHICH MASS BALANCE WAS PERFORMED
30     C      CODIN=INFLUENT COD CONCENTRATION
31     C      CODOUT=EFFLUENT COD CONCENTRATION
32     C      TKNIN=INFLUENT TKN CONCENTRATION
33     C      TKNOUT=EFFLUENT TKN CONCENTRATION
34     C      OUR1=OXYGEN UPTAKE RATE IN THE FIRST AEROBIC REACTOR - A1
35     C      OUR2=OXYGEN UPTAKE RATE IN THE SECOND AEROBIC REACTOR - A2
36     C      NO3AN=NITRATE CONCENTRATION IN THE ANOXIC REACTOR - AN
37     C      NO3A2=NITRATE CONCENTRATION IN THE SECOND AEROBIC REACTOR
38     C      NO3EFF=EFFLUENT NITRATE CONCENTRATION
39     C      DNIT=NITROGEN REMOVED BY THE SYSTEM
40     C      NRECOV=NITROGEN RECOVERY (NITROGEN MASS BALANCE)
41     C      CODREC=COD RECOVERY (COD MASS BALANCE)
42     C      *****
43     C      REAL NO3AN,NO3A2,NO3EFF,NSL,NRECOV
44     C      INTEGER DATE
45     C      READ 104,N
46     C      104 FORMAT(I2)
47     C      PRINT 105
48     C      105 FORMAT(1H ,4X,'DATE',4X,'CODIN',4X,'CODOUT',4X,'TKNIN',4X,'TKNOUT'
49     C      %,4X,'OUR1',4X,'OUR2',4X,'NO3AN',4X,'NO3A2',4X,'NO3EFF',4X,'NRECOV'
50     C      %,4X,'CODBAL',4X,'DNIT',/,1H ,4X,4(' '),4X,5(' '),4X,6(' '),4X,5(' ')
51     C      %'),4X,6(' '),4X,4(' '),4X,4(' '),4X,5(' '),4X,5(' '),4X,6(' '),4X,
52     C      %6(' '),4X,6(' '),4X,4(' '))
53     C      DO 10 I=1,N
54     C      READ101,CODIN,CODOUT,TKNIN,TKNOUT,VSS,P,A,S,Q,QQ,OUR1,OUR2,NO3AN
55     C      READ101,NO3A2,NO3EFF,OXFEED
56     C      READ 103,DATE,MONTH
57     C      103 FORMAT(I2,I2)
58     C      101 FORMAT(
59     C      TNIN=Q*TKNIN
60     C      NSL=0.1*VSS*QQ
61     C      TNOUT=Q*TKNOUT
62     C      DENIT=Q*(A*NO3A2+S*(NO3EFF+NO3A2)/2-(A+S+1.0)*NO3AN)
63     C      TNO3EF=NO3EFF*Q

```



```

64      NRECOV=100.*(TNOUT+TNO3EF+DENIT+NSL)/TNIN
65      RDENIT=TNIN-(TNOUT+TNO3EF+NSL)
66      TCODIN=CODIN*Q
67      CODDEN=RDENIT*2.86
68      CODOUR=OUR1*24.*2.5+OUR2*24.*4.5
69      TOTNIT=(TNO3EF+RDENIT)*4.57
70      CODSL=P*QQ*VSS
71      TCODEF=CODOUT*Q
72      RETOX=Q*(A+S)*1.5
73      CODREC=(CODDEN+CODOUR-TOTNIT+CODSL+TCODEF+RETOX+Q*OXFEED)
74      %*100./TCODIN
75      DNIT=DENIT/Q
76      10 PRINT 100,DATE,MONTH,CODIN,CODOUT,TKNIN,TKNOUT,OUR1,OUR2,NO3AN,NO3
77      %A2,NO3EFF,NRECOV,CODREC,DNIT
78      100 FORMAT(1H ,4X,I2,I2,4X,F5.1,4X,F6.1,4X,F5.1,4X,F6.1,4X,F4.1,4X,F4.
79      %1,4X,F5.1,4X,F5.1,4X,F6.1,4X,F6.2,4X,F6.2,4X,F4.1)
80      END

```

The system depicted in Fig. 3.1 was run under constant flow and load conditions until satisfactory* nitrogen *and* COD mass balances were obtained using *average values* of the measured parameters *for each batch* of sewage. A summary of program B.1 output for all sewage batches is given in Table 3.1 [see Section 3.2.1].

*Satisfactory nitrogen and COD mass balances are defined as above 95% and 90% recovery respectively.

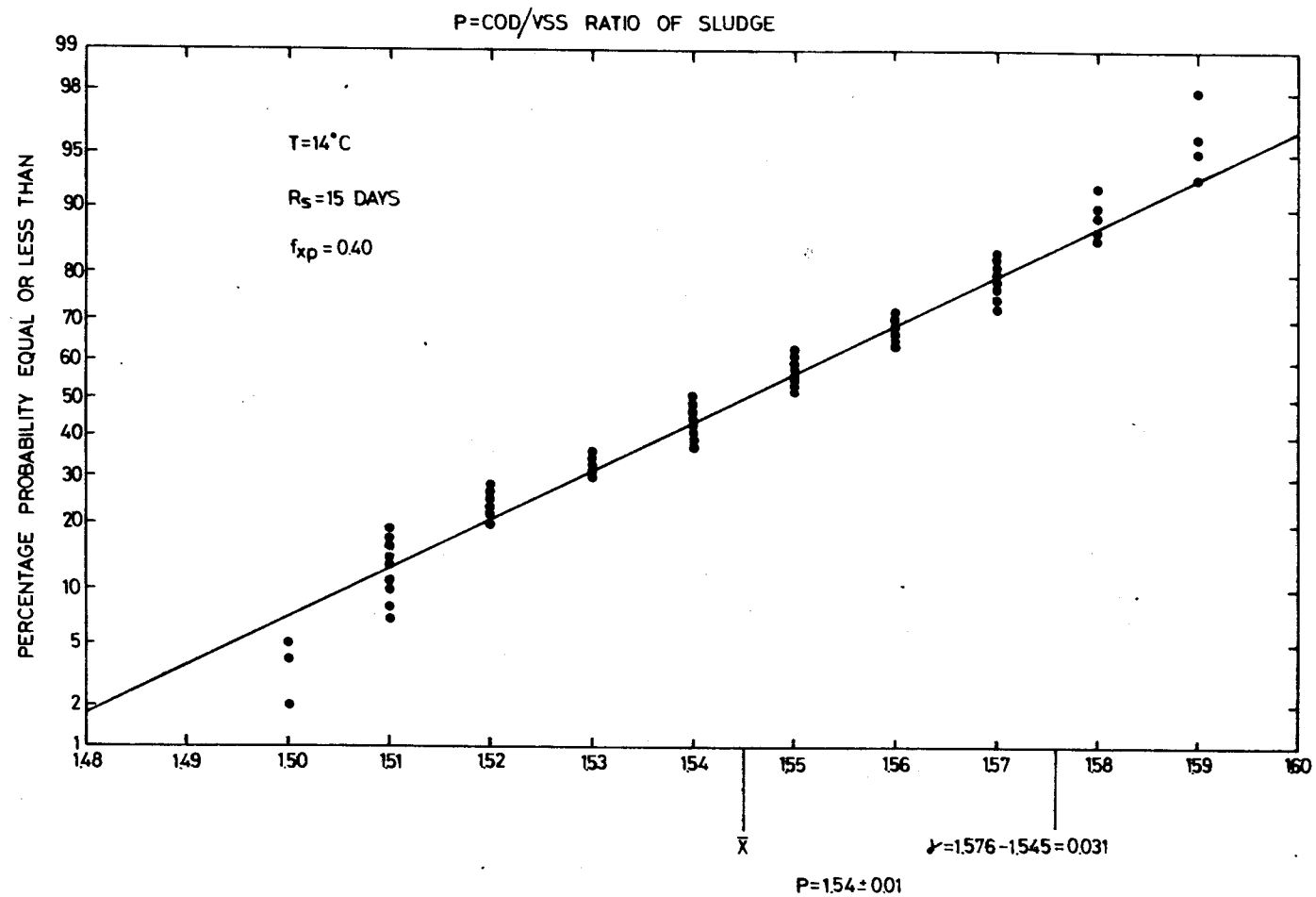


Fig.2.10: Mean value for P (COD content of the sludge) from a statistical analysis.

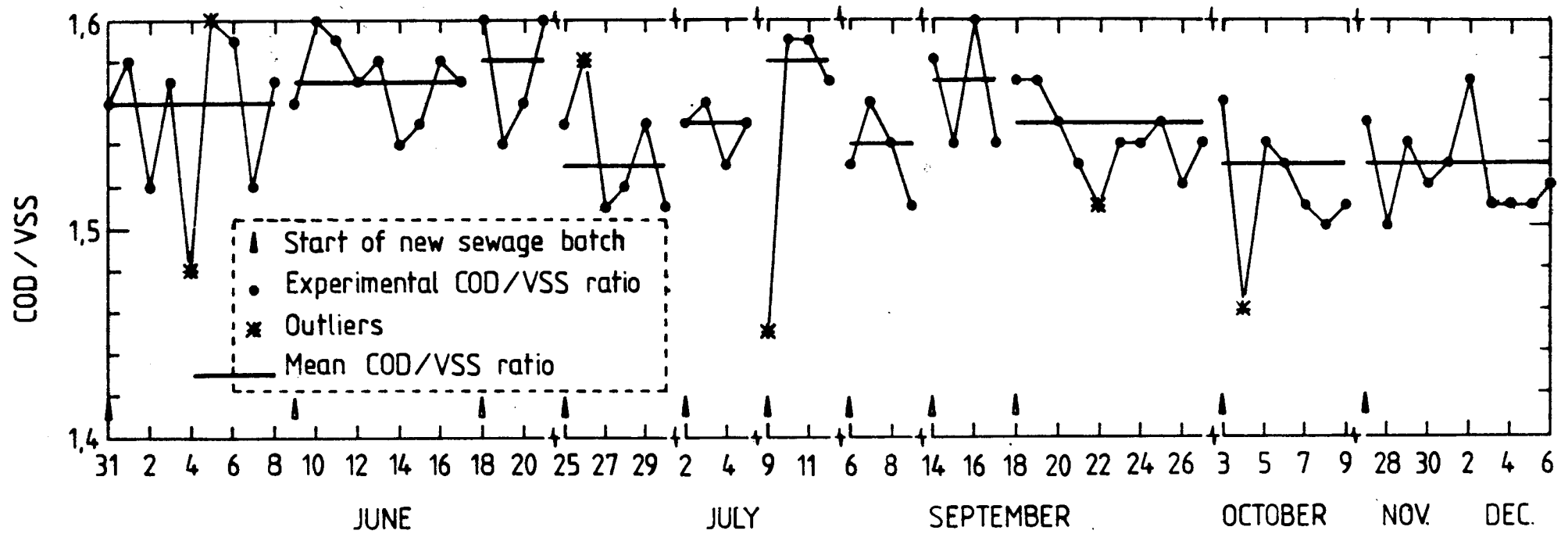


Fig.2.11 Variations in the experimentally measured COD/VSS ratio for different batches of Zeekoevlei sewage outfall at 14°C, pH = 7,2, $R_s = 15$ days.

APPENDIX C

C.1 A critical review of the oxygen uptake rate test

C.2 Experimental determination of oxygen utilization by the organisms (r_u).

C.1 A critical review of the oxygen uptake rate test

Successful application of oxygen uptake rate measurements depends on the ability to identify and quantify any errors inherent to the test.

The observed oxygen uptake rate $[(dD/dt)_{obs}]$ is the algebraic sum of (1) the rate of oxygen utilization by the organisms $[(dD/dt)_u]$, (2) the net rate of change in the oxygen concentration due to the dissolved oxygen concentration gradient (the difference in the dissolved oxygen (DO) concentrations of the influent to the reactor and the effluent from the reactor respectively), $[(dD/dt)_{IE}]$, and (3) the reaeration rate $[(dD/dt)_r]$, i.e.

$$(dD/dt)_{obs} = (dD/dt)_u + (dD/dt)_{IE} + (dD/dt)_r \quad (a)$$

C.1a Effect of (DO) concentration gradient

If the mixed liquor entering the reactor has a DO concentration different from that leaving the reactor (as will, usually, be the case), then there will be a net change in the DO concentration due to this difference. This can be expressed as

$$r_{IE} = -(dD/dt)_{IE} = [Q_a \cdot (D_i - D) / (24 V)] \quad (b)$$

where

D = DO concentration in the reactor $[mg\ O_2 \cdot l^{-1}]$

D_i = DO concentration in the influent mixed liquor to the reactor $[mg\ O_2 \cdot l^{-1}]$

Q_a = Actual flow rate (including any recycles) to the reactor $[l \cdot d^{-1}]$

$r_{IE} = -(dD/dt)_{IE}$ = Change in oxygen uptake rate due to the difference in DO concentrations of the influent to the reactor and the effluent from the reactor, respectively. (Negative sign indicates disappearance of oxygen). $(\Delta [mg\ O_2 \cdot l^{-1} \cdot h^{-1}])$

C.1b Effect of reaeration

Because the mixed liquor is not saturated with respect to DO, there is a certain rate of reaeration, r_r , which depends upon

- (1) The free surface/volume ratio.
- (2) The renewal rate of the free surface, i.e. the overall mixing intensity. The overall mixing intensity is mainly governed by the intensity of stirring and shear stress. The shear stress is the 'drag' effect caused by the introduction of influent to the mixed liquor in the reactor, i.e.

$$r_r = (dD/dt)_r = K_a \cdot (A_s/V) + r_t \quad (c)$$

where

K_a = Transfer coefficient [$\text{mg O}_2 \cdot \text{h}^{-1} \cdot \text{m}^{-2}$]

A_s = Free surface area [m^2]

V = Reactor volume [ℓ]

r_t = Reaeration via the opening in the U-tube preceding the reactor under consideration [$\text{mg O}_2 \cdot \ell^{-1} \cdot \text{h}^{-1}$].

It must be noted that r_r is specific to a certain reactor under certain conditions of flow and mixing and must therefore be determined experimentally for each particular case. Also the value of K_a need not be directly applicable for scale up purposes and is specific to the particular influent waste under consideration. Hence r_r is measured by substituting tap water for mixed liquor in the specific reactor (with its specific mixer), purging the water with nitrogen and thereafter monitoring the increase in DO concentration with time (rate of reaeration), keeping all flow streams operational at their respective experimental values.

In most practical cases r_r is usually so small that it has very little effect, but in bench scale units the ratio A_s/V sometimes is so high that the effect of reaeration becomes significant, especially when the oxygen uptake rate is low and the reactor is intensely stirred.

C.2 Determination of r_u

Substituting into Eq. (a) for $(dD/dt)_{IE}$ and $(dD/dt)_r$ from Eqs. (b) and (c) respectively, the observed rate of change of DO concentration, $r_{obs} = -(dD/dt)_{obs}$, can be written:

$$-(dD/dt)_{obs} = r_{obs} - r_u + r_{IE} - r_r = -(dD/dt)_v - (dD/dt)_{IE} - (dD/dt)_r$$

hence,

$$r_u = r_{obs} + (dD/dt)_{IE} + (dD/dt)_r$$

or

$$r_u = r_{obs} - [Q_a(D_i - D)/(24.V)] + r_r \quad (d)$$

The rate of oxygen utilization by the organisms is estimated by adjusting the observed oxygen uptake rate according to Eq. (d).

The value of r_u is now evaluated in turn for (I) the predenitrification system given in Fig. 3.1, and (II) for the completely mixed single reactor process given in Fig. 3.3.

(I) Considering the system in Fig. 3.1, the reaeration rates r_{rx} for any reactor x have been measured experimentally at 14°C as described in Section C.1(b) and were found to be:

$$r_{rA1} = 7,5 \text{ [mg O.}\ell^{-1}.\text{h}^{-1}\text{]}$$

$$r_{rA2} = 2,5 \text{ [mg O.}\ell^{-1}.\text{h}^{-1}\text{]}$$

(subscripts A1 and A2 refer to aerobic reactors A1 and A2, respectively).

$$\text{i.e. } r_r = \Sigma r_{rx} = 7,5 + 2,5 = 10,0 \text{ [mg O.}\ell^{-1}.\text{h}^{-1}\text{]}$$

For the system in Fig. 3.1 with

$$V_{A1} = 2,5 \text{ [}\ell\text{]}$$

$$V_{A2} = 4,5 \text{ [}\ell\text{]}$$

$$(a+s+1) = 7,$$

and assuming an average reactor DO concentration during the oxygen utilization rate test to be $D_o = 4 \text{ [mg O.}\ell^{-1}\text{]}$ (this seems a fair assumption considering that the oxygen uptake rate test is carried out between an upper DO concentration level of about $6 \text{ [mg O.}\ell^{-1}\text{]}$ and a lower DO concentration level of about $2 \text{ [mg O.}\ell^{-1}\text{]}$),

$$r_{IE_{A1}} = -[Q_a(D_o - D_i)/(24.V)]_{A1} = -17,2.7.(4-0)/(24.2,5) = -8,0 [\text{mg O.}\ell^{-1}.\text{h}^{-1}]$$

$$r_{IE_{A2}} = -[Q_a(D_o - D_i)/(24.V)]_{A2} = -17,2.7.(4-2)/(24.4,5) = -2,2 [\text{mg O.}\ell^{-1}.\text{h}^{-1}]$$

i.e.

$$r_{IE} = \Sigma r_{IE_x} = -8,0 + (-2,2) = -10,2 [\text{mg O.}\ell^{-1}.\text{h}^{-1}]$$

If r_u is calculated by using Eq.(d), it becomes apparent that for the system in Fig. 3.1, r_{IE} and r_r tend to cancel each other out. It therefore becomes unnecessary to consider these effects and hence they are ignored.

(II) Considering the single reactor system in Fig.3.3(that was sequentially subjected to anoxic and aerobic conditions), i.e.

$$V = 15 [\ell]$$

$$Q_a = Q.(s+1) = 30.(2+1) = 90 [\ell.d^{-1}].$$

Experiment (as described in Section C.1b) gives

$$r_r \approx 0,2 [\text{mg O.}\ell^{-1}.\text{h}^{-1}].$$

In view of the very small magnitude of r_r , the effect of the reaeration rate on r_u is ignored in the completely mixed single reactor (CMSR) system.

It is quite meaningless to attempt an accurate evaluation of the reactor's DO concentration and the reactor's inlet DO concentration during the oxygen utilization test. The main reason for that is the continuous underflow recycle (from the settler) with its varying DO concentration, directly into the reactor during the oxygen utilization rate (O.U.R.) test. A very rough indication of the magnitude of r_{IE} can be given if an average reactor DO concentration during the O.U.R. test is accepted as $D_o \approx 4 [\text{mg O.}\ell^{-1}]$ and an average underflow recycle DO concentration is assumed to be $2 [\text{mg O.}\ell^{-1}]$, in which event r_{IE} will be given by

$$r_{IE} = -[Q_a(D_o - D_i)/(24.V)] \approx -[90.(4-2)/(24.15)] = -0,5 [\text{mg O.}\ell^{-1}].$$

The uncertainty associated with the estimation of r_{IE} makes it difficult to accurately determine the value of r_u when using Eq.(d). Fortunately the magnitude of r_{IE} is very small and is not too far

away from the magnitude of r_r , except that it affects r_u in the opposite fashion to the way r_r affects r_u . Hence it becomes unnecessary to consider the effects of r_r and r_{IE} in the CMSR system and they are consequently ignored.